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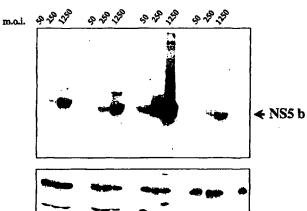
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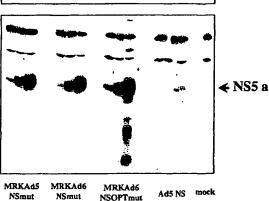
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(54) Title: HEPATITIS C VIRUS VACCINE



(57) Abstract: The present invention features Ad6 vectors and a nucleic acid encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide containing an inactive NS5B RNA-dependent RNA polymerase region. The nucleic acid is particularly useful as a component of an adenovector or DNA plasmid vaccine providing a broad range of antigens for generating an HCV specific cell mediated immune (CMI) response against HCV.





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TITLE OF THE INVENTION HEPATITIS C VIRUS VACCINE

RELATED APPLICATIONS

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The present application claims priority to provisional applications U.S. Serial No. 60/363,774, filed March 13, 2002, and U.S. Serial No. 60/328,655, filed October 11, 2001, each of which are hereby incorporated by reference herein.

BACKGROUND OF THE INVENTION

The references cited in the present application are not admitted to be prior art to the claimed invention.

About 3% of the world's population are infected with the Hepatitis C virus (HCV). (Wasley et al., Semin. Liver Dis. 20, 1-16, 2000.) Exposure to HCV results in an overt acute disease in a small percentage of cases, while in most instances the virus establishes a chronic infection causing liver inflammation and slowly progresses into liver failure and cirrhosis. (Iwarson, FEMS Microbiol. Rev. 14, 201-204, 1994.) In addition, epidemiological surveys indicate an important role of HCV in the pathogenesis of hepatocellular carcinoma. (Kew, FEMS Microbiol. Rev. 14, 211-220, 1994, Alter, Blood 85, 1681-1695, 1995.)

Prior to the implementation of routine blood screening for HCV in 1992, most infections were contracted by inadvertent exposure to contaminated blood, blood products or transplanted organs. In those areas where blood screening of HCV is carried out, HCV is primarily contracted through direct percutaneous exposure to infected blood, *i.e.*, intravenous drug use. Less frequent methods of transmission include perinatal exposure, hemodialysis, and sexual contact with an HCV infected person. (Alter *et al.*, *N. Engl. J. Med.* 341(8), 556-562, 1999, Alter, *J. Hepatol.* 31 Suppl. 88-91, 1999. Semin. Liver. Dis. 201, 1-16, 2000.)

The HCV genome consists of a single strand RNA about 9.5 kb encoding a precursor polyprotein of about 3000 amino acids. (Choo et al., Science 244, 362-364, 1989, Choo et al., Science 244, 359-362, 1989, Takamizawa et al., J. Virol. 65, 1105-1113, 1991.) The HCV polyprotein contains the viral proteins in the order: C-E1-E2-p7-NS2-NS3-NS4A-NS4B-NS5A-NS5B.

Individual viral proteins are produced by proteolysis of the HCV polyprotein. Host cell proteases release the putative structural proteins C, E1, E2, and

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p7, and create the N-terminus of NS2 at amino acid 810. (Mizushima et al., J. Virol. 68, 2731-2734, 1994, Hijikata et al., P.N.A.S. USA 90, 10773-10777, 1993.)

The non-structural proteins NS3, NS4A, NS4B, NS5A and NS5B presumably form the virus replication machinery and are released from the polyprotein. A zinc-dependent protease associated with NS2 and the N-terminus of NS3 is responsible for cleavage between NS2 and NS3. (Grakoui et al., J. Virol. 67, 1385-1395, 1993, Hijikata et al., P.N.A.S. USA 90, 10773-10777, 1993.) A distinct serine protease located in the N-terminal domain of NS3 is responsible for proteolytic cleavages at the NS3/NS4A, NS4A/NS4B, NS4B/NS5A and NS5A/NS5B junctions. (Bartenschlager et al., J. Virol. 67, 3835-3844, 1993, Grakoui et al., Proc. Natl. Acad. 10 Sci. USA 90, 10583-10587, 1993, Tomei et al., J. Virol. 67, 4017-4026, 1993.) NS4A provides a cofactor for NS3 activity. (Failla et al., J. Virol. 68, 3753-3760, 1994, De Francesco et al., U.S. Patent No. 5,739,002.)

NS5A is a highly phosphorylated protein conferring interferon resistance. (De Francesco et al., Semin. Liver Dis., 20(1), 69-83, 2000, Pawlotsky, Viral Hepat. Suppl. 1, 47-48, 1999.)

NS5B provides an RNA-dependent RNA polymerase. (De Francesco et al., International Publication Number WO 96/37619, Behrens et al., EMBO 15, 12-22, 1996, Lohmann et al., Virology 249, 108-118, 1998.)

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SUMMARY OF THE INVENTION

The present invention features Ad6 vectors and a nucleic acid encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide containing an inactive NS5B RNA-dependent RNA polymerase region. The nucleic acid is particularly useful as a component of an adenovector or DNA plasmid vaccine providing a broad range of antigens for generating an HCV specific cell mediated immune (CMI) response against HCV.

A HCV specific CMI response refers to the production of cytotoxic T lymphocytes and T helper cells that recognize an HCV antigen. The CMI response may also include non-HCV specific immune effects.

Preferred nucleic acids encode a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide that is substantially similar to SEQ. ID. NO. 1 and has sufficient protease activity to process itself to produce at least a polypeptide substantially similar to the NS5B region present in SEQ. ID. NO. 1. The produced polypeptide corresponding to NS5B is enzymatically inactive. More preferably, the HCV polypeptide has sufficient

protease activity to produce polypeptides substantially similar to the NS3, NS4A, NS4B, NS5A, and NS5B regions present in SEQ. ID. NO. 1.

Reference to a "substantially similar sequence" indicates an identity of at least about 65% to a reference sequence. Thus, for example, polypeptides having an amino acid sequence substantially similar to SEQ. ID. NO. 1 have an overall amino acid identity of at least about 65% to SEO. ID. NO. 1.

Polypeptides corresponding to NS3, NS4A, NS4B, NS5A, and NS5B have an amino acid sequence identity of at least about 65% to the corresponding region in SEQ. ID. NO. 1. Such corresponding polypeptides are also referred to herein as NS3, NS4A, NS4B, NS5A, and NS5B polypeptides.

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Thus, a first aspect of the present invention describes a nucleic acid comprising a nucleotide sequence encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide substantially similar to SEQ. ID. NO. 1. The encoded polypeptide has sufficient protease activity to process itself to produce an NS5B polypeptide that is enzymatically inactive.

In a preferred embodiment, the nucleic acid is an expression vector capable of expressing the Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide in a desired human cell. Expression inside a human cell has therapeutic applications for actively treating an HCV infection and for prophylactically treating against an HCV infection.

An expression vector contains a nucleotide sequence encoding a polypeptide along with regulatory elements for proper transcription and processing. The regulatory elements that may be present include those naturally associated with the nucleotide sequence encoding the polypeptide and exogenous regulatory elements not naturally associated with the nucleotide sequence. Exogenous regulatory elements such as an exogenous promoter can be useful for expression in a particular host, such as in a human cell. Examples of regulatory elements useful for functional expression include a promoter, a terminator, a ribosome binding site, and a polyadenylation signal.

Another aspect of the present invention describes a nucleic acid comprising a gene expression cassette able to express in a human cell a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide substantially similar to SEQ. ID. NO. 1. The polypeptide can process itself to produce an enzymatically inactive NS5B protein. The gene expression cassette contains at least the following:

a) a promoter transcriptionally coupled to a nucleotide sequence encoding a polypeptide;

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- b) a 5' ribosome binding site functionally coupled to the nucleotide sequence,
 - c) a terminator joined to the 3' end of the nucleotide sequence, and
- d) a 3' polyadenylation signal functionally coupled to the nucleotide sequence.

Reference to "transcriptionally coupled" indicates that the promoter is positioned such that transcription of the nucleotide sequence can be brought about by RNA polymerase binding at the promoter. Transcriptionally coupled does not require that the sequence being transcribed is adjacent to the promoter.

Reference to "functionally coupled" indicates the ability to mediate an effect on the nucleotide sequence. Functionally coupled does not require that the coupled sequences be adjacent to each other. A 3' polyadenylation signal functionally coupled to the nucleotide sequence facilitates cleavage and polyadenylation of the transcribed RNA. A 5' ribosome binding site functionally coupled to the nucleotide sequence facilitates ribosome binding.

In preferred embodiments the nucleic acid is a DNA plasmid vector or an adenovector suitable for either therapeutic application in treating HCV or as an intermediate in the production of a therapeutic vector. Treating HCV includes actively treating an HCV infection and prophylactically treating against an HCV infection.

Another aspect of the present invention describes an adenovector comprising a Met-NS3-NS4A-NS4B-NS5A-NS5B expression cassette able to express a polypeptide substantially similar to SEQ. ID. NO. 1 that is produced by a process involving (a) homologous recombination and (b) adenovector rescue. The homologous recombinant step produces an adenovirus genome plasmid. The adenovector rescue step produces the adenovector from the adenogenome plasmid.

Adenovirus genome plasmids described herein contain a recombinant adenovirus genome having a deletion in the E1 region and optionally in the E3 region and a gene expression cassette inserted into one of the deleted regions. The recombinant adenovirus genome is made of regions substantially similar to one or more adenovirus serotypes.

Another aspect of the present invention describes an adenovector consisting of the nucleic acid sequence of SEQ. ID. NO. 4 or a derivative thereof,

wherein said derivative thereof has the HCV polyprotein encoding sequence present in SEQ. ID. NO. 4 replaced with the HCV polyprotein encoding sequence of either SEQ. ID. NO. 3, SEQ. ID. NO. 10 or SEQ. ID. NO. 11.

Another aspect of the present invention describes a cultured recombinant cell comprising a nucleic acid containing a sequence encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide substantially similar to SEQ. ID. NO. 1. The recombinant cell has a variety of uses such as being used to replicate nucleic acid encoding the polypeptide in vector construction methods.

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Another aspect of the present invention describes a method of making an adenovector comprising a Met-NS3-NS4A-NS4B-NS5A-NS5B expression cassette able to express a polypeptide substantially similar to SEQ. ID. NO. 1. The method involves the steps of (a) producing an adenovirus genome plasmid containing a recombinant adenovirus genome with deletions in the E1 and E3 regions and a gene expression cassette inserted into one of the deleted regions and (b) rescuing the adenovector from the adenovirus genome plasmid.

Another aspect of the present invention describes a pharmaceutical composition comprising a vector for expressing a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide substantially similar to SEQ. ID. NO. 1 and a pharmaceutically acceptable carrier. The vector is suitable for administration and polypeptide expression in a patient.

A "patient" refers to a mammal capable of being infected with HCV. A patient may or may not be infected with HCV. Examples of patients are humans and chimpanzees.

Another aspect of the present invention describes a method of treating a patient comprising the step of administering to the patient an effective amount of a vector expressing a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide substantially similar to SEQ. ID. NO. 1. The vector is suitable for administration and polypeptide expression in the patient.

The patient undergoing treatment may or may not be infected with 30 HCV. For a patient infected with HCV, an effective amount is sufficient to achieve one or more of the following effects: reduce the ability of HCV to replicate, reduce HCV load, increase viral clearance, and increase one or more HCV specific CMI responses. For a patient not infected with HCV, an effective amount is sufficient to achieve one or more of the following: an increased ability to produce one or more components of a HCV specific CMI response to a HCV infection, a reduced

susceptibility to HCV infection, and a reduced ability of the infecting virus to establish persistent infection for chronic disease.

Another aspect of the present invention features a recombinant nucleic acid comprising an Ad6 region and a region not present in Ad6. Reference to "recombinant" nucleic acid indicates the presence of two or more nucleic acid regions not naturally associated with each other. Preferably, the Ad6 recombinant nucleic acid contains Ad6 regions and a gene expression cassette coding for a polypeptide heterologous to Ad6.

Other features and advantages of the present invention are apparent from the additional descriptions provided herein including the different examples. The provided examples illustrate different components and methodology useful in practicing the present invention. The examples do not limit the claimed invention. Based on the present disclosure the skilled artisan can identify and employ other components and methodology useful for practicing the present invention.

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BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A and 1B illustrate SEQ. ID. NO. 1.

Figures 2A, 2B, 2C, and 2D illustrate SEQ. ID. NO. 2. SEQ. ID. NO. 2 provides a nucleotide sequence coding for SEQ. ID. NO. 1 along with an optimized internal ribosome entry site and TAAA termination. Nucleotides 1-6 provides an optimized internal ribosome entry site. Nucleotides 7-5961 code for a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide with nucleotides in positions 5137 to 5145 providing a AlaAlaGly sequence in amino acid positions 1711 to 1713 that renders NS5B inactive. Nucleotides 5962-5965 provide a TAAA termination.

Figures 3A, 3B, 3C, and 3D illustrate SEQ. ID. NO. 3. SEQ. ID. NO. 3 is a codon optimized version of SEQ. ID. NO. 2. Nucleotides 7-5961 encode a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide.

Figures 4A-4M illustrate MRKAd6-NSmut (SEQ. ID. NO. 4). SEQ. ID. NO. 4 is an adenovector containing an expression cassette where the polypeptide of SEQ. ID. NO. 1 is encoded by SEQ. ID. NO. 2. Base pairs 1-450 correspond to the Ad5 bp 1 to 450; base pairs 462 to 1252 correspond to the human CMV promoter; base pairs 1258 to 1267 correspond to the Kozak sequence; base pairs 1264 to 7222 correspond to the NS genes; base pairs 7231 to 7451 correspond to the BGH polyadenylation signal; base pairs 7469 to 9506 correspond to Ad5 base pairs 3511 to 5548; base pairs 9507 to 32121 correspond to Ad6 base pairs 5542 to 28156; base

pairs 32122 to 35117 correspond to Ad6 base pairs 30789 to 33784; and base pairs 35118 to 37089 correspond to Ad5 base pairs 33967 to 35935.

Figures 5A-5O illustrate SEQ. ID. NOs. 5 and 6. SEQ. ID. NO. 5 encodes a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide with an active RNA dependent RNA polymerase. SEQ. ID. NO. 6 provides the amino acid sequence for the polypeptide.

Figures 6A-6C provide the nucleic acid sequence for pV1JnsA (SEQ. ID. NO. 7).

Figures 7A-7N provide the nucleic acid sequence for the Ad6 genome 10 (SEQ. ID. NO. 8).

Figures 8A-8K provide the nucleic acid sequence for the Ad5 genome (SEQ. ID. NO. 9).

Figure 9 illustrates different regions of the Ad6 genome. The linear (35759 bp) ds DNA genome is indicated by two parallel lines and is divided into 100 map units. Transcription units are shown relative to their position and orientation in the genome. Early genes (E1A, E1B, E2A/B, E3 and E4 are indicated by gray arrows. Late genes (L1 to L5), indicated by black arrows, are produced by alternative splicing of a transcript produced from the major late promoter (MLP) and all contain the tripartite leader (1, 2, 3) at their 5' ends. The E1 region is located from approximately 1.0 to 11.5 map units, the E2 region from 75.0 to 11.5 map units, E3 from 76.1 to 86.7 map units, and E4 from 99.5 to 91.2 map units. The major late transcription unit is located between 16.0 and 91.2 map units.

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Figure 10 illustrates homologous recombination to recover pAdE1-E3+ containing Ad6 and Ad5 regions.

Figure 11 illustrates homologous recombinant to recover a pAdE1-E3+ containing Ad6 regions.

Figure 12 illustrates a western blot on whole-cell extracts from 293 cells transfected with plasmid DNA expressing different HCV NS cassettes. Mature NS3 and NS5A products were detected with specific antibodies. "pV1Jns-NS" refers to a pV1JnsA plasmid where a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide is encoded by SEQ. ID. NO. 5, and SEQ. ID. NO. 5 is inserted between bases 1881 and 1912 of SEQ. ID. NO. 7. "pV1Jns-NSmut" refers to a pV1JnsA plasmid where SEQ. ID. NO. 2 is inserted between bases 1882 and 1925 of SEQ. ID. NO. 7. "pV1Jns-NSOPTmut" refers to a pV1JnsA plasmid where SEQ. ID. NO. 3 is inserted between bases 1881 and 1905 of SEQ. ID. NO. 7.

Figures 13A and 13B illustrate T cell responses by IFN γ ELIspot induced in C57black6 mice (A) and BalbC mice (B) by two injections of 25 μ g and 50 μ g, respectively, of plasmid DNA encoding the different HCV NS cassettes with Gene Electro-Transfer (GET).

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Figure 14 illustrates protein expression from different adenovectors upon infection of HeLa cells. MRKAd5-NSmut is an adenovector based on an Ad5 sequence (SEQ. ID. NO. 9), where the Ad5 genome has an E1 deletion of base pairs 451 to 3510, an E3 deletion of base pairs 28134 to 30817, and has the NS3-NS4A-NS4B-NS5A-NS5B expression cassette as provided in base pairs 451 to 7468 of SEQ. ID. NO. 4 inserted between positions 450 and 3511. Ad5-NS is an adenovector based on an Ad5 backbone with an E1 deletion of base pairs 342 to 3523, and E3 deletion of base pairs 28134 to 30817 and containing an expression cassette encoding a NS3-NS4A-NS4B-NS5A-NS5B from SEQ. ID. NO. 5. "MRKAd6-NSOPTmut" refers to an adenovector having a modified SEQ. ID. NO. 4 sequence, wherein base pairs 1258 to 7222 of SEQ. ID. NO. 4 is replaced with SEQ. ID. NO. 3.

Figure 15 illustrates T cell responses by IFN γ ELIspot induced in C57black6 mice by two injections of 10^9 vp of adenovectors containing different HCV non-structural gene cassettes.

Figures 16A-16D illustrate T cell responses by IFN γ ELIspot induced in Rhesus monkeys by one or two injections of 10^{10} vp (A) or 10^{11} vp (B) of adenovectors containing different HCV non-structural gene cassettes.

Figures 17A and 17B illustrates CD8+ T cell responses by IFN γ ICS induced in Rhesus monkeys by two injections of 10^{10} vp (A) or 10^{11} vp (B) of adenovectors encoding the different HCV non-structural gene cassettes.

Figures 18A-18F illustrate T cell responses by bulk CTL assay induced in Rhesus monkeys by two injections of 10¹¹ vp of Ad5-NS (A), MRKAd5-NSmut (B), or MRKAd6-NSmut (C).

Figure 19 illustrates the plasmid pE2.

Figures 20A-D illustrates the partial codon optimized sequence

NSsuboptmut (SEQ. ID. NO. 10). Coding sequence for the Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide is from base 7 to 5961.

DETAILED DESCRIPTION OF THE INVENTION

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The present invention features Ad6 vectors and nucleic acid encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide that contains an inactive NS5B region. Providing an inactive NS5B region supplies NS5B antigens while reducing the possibility of adverse side effects due to an active viral RNA polymerase. Uses of the featured nucleic acid include use as a vaccine component to introduce into a cell an HCV polypeptide that provides a broad range of antigens for generating a CMI response against HCV, and as an intermediate for producing such a vaccine component.

The adaptive cellular immune response can function to recognize viral antigens in HCV infected cells throughout the body due to the ubiquitous distribution of major histocompatibility complex (MHC) class I and II expression, to induce immunological memory, and to maintain immunological memory. These functions are attributed to antigen-specific CD4+ T helper (Th) and CD8+ cytotoxic T cells (CTL).

Upon activation via their specific T cell receptors, HCV specific Th cells fulfill a variety of immunoregulatory functions, most of them mediated by Th1 and Th2 cytokines. HCV specific Th cells assist in the activation and differentiation of B cells and induction and stimulation of virus-specific cytotoxic T cells. Together with CTL, Th cells may also secrete IFN-γ and TNF-α that inhibit replication and gene expression of several viruses. Additionally, Th cells and CTL, the main effector cells, can induce apoptosis and lysis of virus infected cells.

HCV specific CTL are generated from antigens processed by professional antigen presenting cells (pAPCs). Antigens can be either synthesized within or introduced into pAPCs. Antigen synthesis in a pAPC can be brought about by introducing into the cell an expression cassette encoding the antigen.

A preferred route of nucleic acid vaccine administration is an intramuscular route. Intramuscular administration appears to result in the introduction and expression of nucleic acid into somatic cells and pAPCs. HCV antigens produced in the somatic cells can be transferred to pAPCs for presentation in the context of MHC class I molecules. (Donnelly et al., Annu. Rev. Immunol. 15:617-648, 1997.)

pAPCs process longer length antigens into smaller peptide antigens in the proteasome complex. The antigen is translocated into the endoplasmic reticulum/Golgi complex secretory pathway for association with MHC class I

proteins. CD8+ T lymphocytes recognize antigen associated with class I MHC via the T cell receptor (TCR) and the CD8 cell surface protein.

Using a nucleic acid encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide as a vaccine component allows for production of a broad range of antigens capable of generating CMI responses from a single vector. The polypeptide should be able to process itself sufficiently to produce at least a region corresponding to NS5B. Preferred nucleic acids encode an amino acid sequence substantially similar to SEQ. ID. NO. 1 that has sufficient protease activity to process itself to produce individual HCV polypeptides substantially similar to the NS3, NS4A, NS4B, NS5A, and NS5B regions present in SEQ. ID. NO. 1.

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A polypeptide substantially similar to SEQ. ID. NO. 1 with sufficient protease activity to process itself in a cell provides the cell with T cell epitopes that are present in several different HCV strains. Protease activity is provided by NS3 and NS3/NS4A proteins digesting the Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide at the appropriate cleavage sites to release polypeptides corresponding to NS3, NS4A, NS4B, NS5A, and NS5B. Self- processing of the Met-NS3-NS4A-NS4B-NS5A-NS5B generates polypeptides that approximate naturally occurring HCV polypeptides.

Based on the guidance provided herein a sufficiently strong immune response can be generated to achieve beneficial effects in a patient. The provided guidance includes information concerning HCV sequence selection, vector selection, vector production, combination treatment, and administration.

I. HCV SEQUENCES

A variety of different nucleic acid sequences can be used as a vaccine component to supply a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide to a cell or as an intermediate to produce vaccine components. The starting point for obtaining suitable nucleic acid sequences are preferably naturally occurring NS3-NS4A-NS4B-NS5A-NS5B polypeptide sequences modified to produce an inactive NS5B.

The use of a HCV nucleic acid sequence providing HCV non-structural antigens to generate a CMI response is mentioned by Cho et al., Vaccine 17:1136-1144, 1999, Paliard et al., International Publication Number WO 01/30812 (not admitted to be prior art to the claimed invention), and Coit et al., International Publication Number WO 01/38360 (not admitted to be prior art to the claimed invention). Such references fail to describe, for example, a polypeptide that processes

itself to produce an inactive NS5B, and the particular combinations of HCV sequences and delivery vehicles employed herein.

Modifications to a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide sequence can be produced by altering the encoding nucleic acid. Alterations can be performed to create deletions, insertions and substitutions.

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Small modifications can be made in NS5B to produce an inactive polymerase by targeting motifs essentially for replication. Examples of motifs critical for NS5B activity and modifications that can be made to produce an inactive NS5B are described by Lohmann *et al.*, *Journal of Virology 71*:8416-8426, 1997, and Kolykhalov *et al.*, *Journal of Virology 74*:2046-2051, 2000.

Additional factors to take into account when producing modifications to a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide include maintaining the ability to self-process and maintaining T cell antigens. The ability of the HCV polypeptide to process itself is determined to a large extent by a functional NS3 protease. Modifications that maintain NS3 activity protease activity can be obtained by taking into account the NS3 protein, NS4A which serves as a cofactor for NS3, and NS3 protease recognition sites present within the NS3-NS4A-NS4B-NS5A-NS5B polypeptide.

Different modifications can be made to naturally occurring NS3-NS4A-NS4B-NS5A-NS5B polypeptide sequences to produce polypeptides able to elicit a broad range of T cell responses. Factors influencing the ability of a polypeptide to elicit a broad T cell response include the preservation or introduction of HCV specific T cell antigen regions and prevalence of different T cell antigen regions in different HCV isolates.

Numerous examples of naturally occurring HCV isolates are well known in the art. HCV isolates can be classified into the following six major genotypes comprising one or more subtypes: HCV-1/(1a,1b,1c), HCV-2/(2a,2b,2c), HCV-3/(3a,3b,10a), HCV-4/(4a), HCV-5/(5a) and HCV-6/(6a,6b,7b,8b,9a,11a). (Simmonds, J. Gen. Virol., 693-712, 2001.) Examples of particular HCV sequences such as HCV-BK, HCV-J, HCV-N, HCV-H, have been deposited in GenBank and described in various publications. (See, for example, Chamberlain et al., J. Gen. Virol., 1341-1347, 1997.)

HCV T cell antigens can be identified by, for example, empirical experimentation. One way of identifying T cell antigens involves generating a series of overlapping short peptides from a longer length polypeptide and then screening the

T-cell populations from infected patients for positive clones. Positive clones are activated/primed by a particular peptide. Techniques such as IFNY-ELISPOT, IFNY-Intracellular staining and bulk CTL assays can be used to measure peptide activity. Peptides thus identified can be considered to represent T-cell epitopes of the respective pathogen.

HCV T cell antigen regions from different HCV isolates can be introduced into a single sequence by, for example, producing a hybrid NS3-NS4A-NS4B-NS5A-NS5B polypeptide containing regions from two or more naturally occurring sequences. Such a hybrid can contain additional modifications, which preferably do not reduce the ability of the polypeptide to produce an HCV CMI response.

The ability of a modified Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide to process itself and produce a CMI response can be determined using techniques described herein or well known in the art. Such techniques include the use of IFNY-ELISPOT, IFNY-Intracellular staining and bulk CTL assays to measure a HCV specific CMI response.

A. Met-NS3-NS4A-NS4B-NS5A-NS5B Sequences

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SEQ. ID. NO. 1 provides a preferred Met-NS3-NS4A-NS4B-NS5A-NS5B sequence. SEQ. ID. NO. 1 contains a large number of HCV specific T cell antigens that are present in several different HCV isolates. SEQ. ID. NO. 1 is similar to the NS3-NS4A-NS4B-NS5A-NS5B portion of the HCV BK strain nucleotide sequence (GenBank accession number M58335).

In SEQ. ID. NO. 1 anchor positions important for recognition by MHC class I molecules are conserved or represent conservative substitutions for 18 out of 20 known T-cell epitopes in the NS3-NS4A-NS4B-NS5A-NS5B portion of HCV polyproteins. With respect to the remaining two known T-cell epitopes, one has a non-conservative anchor substitution in SEQ. ID. NO. 1 that may still be recognized by a different HLA supertype and one epitope has one anchor residue not conserved. HCV T-cell epitopes are described in Chisari et al., Curr. Top. Microbiol Immunol., 242:299-325, 2000, and Lechner et al. J. Exp. Med. 9:1499-1512, 2000.

Differences between the HCV-BK NS3-NS4A-NS4B-NS5A-NS5B nucleotide sequence and SEQ. ID. NO. 1 include the introduction of a methionine at the 5' end and the presence of modified NS5B active site residues in SEQ. ID. NO. 1.

The modification replaces GlyAspAsp with AlaAlaGly (residues 1711-1713) to inactivate NS5B.

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The encoded HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide preferably has an amino acid sequence substantially similar to SEQ. ID. NO. 1. In different embodiments, the encoded HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide has an amino acid identify to SEQ. ID. NO. 1 of at least 65%, at least 75%, at least 95%, at least 99% or 100%; or differs from SEQ. ID. NO. 1 by 1-2, 1-3, 1-4, 1-5, 1-6, 1-7, 1-8, 1-9, 1-10, 1-11, 1-12, 1-13, 1-14, 1-15, 1-16, 1-17, 1-18, 1-19, or 1-20 amino acids.

Amino acid differences between a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide and SEQ. ID. NO. 1 are calculated by determining the minimum number of amino acid modifications in which the two sequences differ. Amino acid modifications can be deletions, additions, substitutions or any combination thereof.

Amino acid sequence identity is determined by methods well known in the art that compare the amino acid sequence of one polypeptide to the amino acid sequence of a second polypeptide and generate a sequence alignment. Amino acid identity is calculated from the alignment by counting the number of aligned residue pairs that have identical amino acids.

Methods for determining sequence identity include those described by

20 Schuler, G.D. in Bioinformatics: A Practical Guide to the Analysis of Genes and
Proteins, Baxevanis, A.D. and Ouelette, B.F.F., eds., John Wiley & Sons, Inc, 2001;
Yona, et al., in Bioinformatics: Sequence, structure and databanks, Higgins, D. and
Taylor, W. eds, Oxford University Press, 2000; and Bioinformatics: Sequence and
Genome Analysis, Mount, D.W., ed., Cold Spring Harbor Laboratory Press, 2001).

Methods to determine amino acid sequence identity are codified in publicly available
computer programs such as GAP (Wisconsin Package Version 10.2, Genetics
Computer Group (GCG), Madison, Wisc.), BLAST (Altschul et al., J. Mol. Biol.
215(3):403-10, 1990), and FASTA (Pearson, Methods in Enzymology 183:63-98,
1990, R.F. Doolittle, ed.).

In an embodiment of the present invention sequence identity between two polypeptides is determined using the GAP program (Wisconsin Package Version 10.2, Genetics Computer Group (GCG), Madison, Wisc.). GAP uses the alignment method of Needleman and Wunsch. (Needleman, et al., J. Mol. Biol. 48:443-453, 1970.) GAP considers all possible alignments and gap positions between two sequences and creates a global alignment that maximizes the number of matched

residues and minimizes the number and size of gaps. A scoring matrix is used to assign values for symbol matches. In addition, a gap creation penalty and a gap extension penalty are required to limit the insertion of gaps into the alignment. Default program parameters for polypeptide comparisons using GAP are the BLOSUM62 (Henikoff *et al.*, *Proc. Natl. Acad. Sci. USA*, 89:10915-10919, 1992) amino acid scoring matrix (MATrix=blosum62.cmp), a gap creation parameter (GAPweight=8) and a gap extension pararameter (LENgthweight=2).

More preferred HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptides in addition to being substantially similar to SEQ. ID. NO. 1 across their entire length produce individual NS3, NS4A, NS4B, NS5A and NS5B regions that are substantially similar to the corresponding regions present in SEQ. ID. NO. 1. The corresponding regions in SEQ. ID. NO. 1 are provided as follows: Met-NS3 amino acids 1-632; NS4A amino acids 633-686; NS4B amino acids 687-947; NS5A amino acids 948-1394; and NS5B amino acids 1395-1985.

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In different embodiments a NS3, NS4A, NS4B, NS5A and/or NS5B region has an amino acid identity to the corresponding region in SEQ. ID. NO. 1 of at least 65%, at least 75%, at least 85%, at least 95%, at least 99%, or 100%; or an amino acid difference of 1-2, 1-3, 1-4, 1-5, 1-6, 1-7, 1-8, 1-9, 1-10, 1-11, 1-12, 1-13, 1-14, 1-15, 1-16, 1-17, 1-18, 1-19, or 1-20 amino acids.

Amino acid modifications to SEQ. ID. NO. 1 preferably maintain all or most of the T-cell antigen regions. Differences in naturally occurring amino acids are due to different amino acid side chains (R groups). An R group affects different properties of the amino acid such as physical size, charge, and hydrophobicity. Amino acids can be divided into different groups as follows: neutral and hydrophobic (alanine, valine, leucine, isoleucine, proline, tyrptophan, phenylalanine, and methionine); neutral and polar (glycine, serine, threonine, tryosine, cysteine, asparagine, and glutamine); basic (lysine, arginine, and histidine); and acidic (aspartic acid and glutamic acid).

Generally, in substituting different amino acids it is preferable to exchange amino acids having similar properties. Substituting different amino acids within a particular group, such as substituting valine for leucine, arginine for lysine, and asparagine for glutamine are good candidates for not causing a change in polypeptide tertiary structure.

Starting with a particular amino acid sequence and the known degeneracy of the genetic code, a large number of different encoding nucleic acid

sequences can be obtained. The degeneracy of the genetic code arises because almost all amino acids are encoded by different combinations of nucleotide triplets or "codons". The translation of a particular codon into a particular amino acid is well known in the art (see, e.g., Lewin GENES IV, p. 119, Oxford University Press, 1990).

5 Amino acids are encoded by codons as follows:

A=Ala=Alanine: codons GCA, GCC, GCG, GCU

C=Cys=Cysteine: codons UGC, UGU

D=Asp=Aspartic acid: codons GAC, GAU

E=Glu=Glutamic acid: codons GAA, GAG

10 F=Phe=Phenylalanine: codons UUC, UUU

G=Gly=Glycine: codons GGA, GGC, GGG, GGU

H=His=Histidine: codons CAC, CAU

I=Ile=Isoleucine: codons AUA, AUC, AUU

K=Lys=Lysine: codons AAA, AAG

15 L=Leu=Leucine: codons UUA, UUG, CUA, CUC, CUG, CUU

M=Met=Methionine: codon AUG

N=Asn=Asparagine: codons AAC, AAU

P=Pro=Proline: codons CCA, CCC, CCG, CCU

Q=Gln=Glutamine: codons CAA, CAG

20 R=Arg=Arginine: codons AGA, AGG, CGA, CGC, CGG, CGU

S=Ser=Serine: codons AGC, AGU, UCA, UCC, UCG, UCU

T=Thr=Threonine: codons ACA, ACC, ACG, ACU

V=Val=Valine: codons GUA, GUC, GUG, GUU

W=Trp=Tryptophan: codon UGG

25 Y=Tyr=Tyrosine: codons UAC, UAU.

Nucleic acid sequences can be optimized in an effort to enhance expression in a host. Factors to be considered include C:G content, preferred codons, and the avoidance of inhibitory secondary structure. These factors can be combined in different ways in an attempt to obtain nucleic acid sequences having enhanced

30 expression in a particular host. (See, for example, Donnelly *et al.*, International Publication Number WO 97/47358.)

The ability of a particular sequence to have enhanced expression in a particular host involves some empirical experimentation. Such experimentation involves measuring expression of a prospective nucleic acid sequence and, if needed,

35 altering the sequence.

B. Encoding Nucleotide Sequences

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SEQ. ID. NOs. 2 and 3 provide two examples of nucleotide sequences encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B sequence. The coding sequence of SEQ. ID. NO. 2 is similar (99.4% nucleotide sequence identity) to the NS3-NS4A-NS4B-NS5A-NS5B region of the naturally occurring HCV-BK sequence (GenBank accession number M58335). SEQ. ID. NO. 3 is a codon-optimized version of SEQ. ID. NO. 2. SEQ. ID. NOs. 2 and 3 have a nucleotide sequence identity of 78.3%.

Differences between the HCV-BK NS3-NS4A-NS4B-NS5A-NS5B nucleotide (GenBank accession number M58335) and SEQ. ID. NO. 2, include SEQ. ID. NO. 2 having a ribosome binding site, an ATG methionine codon, a region coding for a modified NS5B catalytic domain, a TAAA stop signal and an additional 30 nucleotide differences. The modified catalytic domain codes for a AlaAlaGly (residues 1711-1713) instead of GlyAspAsp to inactivate NS5B.

A nucleotide sequence encoding a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide is preferably substantially similar to the SEQ. ID. NO. 2 coding region. In different embodiments, the nucleotide sequence encoding a HCV Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide has a nucleotide sequence identify to the SEQ. ID. NO. 2 coding region of at least 65%, at least 75%, at least 85%, at least 95%, at least 99%, or 100%; or differs from SEQ. ID. NO. 2 by 1-2, 1-3, 1-4, 1-5, 1-6, 1-7, 1-8, 1-9, 1-10, 1-11, 1-12, 1-13, 1-14, 1-15, 1-16, 1-17, 1-18, 1-19, 1-20, 1-25, 1-30, 1-35, 1-40, 1-45, or 1-50 nucleotides.

Nucleotide differences between a sequence coding Met-NS3-NS4A-NS4B-NS5A-NS5B and the SEQ. ID. NO. 2 coding region are calculated by determining the minimum number of nucleotide modifications in which the two sequences differ. Nucleotide modifications can be deletions, additions, substitutions or any combination thereof.

Nucleotide sequence identity is determined by methods well known in the art that compare the nucleotide sequence of one sequence to the nucleotide sequence of a second sequence and generate a sequence alignment. Sequence identity is determined from the alignment by counting the number of aligned positions having identical nucleotides.

Methods for determining nucleotide sequence identity between two polynucleotides include those described by Schuler, in *Bioinformatics: A Practical Guide to the Analysis of Genes and Proteins*, Baxevanis, A.D. and Ouelette, B.F.F.,

eds., John Wiley & Sons, Inc, 2001; Yona et al., in Bioinformatics: Sequence, structure and databanks, Higgins, D. and Taylor, W. eds, Oxford University Press, 2000; and Bioinformatics: Sequence and Genome Analysis, Mount, D.W., ed., Cold Spring Harbor Laboratory Press, 2001). Methods to determine nucleotide sequence identity are codified in publicly available computer programs such as GAP (Wisconsin Package Version 10.2, Genetics Computer Group (GCG), Madison, Wisc.), BLAST (Altschul et al., J. Mol. Biol. 215(3):403-10, 1990), and FASTA (Pearson, W.R., Methods in Enzymology 183:63-98, 1990, R.F. Doolittle, ed.).

In an embodiment of the present ivnention, sequence identity between two polynucleotides is determined by application of GAP (Wisconsin Package Version 10.2, Genetics Computer Group (GCG), Madison, Wisc.). GAP uses the alignment method of Needleman and Wunsch. (Needleman et al., J. Mol. Biol. 48:443-453, 1970.) GAP considers all possible alignments and gap positions between two sequences and creates a global alignment that maximizes the number of matched residues and minimizes the number and size of gaps. A scoring matrix is used to assign values for symbol matches. In addition, a gap creation penalty and a gap extension penalty are required to limit the insertion of gaps into the alignment. Default program parameters for polynucleotide comparisons using GAP are the nwsgapdna.cmp scoring matrix (MATrix=nwsgapdna.cmp), a gap creation parameter (GAPweight=50) and a gap extension pararameter (LENgthweight=3).

More preferred HCV Met-NS3-NS4A-NS4B-NS5A-NS5B nucleotide sequences in addition to being substantially similar across its entire length, produce individual NS3, NS4A, NS4B, NS5A and NS5B regions that are substantially similar to the corresponding regions present in SEQ. ID. NO. 2. The corresponding coding regions in SEQ. ID. NO. 2 are provided as follows: Met-NS3, nucleotides 7-1902; NS4A nucleotides 1903-2064; NS4B nucleotides 2065-2847; NS5A nucleotides 2848-4188: NS5B nucleotides 4189-5661.

In different embodiments a NS3, NS4A, NS4B, NS5A and/or NS5B encoding region has a nucleotide sequence identity to the corresponding region in SEQ. ID. NO. 2 of at least 65%, at least 75%, at least 85%, at least 95%, at least 99% or 100%; or a nucleotide difference to SEQ. ID. NO. 2 of 1-2, 1-3, 1-4, 1-5, 1-6, 1-7, 1-8, 1-9, 1-10, 1-11, 1-12, 1-13, 1-14, 1-15, 1-16, 1-17, 1-18, 1-19, 1-20, 1-25, 1-30, 1-35, 1-40, 1-45, or 1-50 nucleotides.

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C. Gene Expression Cassettes

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A gene expression cassette contains elements needed for polypeptide expression. Reference to "polypeptide" does not provide a size limitation and includes protein. Regulatory elements present in a gene expression cassette generally include: (a) a promoter transcriptionally coupled to a nucleotide sequence encoding the polypeptide, (b) a 5' ribosome binding site functionally coupled to the nucleotide sequence, (c) a terminator joined to the 3' end of the nucleotide sequence, and (d) a 3' polyadenylation signal functionally coupled to the nucleotide sequence. Additional regulatory elements useful for enhancing or regulating gene expression or polypeptide processing may also be present.

Promoters are genetic elements that are recognized by an RNA polymerase and mediate transcription of downstream regions. Preferred promoters are strong promoters that provide for increased levels of transcription. Examples of strong promoters are the immediate early human cytomegalovirus promoter (CMV), and CMV with intron A. (Chapman *et al*, *Nucl. Acids Res.* 19:3979-3986, 1991.) Additional examples of promoters include naturally occurring promoters such as the EF1 alpha promoter, the murine CMV promoter, Rous sarcoma virus promoter, and SV40 early/late promoters and the β-actin promoter; and artificial promoters such as a synthetic muscle specific promoter and a chimeric muscle-specific/CMV promoter (Li *et al.*, *Nat. Biotechnol. 17*:241-245, 1999, Hagstrom *et al.*, *Blood 95*:2536-2542, 2000).

The ribosome binding site is located at or near the initiation codon. Examples of preferred ribosome binding sites include CCACCAUGG, CCGCCAUGG, and ACCAUGG, where AUG is the initiation codon. (Kozak, *Cell* 44:283-292, 1986). Another example of a ribosome binding site is GCCACCAUGG (SEQ. ID. NO. 12).

The polyadenylation signal is responsible for cleaving the transcribed RNA and the addition of a poly (A) tail to the RNA. The polyadenylation signal in higher eukaryotes contains an AAUAAA sequence about 11-30 nucleotides from the polyadenylation addition site. The AAUAAA sequence is involved in signaling RNA cleavage. (Lewin, Genes IV, Oxford University Press, NY, 1990.) The poly (A) tail is important for the mRNA processing.

Polyadenylation signals that can be used as part of a gene expression cassette include the minimal rabbit β -globin polyadenylation signal and the bovine growth hormone polyadenylation (BGH). (Xu et al., Gene 272:149-156, 2001, Post et

al., U.S. Patent U. S. 5,122,458.) Additional examples include the Synthetic Polyadenylation Signal (SPA) and SV40 polyadenylation signal. The SPA sequence is as follows: AAUAAAAGAUCUUUAUUUUCAUUAGAUCUGUGUGUUUUUUUGUGUG (SEQ. ID. NO. 13).

Examples of additional regulatory elements useful for enhancing or regulating gene expression or polypeptide processing that may be present include an enhancer, a leader sequence and an operator. An enhancer region increases transcription. Examples of enhancer regions include the CMV enhancer and the SV40 enhancer. (Hitt et al., Methods in Molecular Genetics 7:13-30, 1995, Xu, et al., Gene 272:149-156, 2001.) An enhancer region can be associated with a promoter.

A leader sequence is an amino acid region on a polypeptide that directs the polypeptide into the proteasome. Nucleic acid encoding the leader sequence is 5' of a structural gene and is transcribed along the structural gene. An example of a leader sequences is tPA.

An operator sequence can be used to regulate gene expression. For example, the Tet operator sequence can be used to repress gene expression.

II. THERAPEUTIC VECTORS

Nucleic acid encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide can be introduced into a patient using vectors suitable for therapeutic administration. Suitable vectors can deliver nucleic acid into a target cell without causing an unacceptable side effect.

Cellular expression is achieved using a gene expression cassette encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide. The gene expression cassette contains regulatory elements for producing and processing a sufficient amount of nucleic acid inside a target cell to achieve a beneficial effect.

Examples of vectors that can be used for therapeutic applications include first and second generation adenovectors, helper dependent adenovectors, adeno-associated viral vectors, retroviral vectors, alpha virus vectors, Venezuelan Equine Encephalitis virus vector, and plasmid vectors. (Hitt, et al., Advances in Pharmacology 40:137-206, 1997, Johnston et al., U.S. Patent No. 6,156,588, and Johnston et al., International Publication Number WO 95/32733.) Preferred vectors for introducing a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide into a subject are first generation adenoviral vectors and plasmid DNA vectors.

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A. First Generation Adenovectors

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First generation adenovector for expressing a gene expression cassette contain the expression cassette in an E1 and optionally E3 deleted recombinant adenovirus genome. The deletion in the E1 region is sufficiently large to remove elements needed for adenoviral replication.

First generation adenovectors for expressing a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide contain a E1 and E3 deleted recombinant adenovirus genome. The deletion in the E1 region is sufficiently large to remove elements needed for adenoviral replication. The combinations of deletions of the E1 and E3 regions are sufficiently large to accommodate a gene expression cassette encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide.

The adenovirus has a double-stranded linear genome with inverted terminal repeats at both ends. During viral replication, the genome is packaged inside a viral capsid to form a virion. The virus enters its target cell through viral attachment followed by internalization. (Hitt et al., Advances in Pharmacology 40:137-206, 1997.)

Adenovectors can be based on different adenovirus serotypes such as those found in humans or animals. Examples of animal adenoviruses include bovine, porcine, chimp, murine, canine, and avian (CELO). Preferred adenovectors are based on human serotypes, more preferably Group B, C, or D serotypes. Examples of human adenovirus Group B, C, D, or E serotypes include types 2 ("Ad2"), 4 ("Ad4"), 5 ("Ad5"), 6 ("Ad6"), 24 ("Ad24"), 26 ("Ad26"), 34 ("Ad34") and 35 ("Ad35"). Adenovectors can contain regions from a single adenovirus or from two or more adenovirus.

In different embodiments adenovectors are based on Ad5, Ad6, or a combination thereof. Ad5 is described by Chroboczek, et al., J. Virology 186:280-285, 1992. Ad6 is described in Figures 7A-7N. An Ad6 based vector containing Ad5 regions is described in the Example section provided below.

Adenovectors do not need to have their E1 and E3 regions completely removed. Rather, a sufficient amount the E1 region is removed to render the vector replication incompetent in the absence of the E1 proteins being supplied in *trans*; and the E1 deletion or the combination of the E1 and E3 deletions are sufficiently large enough to accommodate a gene expression cassette.

E1 deletions can be obtained starting at about base pair 342 going up to about base pair 3523 of Ad5, or a corresponding region from other adenoviruses.

Preferably, the deleted region involves removing a region from about base pair 450 to about base pair 3511 of Ad5, or a corresponding region from other adenoviruses. Larger E1 region deletions starting at about base pair 341 removes elements that facilitate virus packaging.

E3 deletions can be obtained starting at about base pair 27865 to about base pair 30995 of Ad5, or the corresponding region of other adenovectors.

Preferably the deletion region involves removing a region from about base pair 28134 up to about base pair 30817 of Ad5, or the corresponding region of other adenovectors.

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The combination of deletions to the E1 region and optionally the E3 region should be sufficiently large so that the overall size of the recombinant genome containing the gene expression cassette does not exceed about 105% of the wild type adenovirus genome. For example, as recombinant adenovirus Ad5 genomes increase size above about 105% the genome becomes unstable. (Bett et al., Journal of Virology 67:5911-5921, 1993.)

Preferably, the size of the recombinant adenovirus genome containing the gene expression cassette is about 85% to about 105% the size of the wild type adenovirus genome. In different embodiments, the size of the recombinant adenovirus genome containing the expression cassette is about 100% to about 105.2%, or about 100%, the size of the wild type genome.

Approximately 7,500 kb can be inserted into an adenovirus genome with a E1 and E3 deletion. Without any deletion, the Ad5 genome is 35,935 base pairs and the Ad6 genome is 35,759 base pairs.

Replication of first generation adenovectors can be performed by supplying the E1 gene products in *trans*. The E1 gene product can be supplied in *trans*, for example, by using cell lines that have been transformed with the adenovirus E1 region. Examples of cells and cells lines transformed with the adenovirus E1 region are HEK 293 cells, 911 cells, PERC.6TM cells, and transfected primary human aminocytes cells. (Graham *et al.*, *Journal of Virology 36*:59-72, 1977, Schiedner *et al.*, *Human Gene Therapy 11*:2105-2116, 2000, Fallaux *et al.*, *Human Gene Therapy* 9:1909-1917, 1998, Bout *et al.*, U.S. Patent No. 6,033,908.)

A Met-NS3-NS4A-NS4B-NS5A-NS5B expression cassette should be inserted into a recombinant adenovirus genome in the region corresponding to the deleted E1 region or the deleted E3 region. The expression cassette can have a parallel or anti-parallel orientation. In a parallel orientation the transcription direction

of the inserted gene is the same direction as the deleted E1 or E3 gene. In an antiparallel orientation transcription the opposite strand serves as a template and the transcription direction is in the opposite direction.

In an embodiment of the present invention the adenovector has a gene expression cassette inserted in the E1 deleted region. The vector contains:

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- a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;
- b) a gene expression cassette in a E1 parallel or E1 anti-parallel orientation joined to the first region;
- c) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to the expression cassette;
- d) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to the second region;
- e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to the third region; and
- f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6 joined to the fourth region.

In another embodiment of the present invention the adenovector has an expression cassette inserted in the E3 deleted region. The vector contains:

- a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;
 - b) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to the first region;
- c) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to the second region;
- d) a gene expression cassette in a E3 parallel or E3 anti-parallel orientation joined to the third region;

e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to the gene expression cassette; and

f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to the fourth region.

In preferred different embodiments concerning adenovirus regions that are present: (1) the first, second, third, fourth, and fifth region corresponds to Ad5; (2) the first, second, third, fourth, and fifth region corresponds to Ad6; and (3) the first region corresponds to Ad5, the second region corresponds to Ad5, the third region corresponds to Ad6, the fourth region corresponds to Ad6, and the fifth region corresponds to Ad5.

B. DNA Plasmid Vectors

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DNA vaccine plasmid vectors contain a gene expression cassette along with elements facilitating replication and preferably vector selection. Preferred elements provide for replication in non-mammalian cells and a selectable marker. The vectors should not contain elements providing for replication in human cells or for integration into human nucleic acid.

The selectable marker facilitates selection of nucleic acids containing the marker. Preferred selectable markers are those that confer antibiotic resistance. Examples of antibiotic selection genes include nucleic acid encoding resistance to ampicillin, neomycin, and kanamycin.

Suitable DNA vaccine vectors can be produced starting with a plasmid containing a bacterial origin of replication and a selectable marker. Examples of bacterial origins of replication providing for higher yields include the ColE1 plasmid-derived bacterial origin of replication. (Donnelly et al., Annu. Rev. Immunol. 15:617-648, 1997.)

The presence of the bacterial origin of replication and selectable marker allows for the production of the DNA vector in a bacterial strain such as *E. coli*. The selectable marker is used to eliminate bacteria not containing the DNA vector.

III. AD6 RECOMBINANT NUCLEIC ACID

Ad6 recombinant nucleic acid comprises an Ad6 region substantially similar to an Ad6 region found in SEQ. ID. NO. 8, and a region not present in Ad6 nucleic acid. Recombinant nucleic acid comprising Ad6 regions have different uses such as in producing different Ad6 regions, as intermediates in the production of Ad6 based vectors, and as a vector for delivering a recombinant gene.

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As depicted in Figure 9, the genomic organization of Ad6 is very similar to the genomic organization of Ad5. The homology between Ad5 and Ad6 is approximately 98%.

In different embodiments, the Ad6 recombinant nucleic acid comprises a nucleotide region substantially similar to E1A, E1B, E2B, E2A, E3, E4, L1, L2, L3, or L4, or any combination thereof. A substantially similar nucleic acid region to an Ad6 region has a nucleotide sequence identity of at least 65%, at least 75%, at least 85%, at least 95%, at least 99% or 100%; or a nucleotide difference of 1-2, 1-3, 1-4, 1-5, 1-6, 1-7, 1-8, 1-9, 1-10, 1-11, 1-12, 1-13, 1-14, 1-15, 1-16, 1-17, 1-18, 1-19, 1-20, 1-25, 1-30, 1-35, 1-40, 1-45, or 1-50 nucleotides. Techniques and embodiments for determining substantially similar nucleic acid sequences are described in Section I.B. supra.

Preferably, the recombinant Ad6 nucleic acid contains an expression cassette coding for a polypeptide not found in Ad6. Examples of expression cassettes include those coding for HCV regions and those coding for other types of polypeptides.

Different types of adenoviral vectors can be produced incorporating different amounts of Ad6, such as first and second generation adenovectors. As noted in Section II.A. *supra*. first generation adenovectors are defective in E1 and can replicate when E1 is supplied *in trans*.

Second generation adenovectors contain less adenoviral genome than first generation vectors and can be used in conjugation with complementing cell lines and/or helper vectors supplying adenoviral proteins. Second generation adenovectors are described in different references such as Russell, *Journal of General Virology* 81:2573-2604, 2000; Hitt et al., 1997, Human Ad vectors for Gene Transfer, Advances in Pharmacology, Vol 40 Academic Press.

In an embodiment of the present invention, the Ad6 recombinant nucleic acid is an adenovirus vector defective in E1 that is able to replicate when E1 is

supplied in trans. Expression cassettes can be inserted into a deleted E1 region and/or a deleted E3 region.

An example of an Ad6 based adenoviral vector with an expression cassette provided in a deleted E1 region comprises or consists of:

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- a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;
- b) a gene expression cassette in a E1 parallel or E1 anti-parallel orientation joined to the first region;
- c) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to the expression cassette;
- d) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to the second region;
- e) an optionally present fourth region from about base pair 28134 to about base pair 30817 corresponding to Ad5, or from about base pair 28157 to about base pair 30788 corresponding to Ad6, joined to the third region;
- f) a fifth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, wherein the fifth region is joined to the fourth region if the fourth region is present, or the fifth is joined to the third region if the fourth region is not present; and
- g) a sixth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to the fifth region;

wherein at least one Ad6 region is present.

In different embodiments of the invention, all of the regions are from Ad6; all of the regions expect for the first and second are from Ad6; and 1, 2, 3, or 4 regions selected from the second, third, fourth, and fifth regions are from Ad6.

An example of an Ad6 based adenoviral vector with an expression cassette provided in a deleted E3 region comprises or consists of:

a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;

b) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to the first region;

c) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to the second region;

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- d) a gene expression cassette in a E3 parallel or E3 anti-parallel orientation joined to the third region;
- e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to the gene expression cassette; and
- f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to the fourth region;

wherein at least one Ad6 region is present.

In different embodiment of the invention, all of the regions are from Ad6; all of the regions expect for the first and second are from Ad6; and 1, 2, 3, or 4 regions selected from the second, third, fourth and fifth regions are from Ad6.

IV. VECTOR PRODUCTION

Vectors can be produced using recombinant nucleic acid techniques such as those involving the use of restriction enzymes, nucleic acid ligation, and homologous recombination. Recombinant nucleic acid techniques are well known in the art. (Ausubel, Current Protocols in Molecular Biology, John Wiley, 1987-1998, and Sambrook et al., Molecular Cloning, A Laboratory Manual, 2nd Edition, Cold Spring Harbor Laboratory Press, 1989.)

Intermediate vectors are used to derive a therapeutic vector or to transfer an expression cassette or portion thereof from one vector to another vector. Examples of intermediate vectors include adenovirus genome plasmids and shuttle vectors.

Useful elements in an intermediate vector include an origin of replication, a selectable marker, homologous recombination regions, and convenient restriction sites. Convenient restriction sites can be used to facilitate cloning or release of a nucleic acid sequence.

Homologous recombination regions provide nucleic acid sequence regions that are homologous to a target region in another nucleic acid molecule. The homologous regions flank the nucleic acid sequence that is being inserted into the target region. In different embodiments homologous regions are preferably about 150 to 600 nucleotides in length, or about 100 to 500 nucleotides in length.

An embodiment of the present invention describes a shuttle vector containing a Met-NS3-NS4A-NS4B-NS5A-NS5B expression cassette, a selectable marker, a bacterial origin of replication, a first adenovirus homology region and a second adenovirus homologous region that target the expression cassette to insert in or replace an E1 region. The first and second homology regions flank the expression cassette. The first homology region contains at least about 100 base pairs substantially homologous to at least the right end (3' end) of a wild-type adenovirus region from about base pairs 4-450. The second homology contains at least about 100 base pairs substantially homologous to at least the left end (5' end) of Ad5 from about base pairs 3511-5792, or the corresponding region from another adenovirus.

Reference to "substantially homologous" indicates a sufficient degree of homology to specifically recombine with a target region. In different embodiments substantially homologous refers to at least 85%, at least 95%, or 100% sequence identity. Sequence identity can be calculated as described in Section I.B. supra.

One method of producing adenovectors is through the creation of an adenovirus genome plasmid containing an expression cassette. The pre-Adenovirus plasmid contains all the adenovirus sequences needed for replication in the desired complimenting cell line. The pre-Adenovirus plasmid is then digested with a restriction enzyme to release the viral ITR's and transfected into the complementing cell line for virus rescue. The ITR's must be released from plasmid sequences to allow replication to occur. Adenovector rescue results in the production on an adenovector containing the expression cassette.

A. Adenovirus Genome Plasmids

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Adenovirus genome plasmids contain an adenovector sequence inside a longer-length plasmid (which may be a cosmid). The longer-length plasmid may contain additional elements such as those facilitating growth and selection in eukaryotic or bacterial cells depending upon the procedures employed to produce and maintain the plasmid. Techniques for producing adenovirus genome plasmids include those involving the use of shuttle vectors and homologous recombination, and those

involving the insertion of a gene expression cassette into an adenovirus cosmid. (Hitt et al., Methods in Molecular Genetics 7:13-30, 1995, Danthinne et al., Gene Therapy 7:1707-1714, 2000.)

Adenovirus genome plasmids preferably have a gene expression cassette inserted into a E1 or E3 deleted region. In an embodiment of the present invention, the adenovirus genome plasmid contains a gene expression cassette inserted in the E1 deleted region, an origin of replication, a selectable marker, and the recombinant adenovirus region is made up of:

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- a) a first adenovirus region from about base pair 1 to about base
 450 corresponding to either Ad5 or Ad6;
 - b) a gene expression cassette in a E1 parallel or E1 anti-parallel orientation joined to the first region;
 - c) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to the expression cassette;
 - d) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to the second region;
 - e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to the third region;
 - f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to the fourth region, and
 - g) an optionally present E3 region corresponding to all or part of the E3 region present in Ad5 or Ad6, which may be present for smaller inserts taking into account the overall size of the desired adenovector.

In another embodiment of the present invention the recombinant adenovirus genome plasmid has the gene expression cassette inserted in the E3 deleted region. The vector contains an origin of replication, a selectable marker, and the following:

a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;

b) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to the expression cassette;

c) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to the second region;

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- d) the gene expression cassette in a E3 parallel or E3 anti-parallel orientation joined to the third region;
- e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to the gene expression cassette; and
- f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to the fourth region.

In different embodiments concerning adenovirus regions that are present: (1) the first, second, third, fourth, and fifth region corresponds to Ad5; (2) the first, second, third, fourth, and fifth region corresponds to Ad6; and (3) the first region corresponds to Ad5, the second region corresponds to Ad5, the third region corresponds to Ad6, the fourth region corresponds to Ad6, and the fifth region corresponds to Ad5.

An embodiment of the present invention describes a method of making an adenovector involving a homologous recombination step to produce a adenovirus genome plasmid and an adenovirus rescue step. The homologous recombination step involves the use of a shuttle vector containing a Met-NS3-NS4A-NS4B-NS5A-NS5B expression cassette flanked by adenovirus homology regions. The adenovirus homology regions target the expression cassette into either the E1 or E3 deleted region.

In an embodiment of the present invention concerning the production of an adenovirus genome plasmid, the gene expression cassette is inserted into a vector comprising: a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6; a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to the second region; a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6,

joined to the second region; a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to the third region; and a fifth adenovirus region from about 33967 to about 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to the fourth region. The adenovirus genome plasmid should contain an origin of replication and a selectable marker, and may contain all or part of the Ad5 or Ad6 E3 region.

In different embodiments concerning adenovirus regions that are present: (1) the first, second, third, fourth, and fifth region corresponds to Ad5; (2) the first, second, third, fourth, and fifth region corresponds to Ad6; and (3) the first region corresponds to Ad5, the second region corresponds to Ad5, the third region corresponds to Ad6, the fourth region corresponds to Ad6, and the fifth region corresponds to Ad5.

15 B. Adenovector Rescue

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An adenovector can be rescued from a recombinant adenovirus genome plasmid using techniques known in the art or described herein. Examples of techniques for adenovirus rescue well known in the art are provided by Hitt *et al.*, *Methods in Molecular Genetics* 7:13-30, 1995, and Danthinne *et al.*, *Gene Therapy* 7:1707-1714, 2000.

A preferred method of rescuing an adenovector described herein involves boosting adenoviral replication. Boosting adenoviral replication can be performed, for example, by supplying adenoviral functions such as E2 proteins (polymerase, pre-terminal protein and DNA binding protein) as well as E4 orf6 on a separate plasmid. Example 10 *infra*. illustrates the boosting of adenoviral replication to rescue an adenovector containing a codon optimized Met-NS3-NS4A-NS4B-NS5B expression cassette.

V. PARTIAL-OPITIMIZED HCV ENCODING SEQUENCES

Partial optimization of HCV polyprotein encoding nucleic acid provides for a lesser amount of codons optimized for expression in a human than complete optimization. The overall objective is to provide the benefits of increased expression due to codon optimization, while facilitating the production of an adenovector containing HCV polyprotein encoding nucleic acid having optimized codons.

Complete optimization of an HCV polyprotein encoding sequence provides the most frequently observed human codon for each amino acid. Complete optimization can be performed using codon frequency tables well known in the art and using programs such as the BACKTRANSLATE program (Wisconsin Package version 10, Genetics Computer Group, GCG, Madison, Wisc.).

Partial optimization can be preformed on an entire HCV polyprotein encoding sequence that is present (e.g., NS3-NS5B), or one or more local regions that are present. In different embodiments the GC content for the entire HCV encoded polyprotein that is present is no greater than at least about 65%; and the GC content for one or more local regions is no greater than about 70%.

Local regions are regions present in HCV encoding nucleic acid, and can vary in size. For example, local regions can be about 60, about 70, about 80, about 90 or about 100 nucleotides in length.

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Partial optimization can be achieved by initially constructing an HCV encoding polyprotein sequence to be partially optimized based on a naturally ocurring sequence. Alternatively, an optimized HCV encoding sequence can be used as basis of comparison to produce a partial optimized sequence.

VI. HCV COMBINATION TREATMENT

The HCV Met-NS3-NS4A-NS4B-NS5A-NS5B vaccine can be used by itself to treat a patient, can be used in conjunction with other HCV therapeutics, and can be used with agents targeting other types of diseases. Additional therapeutics include additional therapeutic agents to treat HCV and diseases having a high prevalence in HCV infected persons. Agents targeting other types of disease include vaccines directed against HIV and HBV.

Additional therapeutics for treating HCV include vaccines and non-vaccine agents. (Zein, Expert Opin. Investig. Drugs 10:1457-1469, 2001.) Examples of additional HCV vaccines include vaccines designed to elicit an immune response against an HCV core antigen and the HCV E1, E2 or p7 region. Vaccine components can be naturally occurring HCV polypeptides, HCV mimotope polypeptides or nucleic acid encoding such polypeptides.

HCV mimotope polypeptides contain HCV epitopes, but have a different sequence than a naturally occurring HCV antigen. A HCV mimotope can be fused to a naturally occurring HCV antigen. References describing techniques for producing mimotopes in general and describing different HCV mimotopes are

provided in Felici et al. U.S. Patent No. 5,994,083 and Nicosia et al., International Application Number WO 99/60132.

VII. PHARMACEUTICAL ADMINISTRATION

HCV vaccines can be formulated and administered to a patient using the guidance provided herein along with techniques well known in the art. Guidelines for pharmaceutical administration in general are provided in, for example, *Modern Vaccinology*, Ed. Kurstak, Plenum Med. Co. 1994; *Remington's Pharmaceutical Sciences 18th Edition*, Ed. Gennaro, Mack Publishing, 1990; and *Modern Pharmaceutics 2nd Edition*, Eds. Banker and Rhodes, Marcel Dekker, Inc., 1990, each of which are hereby incorporated by reference herein.

HCV vaccines can be administered by different routes such intravenous, intraperitoneal, subcutaneous, intramuscular, intradermal, impression through the skin, or nasal. A preferred route is intramuscular.

Intramuscular administration can be preformed using different techniques such as by injection with or without one or more electric pulses. Electric mediated transfer can assist genetic immunization by stimulating both humoral and cellular immune responses.

Vaccine injection can be performed using different techniques, such as by employing a needle or a needless injection system. An example of a needless injection system is a jet injection device. (Donnelly *et al.*, International Publication Number WO 99/52463.)

A. Electrically Mediated Transfer

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Electrically mediated transfer or Gene Electro-Transfer (GET) can be performed by delivering suitable electric pulses after nucleic acid injection. (See Mathiesen, International Publication Number WO 98/43702). Plasmid injection and electroporation can be performed using stainless needles. Needles can be used in couples, triplets or more complex patterns. In one configuration the needles are soldered on a printed circuit board that is a mechanical support and connects the needles to the electrical field generator by means of suitable cables.

The electrical stimulus is given in the form of electrical pulses. Pulses can be of different forms (square, sinusoidal, triangular, exponential decay) and different polarity (monopolar of positive or negative polarity, bipolar). Pulses can be delivered either at constant voltage or constant current modality.

Different patterns of electric treatment can be used to introduce nucleic acid vaccines including HCV and other nucleic acid vaccines into a patient. Possible patterns of electric treatment include the following:

Treatment 1: 10 trains of 1000 square bipolar pulses delivered every other second, pulse length 0.2 msec/phase, frequency 1000 Hz, constant voltage mode, 45 Volts/phase, floating current.

Treatment 2: 2 trains of 100 square bipolar pulses delivered every other second, pulse length 2 msec/phase, frequency 100 Hz, constant current mode, 100 mA/phase, floating voltage.

Treatment 3: 2 trains of bipolar pulses at a pulse length of about 2 msec/phase, for a total length of about 3 seconds, where the actual current going through the tissue is fixed at about 50 mA.

Electric pulses are delivered through an electric field generator. A suitable generator can be composed of three independent hardware elements assembled in a common chassis and driven by a portable PC which runs the driving program. The software manages both basic and accessory functions. The elements of the device are: (1) signal generator driven by a microprocessor, (2) power amplifier and (3) digital oscilloscope.

The signal generator delivers signals having arbitrary frequency and shape in a given range under software control. The same software has an interactive editor for the waveform to be delivered. The generator features a digitally controlled current limiting device (a safety feature to control the maximal current output). The power amplifier can amplify the signal generated up to +/- 150 V. The oscilloscope is digital and is able to sample both the voltage and the current being delivered by the amplifier.

B. Pharmaceutical Carriers

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Pharmaceutically acceptable carriers facilitate storage and administration of a vaccine to a subject. Examples of pharmaceutically acceptable carriers are described herein. Additional pharmaceutical acceptable carriers are well known in the art.

Pharmaceutically acceptable carriers may contain different components such a buffer, normal saline or phosphate buffered saline, sucrose, salts and polysorbate. An example of a pharmaceutically acceptable carrier is follows: 2.5-10 mM TRIS buffer, preferably about 5 mM TRIS buffer; 25-100 mM NaCl, preferably

about 75 mM NaCl; 2.5-10% sucrose, preferably about 5% sucrose; 0.01 -2 mM MgCl₂; and 0.001%-0.01% polysorbate 80 (plant derived). The pH is preferably from about 7.0-9.0, more preferably about 8.0. A specific example of a carrier contains 5 mM TRIS, 75 mM NaCl, 5% sucrose, 1 mM MgCl₂, 0.005% polysorbate 80 at pH 8.0.

C. Dosing Regimes

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Suitable dosing regimens can be determined taking into account the efficacy of a particular vaccine and factors such as age, weight, sex and medical condition of a patient; the route of administration; the desired effect; and the number of doses. The efficacy of a particular vaccine depends on different factors such as the ability of a particular vaccine to produce polypeptide that is expressed and processed in a cell and presented in the context of MHC class I and II complexes.

HCV encoding nucleic acid administered to a patient can be part of different types of vectors including viral vectors such as adenovector, and DNA plasmid vaccines. In different embodiments concerning administration of a DNA plasmid, about 0.1 to 10 mg of plasmid is administered to a patient, and about 1 to 5 mg of plasmid is administered to a patient. In different embodiments concerning administration of a viral vector, preferably an adenoviral vector, about 105 to 1011 viral particles are administered to a patient, and about 107 to 1010 viral particles are administered to a patient.

Viral vector vaccines and DNA plasmid vaccines may be administered alone, or may be part of a prime and boost administration regimen. A mixed modality priming and booster inoculation involves either priming with a DNA vaccine and boosting with viral vector vaccine, or priming with a viral vector vaccine and boosting with a DNA vaccine.

Multiple priming, for example, about to 2-4 or more may be used. The length of time between priming and boost may typically vary from about four months to a year, but other time frames may be used. The use of a priming regimen with a DNA vaccine may be preferred in situations where a person has a pre-existing anti-adenovirus immune response.

In an embodiment of the present invention, $1x10^7$ to $1x10^{12}$ particles and preferably about $1x10^{10}$ to $1x10^{11}$ particles of adenovector is administered directly into muscle tissue. Following initial vaccination a boost is performed with an adenovector or DNA vaccine.

In another embodiment of the present invention initial vaccination is performed with a DNA vaccine directly into muscle tissue. Following initial vaccination a boost is performed with an adenovector or DNA vaccine.

Agents such as interleukin-12, GM-CSF, B7-1, B7-2, IP10, Mig-1 can be coadministered to boost the immune response. The agents can be coadministered as proteins or through use of nucleic acid vectors.

D. Heterologous Prime-Boost

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Heterologous prime-boost is a mixed modality involving the use of one type of viral vector for priming and another type of viral vector for boosting. The heterologous prime-boost can involve related vectors such as vectors based on different adenovirus serotypes and more distantly related viruses such adenovirus and poxvirus. The use of poxvirus and adenovirus vectors to protect mice against malaria is illustrated by Gilbert *et al.*, *Vaccine* 20:1039-1045, 2002.

Different embodiments concerning priming and boosting involve the following types of vectors expressing desired antigens such as Met-NS3-NS4A-NS4B-NS5A-NS5B: Ad5 vector followed by Ad6 vector; Ad6 vector followed by Ad5 vector; Ad5 vector followed by poxvirus vector; poxvirus vector followed by Ad5 vector; Ad6 vector followed by poxvirus vector; and poxvirus vector followed by Ad6 vector.

The length of time between priming and boosting typically varies from about four months to a year, but other time frames may be used. The minimum time frame should be sufficient to allow for an immunological rest. In an embodiment, this rest is for a period of at least 6 months. Priming may involve multiple priming with one type of vector, such as 2-4 primings.

Expression cassettes present in a poxvirus vector should contain a promoter either native to, or derived from, the poxvirus of interest or another poxvirus member. Different strategies for constructing and employing different types of poxvirus based vectors including those based on vaccinia virus, modified vaccinia virus, avipoxvirus, raccoon poxvirus, modified vaccinia virus Ankara, canarypoxviruses (such as ALVAC), fowlpoxviruses, cowpoxviruses, and NYVAC are well known in the art. (Moss, Current Topics in Microbiology and Immunology 158:25-38, 1982; Earl et al., In Current Protocols in Molecular Biology, Ausubel et al. eds., New York: Greene Publishing Associates & Wiley Interscience;

35 1991:16.16.1-16.16.7, Child et al., Virology 174(2):625-9, 1990; Tartaglia et al.,

Virology 188:217-232, 1992; U.S. Patent Nos., 4,603,112, 4,722,848, 4,769,330, 5,110,587, 5,174,993, 5,185,146, 5,266,313, 5,505,941, 5,863,542, and 5,942,235.

E. Adjuvants

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HCV vaccines can be formulated with an adjuvant. Adjuvants are particularly useful for DNA plasmid vaccines. Examples of adjuvants are alum, AlPO4, alhydrogel, Lipid-A and derivatives or variants thereof, Freund's incomplete adjuvant, neutral liposomes, liposomes containing the vaccine and cytokines, non-ionic block copolymers, and chemokines.

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Non-ionic block polymers containing polyoxyethylene (POE) and polyxylpropylene (POP), such as POE-POP-POE block copolymers may be used as an adjuvant. (Newman et al., Critical Reviews in Therapeutic Drug Carrier Systems 15:89-142, 1998.) The immune response of a nucleic acid can be enhanced using a non-ionic block copolymer combined with an anionic surfactant.

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A specific example of an adjuvant formulation is one containing CRL-1005 (CytRx Research Laboratories), DNA, and benzylalkonium chloride (BAK). The formulation can be prepared by adding pure polymer to a cold (<5°C) solution of plasmid DNA in PBS using a positive displacement pipette. The solution is then vortexed to solubilize the polymer. After complete solubilization of the polymer a clear solution is obtained at temperatures below the cloud point of the polymer (~6-7°C). Approximately 4 mM BAK is then added to the DNA/CRL-1005 solution in PBS, by slow addition of a dilute solution of BAK dissolved in PBS. The initial DNA concentration is approximately 6 mg/mL before the addition of polymer and BAK, and the final DNA concentration is about 5 mg/mL. After BAK addition the formulation is vortexed extensively, while the temperature is allowed to increase from ~ 2°C to above the cloud point. The formulation is then placed on ice to decrease the temperature below the cloud point. Then, the formulation is vortexed while the temperature is allowed to increase from ~2°C to above the cloud point. Cooling and mixing while the temperature is allowed to increase from ~2°C to above the cloud point is repeated several times, until the particle size of the formulation is about 200-500 nm, as measured by dynamic light scattering. The formulation is then stored on ice until the solution is clear, then placed in storage at -70°C. Before use, the formulation is allowed to thaw at room temperature.

F. Vaccine Storage

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Adenovector and DNA vaccines can be stored using different types of buffers. For example, buffer A105 described in Example 9 *infra*. can be used to for vector storage.

Storage of DNA can be enhanced by removal or chelation of trace metal ions. Reagents such as succinic or malic acid, and chelators can be used to enhance DNA vaccine stability. Examples of chelators include multiple phosphate ligands and EDTA. The inclusion of non-reducing free radical scavengers, such as ethanol or glycerol, can also be useful to prevent damage of DNA plasmid from free radical production. Furthermore, the buffer type, pH, salt concentration, light exposure, as well as the type of sterilization process used to prepare the vials, may be controlled in the formulation to optimize the stability of the DNA vaccine.

VII. EXAMPLES

Examples are provided below to further illustrate different features of the present invention. The examples also illustrate useful methodology for practicing the invention. These examples do not limit the claimed invention.

Example 1: Met-NS3-NS4A-NS4B-NS5A-NS5B Expression Cassettes

Different gene expression cassettes encoding HCV NS3-NS4A-NS4B-NS5A-NS5B were constructed based on a 1b subtype HCV BK strain. The encoded sequences had either (1) an active NS5B sequence ("NS"), (2) an inactive NS5B sequence ("NSmut"), (3) a codon optimized sequence with an inactive NS5B sequence ("NSOPTmut"). The expression cassettes also contained a CMV promoter/enhancer and the BGH polyadenylation signal.

The NS nucleotide sequence (SEQ. ID. NO. 5) differs from HCV BK strain GenBank accession number M58335 by 30 out of 5952 nucleotides. The NS amino acid sequence (SEQ. ID. NO. 6) differs from the corresponding 1b genotype HCV BK strain by 7 out of 1984 amino acids. To allow for initiation of translation an ATG codon is present at the 5' end of the NS sequence. A TGA termination sequence is present at the 3' end of the NS sequence.

The NSmut nucleotide sequence (SEQ. ID. NO. 2, Figure 2), is similar to the NS sequence. The differences between NSmut and NS include NSmut having an altered NS5B catalytic site; an optimal ribosome binding site at the 5' end; and a TAAA termination sequence at the 3' end. The alterations in NS5B comprise bases

5138 to 5146, which encode amino acids 1711 to 1713. The alterations result in a change of amino acids GlyAspAsp into AlaAlaGly and creates an inactive form of the NS5B RNA-dependent RNA-polymerase NS5B.

The NSOPTmut sequence (SEQ. ID. NO. 3, Figure 3) was designed

based on the amino acid sequence encoded by NSmut. The NSmut amino acid
sequence was back translated into a nucleotide sequence with the GCG (Wisconsin
Package version 10, Genetics Computer Group, GCG, Madison, Wisc.)

BACKTRANSLATE program. To generate a NSOPTmut nucleotide sequence where
each amino acid is coded for by the corresponding most frequently observed human
codon, the program was run choosing as parameter the generation of the most
probable nucleotide sequence and specifying the codon frequency table of highly
expressed human genes (human_high.cod) available within the GCG Package as
translation scheme.

Example 2: Generation pV1Jns plasmid with NS, NSmut or NSOPTmut Sequences

pV1Jns plasmids containing either the NS sequence, NSmut sequence
or NSOPTmut sequences were generated and characterised as follows:

pV1Ins Plasmid with the NS Sequence

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The coding region Met-NS3-NS4A-NS4B-NS5A and the coding region Met-NS3-NS4A-NS4B-NS5A-NS5B from a HCV BK type strain (Tomei *et al., J. Virol.* 67:4017-4026, 1993) were cloned into pcDNA3 plasmid (Invitrogen), generating pcD3-5a and pcD3-5b vectors, respectively. PcD3-5A was digested with Hind III, blunt-ended with Klenow fill-in and subsequently digested with Xba I, to generate a fragment corresponding to the coding region of Met-NS3-NS4A-NS4B-NS5A. The fragment was cloned into pV1Jns-poly, digested with Bgl II blunt-ended with Klenow fill-in and subsequently digested with Xba I, generating pV1JnsNS3-5A.

pV1Jns-poly is a derivative of pV1JnsA plasmid (Montgomery et al., DNA and Cell Biol. 12:777-783, 1993), modified by insertion of a polylinker containing recognition sites for XbaI, PmeI, PacI into the unique BgIII and NotI restriction sites. The pV1Jns plasmid with the NS sequence (pV1JnsNS3-5B) was obtained by homologous recombination into the bacterial strain BJ5183, cotransforming pV1JNS3-5A linearized with XbaI and NotI digestion and a PCR fragment containing approximately 200 bp of NS5A, NS5B coding sequence and

approximately 60 bp of the BGH polyadenylation signal. The resulting plasmid represents pV1Jns-NS.

pV1Jns-NS can be summarized as follows:

Bases 1 to 1881 of pV1JnsA

5 an additional AGCTT

then the Met-NS3-NS5B sequence (SEQ. ID. NO. 5)

then the wt TGA stop

an additional TCTAGAGCGTTTAAACCCTTAATTAAGG (SEQ. ID.

NO. 14)

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10 Bases 1912 to 4909 of pV1JnsA

pV1Ins Plasmid with the NSmut Sequence

The V1JnsNS3-5A plasmid was modified at the 5' of the NS3 coding sequence by addition of a full Kozak sequence. The plasmid (V1JNS3-5Akozak) was obtained by homologous recombination into the bacterial strain BJ5183, cotransforming V1JNS3-5A linearized by AfIII digestion and a PCR fragment containing the proximal part of Intron A, the restriction site BgIII, a full Kozak translation initiation sequence and part of the NS3 coding sequence.

The resulting plasmid (V1JNS3-5Akozak) was linearized with Xba I

digestion and co-transformed into the bacterial strain BJ5183 with a PCR fragment,
containing approximately 200 bp of NS5A, the NS5B mutated sequence, the strong
translation termination TAAA and approximately 60 bp of the BGH polyadenylation
signal. The PCR fragment was obtained by assembling two 22bp-overlapping
fragments where mutations were introduced by the oligonucleotides used for their
amplification. The resulting plasmid represents pV1Jns-NSmut.

pV1Jns-NSmut can be summarized as follows:

Bases 1 to 1882 of pV1JnsA

then the kozak Met-NS3-NS5B(mut) TAAA sequence (SEQ. ID. NO. 2)

an additional TCTAGA

30 Bases 1925 to 4909 of pV1JnsA

pVIJns Plasmid with the NSOPTmut Sequence

The human codon-optimized synthetic gene (NSOPTmut) with mutated NS5B to abrogate enzymatic activity, full Kozak translation initiation sequence and a strong translation termination was digested with BamHI and SalI

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restriction sites present at the 5' and 3' end of the gene. The gene was then cloned into the BglII and SalI restriction sites present in the polylinker of pV1JnsA plasmid, generating pV1Jns-NSOPTmut.

pV1Jns-NSOPTmut can be summarized as follows:

5 Bases 1 to 1881 of pV1JnsA

an additional C

then

kozak Met-NS3-NS5B(optmut) TAAA sequence (SEQ. ID. NO. 3)

an additional TTTAAATGTTTAAAC (SEQ. ID. NO. 15)

Bases

1905 to 4909 of pV1JnsA

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Plasmids Characterization

Expression of HCV NS proteins was tested by transfection of HEK 293 cells, grown in 10% FCS/DMEM supplemented by L-glutamine (final 4 mM). Twenty-four hours before transfection, cells were plated in 6-well 35 mm diameter, to reach 90-95% confluence on the day of transfection. Forty nanograms of plasmid DNA (previously assessed as a non-saturating DNA amount) were co-transfected with 100 ng of pRSV-Luc plasmid containing the luciferase reporter gene under the control of Rous sarcoma virus promoter, using the LIPOFECTAMINE 2000 reagent. Cells were kept in a CO₂ incubator for 48 hours at 37 °C.

Cell extracts were prepared in 1% Triton/TEN buffer. The extracts were normalized for Luciferase activity, and run in serial dilution on 10% SDSacrylamide gel. Proteins were transferred on nitrocellulose and assayed with antibodies directed against NS3, NS5A and NS5B to assess strength of expression and correct proteolytic cleavage. Mock-transfected cells were used as a negative control.

Results from representative experiments testing pV1JnsNS, pV1JnsNSmut and pV1JnsNSOPTmut are shown in Figure 12.

Example 3: Mice Immunization with Plasmid DNA Vectors

The DNA plasmids pV1Jns-NS, pV1Jns-NSmut and pV1Jns-NSOPT mut were injected in different mice strains to evaluate their potential to elicit anti-HCV immune responses. Two different strains (Balb/C and C57Black6, N=9-10) were injected intramuscularly with 25 or 50 µg of DNA followed by electrical pluses. Each animal received two doses at three weeks interval.

Humoral immune response elicited in C57Black6 mice against the NS3 protein was measured in post dose two sera by ELISA on bacterially expressed NS3 35

protease domain. Antibodies specific for the tested antigen were detected in animals immunized with all three vectors with geometric mean titers (GMT) ranging from 94000 to 133000 (Tables 1-3).

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Table 1: pV1ins-NS

		· ·				_				GMT
Mice n.	l	2	3	4	5	6	7	8	9	
Titer	105466	891980	78799	39496	543542	182139	32351	95028	67800	94553

Table 2: pV1jns-NSmut

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											GMT
Mice n.	11	12	13	14	15	16	17	18	19	20	
Titer	202981	55670	130786	49748	17672	174958	44304	37337	78182	193695	75083

Table 3: pV1jns-NSOPTmut

	· · · · · · · · · · · · · · · · · · ·										GMT
Mice n.	21	22	23	24	25	26	27	28	29	30	
Titer	310349	43645	63496	82174	630778	297259	66861	146735	173506	77732	133165

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A T cell response was measured in C57Black6 mice immunized with two intramuscular injections at three weeks interval with 25 μg of plasmid DNA. Quantitative ELIspot assay was performed to determine the number of IFNγ secreting T cells in response to five pools of 20mer peptides overlapping by ten residues encompassing the NS3-NS5B sequence. Specific CD8+ response was analyzed by the same assay using a 20mer peptide encompassing a CD8+ epitope for C57Black6 mice (pep1480).

Cells secreting IFN γ in an antigen specific-manner were detected using a standard ELIspot assay. T cell response in C57Black6 mice immunized with two intramuscular injections at three weeks interval with 50 μ g of plasmid DNA, was

analyzed by the same ELIspot assay measuring the number of IFN γ secreting T cells in response to five pools of 20mer peptides overlapping by ten residues encompassing the NS3-NS5B sequence.

Spleen cells were prepared from immunized mice and re-suspended in R10 medium (RPMI 1640 supplemented with 10% FCS, 2 mM L-Glutamine, 50 U/ml-50μg/ml Penicillin/Streptomycin, 10 mM Hepes, 50 μM 2-mercapto-ethanol). Multiscreen 96-well Filtration Plates (Millipore, Cat. No. MAIPS4510, Millipore Corporation, 80 Ashby Road Bedford, MA) were coated with purified rat anti-mouse INFγ antibody (PharMingen, Cat. No. 18181D, PharmiMingen, 10975 Torreyana Road, San Diego, California 92121-1111 USA). After overnight incubation, plates were washed with PBS 1X/0.005% Tween and blocked with 250 μl/well of R10 medium.

Splenocytes from immunized mice were prepared and incubated for twenty-four hours in the presence or absence of 10 μM peptide at a density of 2.5 X 10⁵/well or 5 X 10⁵/well. After extensive washing (PBS 1X/0.005% Tween), biotinylated rat anti-mouse IFNγ antibody (PharMingen, Cat. No. 18112D, PharMingen, 10975 Torreyana Road, San Diego, California 92121-1111 USA) was added and incubated overnight at 4° C. For development, streptavidin-AKP (PharMingen, Cat. No. 13043E, PharMingen, 10975 Torreyana Road, San Diego, California 92121-1111 USA) and 1-StepTM NBT-BCIP development solution (Pierce, Cat. No. 34042, Pierce, P.O. Box 117, Rockford, IL 61105 USA) were added.

Pools of 20mer overlapping peptides encompassing the entire sequence of the HCV BK strain NS3 to NS5B were used to reveal HCV-specific IFNγ-secreting T cells. Similarly a single 20mer peptide encompassing a CD8+ epitope for C57Black6 mice was used to detect CD8 response. Representative data from groups of C57Black6 and Balb/C mice (N=9-10) immunized with two injections of 25 or 50 μg of plasmid vectors pV1Jns-NS, pV1Jns-NSmut and pV1Jns-NSOPTmut are shown in Figures 13A and 13B.

30 Example 4: Immunization of Rhesus Macaques

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Rhesus macaques (N=3) were immunized by intramuscular injection with 5mg of plasmid pV1Jns-NSOPTmut in 7.5mg/ml CRL1005, Benzalkonium chloride 0.6 mM. Each animal received two doses in the deltoid muscle at 0, and 4 weeks.

CMI was measured at different time points by IFN- γ ELISPOT. This assay measures HCV antigen-specific CD8+ and CD4+ T lymphocyte responses, and can be used for a variety of mammals, such as humans, rhesus monkeys, mice, and rats.

The use of a specific peptide or a pool of peptides can simplify antigen presentation in CTL cytotoxicity assays, interferon-gamma ELISPOT assays and interferon-gamma intracellular staining assays. Peptides based on the amino acid sequence of various HCV proteins (core, E2, NS3, NS4A, NS4B, NS5A, NS5B) were prepared for use in these assays to measure immune responses in HCV DNA and adenovirus vector vaccinated rhesus monkeys, as well as in HCV-infected humans. The individual peptides are overlapping 20-mers, offset by 10 amino acids. Large pools of peptides can be used to detect an overall response to HCV proteins while smaller pools and individual peptides may be used to define the epitope specificity of a response.

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IFNy ELISPOT

The IFNγ-ELISPOT assay provides a quantitative determination of HCV-specific T lymphocyte responses. PBMC are serially diluted and placed in microplate wells coated with anti-rhesus IFN-γ antibody (MD-1 U-Cytech). They are cultured with a HCV peptide pool for 20 hours, resulting in the restimulation of the precursor cells and secretion of IFN-γ. The cells are washed away, leaving the secreted IFN bound to the antibody-coated wells in concentrated areas where the cells were sitting. The captured IFN is detected with biotinylated anti-rhesus IFN antibody (detector Ab U-Cytech) followed by alkaline phosphatase-conjugated streptavidin (Pharmingen 13043E). The addition of insoluble alkaline phosphatase substrate results in dark spots in the wells at the sites where the cells were located, leaving one spot for each T cell that secreted IFN-γ.

The number of spots per well is directly related to the precursor frequency of antigen-specific T cells. Gamma interferon was selected as the cytokine visualized in this assay (using species specific anti-gamma interferon monoclonal antibodies) because it is the most common, and one of the most abundant cytokines synthesized and secreted by activated T lymphocytes. For this assay, the number of spot forming cells (SFC) per million PBMCs is determined for samples in the

presence and absence (media control) of peptide antigens. Data from Rhesus macaques on PBMC from post dose two material are shown in Table 4.

Table 4

	PV1J-NSOPTmut								
Pep pools	21G	99C161	99C166						
F (NS3p)	8	10	170						
G (NS3h)	7	592	229						
H (NS4)	3	14	16						
I (NS5a)	5	71	36						
L (NS5b)	14	23	11						
M (NS5b)	3	35	8						
DMSO	2	4	5						

INFγELISPOT on PBMC from Rhesus monkeys immunized with two injections of 5 mg DNA/dose in OPTIVAX/BAK of plasmid pV1Jns-NSOPTmut. Data are expressed as SFC7 106 PBMC.

Example 5: Construction of Ad6 Pre-Adenovirus Plasmids

Ad6 pre-adenovirus plasmids were obtained as follows:

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Construction of pAd6 E1-E3+ Pre-adenovirus Plasmid

An Ad6 based pre-adenovirus plasmid which can be used to generate first generation Ad6 vectors was constructed either taking advantage of the extensive sequence identity (approx. 98%) between Ad5 and Ad6 or containing only Ad6 regions. Homologous recombination was used to clone wtAd6 sequences into a bacterial plasmid.

A general strategy used to recover pAd6E1-E3+ as a bacterial plasmid containing Ad5 and Ad6 regions is illustrated in Figure 10. Cotransformation of BJ 5183 bacteria with purified wt Ad6 viral DNA and a second DNA fragment termed the Ad5 ITR cassette resulted in the circularization of the viral genome by homologous recombination. The ITR cassette contains sequences from the right (bp 33798 to 35935) and left (bp 1 to 341 and bp 3525 to 5767) end of the Ad5 genome separated by plasmid sequences containing a bacterial origin of replication and an ampicillin resistance gene. The ITR cassette contains a deletion of E1 sequences from

Ad5 342 to 3524. The Ad5 sequences in the ITR cassette provide regions of homology with the purified Ad6 viral DNA in which recombination can occur.

Potential clones were screened by restriction analysis and one clone was selected as pAd6E1-E3+. This clone was then sequenced in it entirety. pAd6E1-E3+ contains Ad5 sequences from bp 1 to 341 and from bp 3525 to 5548, Ad6 bp 5542 to 33784, and Ad5 bp 33967 to 35935 (bp numbers refer to the wt sequence for both Ad5 and Ad6). pAd6E1-E3+ contains the coding sequences for all Ad6 virion structural proteins which constitute its serotype specificity.

A general strategy used to recover pAd6E1-E3+ as a bacterial plasmid

containing Ad6 regions is illustrated in Figure 11. Cotransformation of BJ 5183

bacteria with purified wt Ad6 viral DNA and a second DNA fragment termed the Ad6

ITR cassette resulted in the circularization of the viral genome by homologous

recombination. The ITR cassette contains sequences from the right (bp 35460 to

35759) and left (bp 1 to 450 and bp 3508 to 3807) end of the Ad6 genome separated

by plasmid sequences containing a bacterial origin of replication and an ampicillin

resistance gene. These three segments were generated by PCR and cloned

sequentially into pNEB193, generating pNEBAd6-3 (the ITR cassette). The ITR

cassette contains a deletion of E1 sequences from Ad5 451 to 3507. The Ad6

sequences in the ITR cassette provide regions of homology with the purified Ad6 viral

DNA in which recombination can occur.

Construction of pAd6 E1-E3- pre-adenovirus plasmids

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Ad6 based vectors containing A5 regions and deleted in the E3 region were constructed starting with pAd6E1-E3+ containing Ad5 regions. A 5322 bp subfragment of pAd6E1-E3+ containing the E3 region (Ad6 bp 25871 to 31192) was subcloned into pABS.3 generating pABSAd6E3. Three E3 deletions were then made in this plasmid generating three new plasmids pABSAd6E3(1.8Kb) (deleted for Ad6 bp 28602 to 30440), pABSAd6E3(2.3Kb) (deleted for Ad6 bp 28157 to 30437) and pABSAd6E3(2.6Kb) (deleted for Ad6 bp 28157 to 30788). Bacterial recombination was then used to substitute the three E3 deletions back into pAd6E1-E3+ generating the Ad6 genome plasmids pAd6E1-E3-1.8Kb, pAd6E1-E3-2.3Kb and pAd6E1-E3-2.6Kb.

Example 6: Generation of Ad5 Genome Plasmid with the NS Sequence

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A pcDNA3 plasmid (Invitrogen) containing the coding region NS3-NS4A-NS4B-NS5A was digested with *Xmn*I and *Nru*I restriction sites and the DNA fragment containing the CMV promoter, the NS3-NS4A-NS4B-NS5A coding sequence and the Bovine Growth Hormone (BGH) polyadenylation signal was cloned into the unique *EcorV* restriction site of the shuttle vector pDelE1Spa, generating the Sva3-5A vector.

A pcDNA3 plasmid containing the coding region NS3-NS4A-NS4B-NS5A-NS5B was digested with *XmnI* and *EcorI* (partial digestion), and the DNA fragment containing part of NS5A, NS5B gene and the BGH polyadenylation signal was cloned into the Sva3-5A vector, digested *EcorI* and *BgIII* blunted with Klenow, generating the Sva3-5B vector.

The Sva3-5B vector was finally digested *SspI* and *Bst*1107I restriction sites and the DNA fragment containing the expression cassette (CMV promoter, NS3-NS4A-NS4B-NS5A-NS5B coding sequence and the BGH polyadenylation signal) flanked by adenovirus sequences was co-transformed with pAd5HVO (E1-,E3-) ClaI linearized genome plasmid into the bacterial strain BJ5183, to generate pAd5HVONS. pAd5HVO contains Ad5 bp 1 to 341, bp 3525 to 28133 and bp 30818 to 35935.

Example 7: Generation of Adenovirus Genome Plasmids with the NSmut Sequence
Adenovirus genome plasmids containing an NS-mut sequence were
generated in an Ad5 or Ad6 background. The Ad6 background contained Ad5 regions
at bases 1 to 450, 3511 to 5548 and 33967 to 35935.

pV1JNS3-5Akozak was digested with *Bgl*II and *Xba*I restriction enzymes and the DNA fragment containing the Kozak sequence and the sequence coding NS3-NS4A-NS4B-NS5A was cloned into a *Bgl*II and XbaI digested polypMRKpdelE1 shuttle vector. The resulting vector was designated shNS3-5Akozak.

PolypMRKpdelE1 is a derivative of RKpdelE1(Pac/pIX/pack450) + CMVmin+BGHpA(str.) modified by the insertion of a polylinker containing recognition sites for BglII, PmeI, SwaI, XbaI, SalI, into the unique BglII restriction site present downstream the CMV promoter. MRKpdelE1(Pac/pIX/pack450) + CMVmin + BGHpA(str.) contains Ad5 sequences from bp 1 to 5792 with a deletion of E1 sequences from bp 451 to 3510. The human CMV promoter and BGH polyadenylation signal were inserted into the E1 deletion in an E1 parallel orientation with a unique BglII site separating them.

The NS5B fragment, mutated to abrogate enzymatic activity and with a strong translation termination at the 3' end, was obtained by assembly PCR and inserted into the shNS3-5Akozak vector via homologous recombination, generating polypMRKpdelE1NSmut. In polypMRKpdelE1NSmut the NS-mut coding sequence is under the control of CMV promoter and the BGH polyadenylation signal is present downstream.

The gene expression cassette and the flanking regions which contain adenovirus sequences allowing homologous recombination were excised by digestion with *PacI* and *Bst*1107I restriction enzymes and co-transformed with either pAd5HVO (E1-,E3-) or pAd6E1-E3-2.6Kb *ClaI* linearized genome plasmids into the bacterial strain BJ5183, to generate pAd5HVONSmut and pAd6E1-,E3-NSmut, respectively.

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pAd6E1-E3-2.6Kb contains Ad5 bp 1 to 341 and from bp 3525 to 5548, Ad6 bp 5542 to 28157 and from bp 30788 to 33784, and Ad5 bp 33967 to 35935 (bp numbers refer to the wt sequence for both Ad5 and Ad6). In both plasmids the viral ITR's are joined by plasmid sequences that contain the bacterial origin of replication and an ampicillin resistance gene.

Example 8: Generation of Adenovirus Genome Plasmids with the NSOPTmut

The human codon-optimized synthetic gene (NSOPTmut) provided by SEQ. ID. NO. 3 cloned into a pCRBlunt vector (Invitrogen) was digested with BamH1 and SalI restriction enzymes and cloned into BgIII and SalI restriction sites present in the shuttle vector polypMRKpdelE1. The resulting clone (polypMRKpdelE1NSOPTmut) was digested with PacI and Bst1107I restriction enzymes and co-transformed with either pAd5HVO (E1-,E3-) or pAd6E1-E3-2.6Kb

ClaI linearized genome plasmids, into the bacterial strain BJ5183, to generate pAd5HVONSOPTmut and pAd6E1-,E3-NSOPTmut, respectively.

Example 9: Rescue and Amplification of Adenovirus Vectors

Adenovectors were rescued in Per.6 cells. Per.C6 were grown in 10% FCS / DMEM supplemented by L-glutamine (final 4mM), penicillin/streptomycin (final 100 IU/ml) and 10 mM MgCl₂. After infection, cells were kept in the same medium supplemented by 5% horse serum (HS). For viral rescue, 2.5 X 10⁶ Per.C6 were plated in 6 cm Ø Petri dishes.

Twenty-four hours after plating, cells were transfected by calcium phosphate method with 10 μ g of the *Pac I* linearized adenoviral DNA. The DNA precipitate was left on the cells for 4 hours. The medium was removed and 5% HS/DMEM was added.

Cells were kept in a CO₂ incubator until a cytopathic effect was visible (1 week). Cells and supernatant were recovered and subjected to 3X freeze/thawing cycles (liquid nitrogen / water bath at 37°C). The lysate was centrifuged at 3000 rpm at - 4°C for 20 minutes and the recovered supernatant (corresponding to a cell lysate containing virus passed on cells only once; P1) was used, in the amount of 1 ml/dish, to infect 80-90% confluent Per.C6 in 10 cm ø Petri dishes. The infected cells were incubated until a cytopathic effect was visible, cells and supernatant recovered and the lysate prepared as described above (P2).

P2 lysate (4 ml) were used to infect 2 X 15 cm ø Petri dishes. The lysate recovered from this infection (P3) was kept in aliquots at -80°C as a stock of virus to be used as starting point for big viral preparations. In this case, 1 ml of the stock was enough to infect 2 X 15 cm ø Petri dishes and resulting lysate (P4) was used for the infection of the Petri dishes devoted to the large scale infection.

Further amplification was obtained from the P4 lysate which was diluted in medium without FCS and used to infect 30 X 15 cm Ø Petri dishes (with Per.C6 80%-90% confluent) in the amount of 10 ml/dish. Cells were incubated 1 hour in the CO₂ incubator, mixing gently every 20 minutes. 12 ml / dish of 5% HS / DMEM was added and cells were incubated until a cytopathic effect was visible (about 48 hours).

Cells and supernatant were collected and centrifuged at 2K rpm for 20 minutes at 4° C. The pellet was resuspended in 15 ml of 0.1 M Tris pH=8.0. Cells were lysed by 3X freeze/thawing cycles (liquid nitrogen / water bath at 37° C). 150 μ l of 2 M MgCl₂ and 75 μ l of DNAse (10 mg of bovine pancreatic deoxyribonuclease I in 10 ml of 20 mM Tris-HCl pH= 7.4, 50 mM NaCl, 1 mM dithiothreitol, 0.1 mg/ml bovine serum albumin, 50% glycerol) were added. After a 1 hour incubation at 37° C in a water bath (vortex every 15 minutes) the lysate was centrifuged at 4K rpm for 15 minutes at 4° C. The recovered supernatant was ready to be applied on CsCl gradient.

The CsCl gradients were prepared in SW40 ultra-clear tubes as

follows:

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0.5 ml of 1.5d CsCl

35 3 ml of 1.35d CsCl

3 ml of 1.25d CsCl

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5-ml/ tube of viral supernatant was applied.

If necessary, the tubes were topped up with 0.1 M tris-Cl pH=8.0. Tubes were centrifuged at 35K rpm for 1 hour at -10°C with rotor SW40. The viral bands (located at the 1.25/1.35 interface) were collected using a syringe.

The virus was transferred into a new SW40 ultraclear tube and 1.35d CsCl was added to top the tube up. After centrifugation at 35K rpm for 24 hours at 10° C in the rotor SW40, the virus was collected in the smallest possible volume and dialyzed extensively against buffer A105 (5 mM Tris, 5% sucrose, 75 mM NaCl, 1 mM MgCl₂, 0.005% polysorbate 80 pH=8.0). After dialysis, glycerol was added to final 10% and the virus was stored in aliquots at -80° C.

Example 10: Enhanced Adenovector Rescue

First generation Ad5 and Ad6 vectors carrying HCV NSOPTmut

transgene were found to be difficult to rescue. A possible block in the rescue process
might be attributed to an inefficient replication of plasmid DNA that is a sub-optimal
template for the replication machinery of adenovirus. The absence of the terminal
protein linked to the 5'ends of the DNA (normally present in the viral DNA),
associated with the very high G-C content of the transgene inserted in the E1 region of
the vector, may be causing a substantial reduction in replication rate of the plasmidderived adenovirus.

To set up a more efficient and reproducible procedure for rescuing Ad vectors, an expression vector (pE2; Figure 19) containing all E2 proteins (polymerase, pre-terminal protein and DNA binding protein) as well as E4 orf6 under the control of tet-inducible promoter was employed. The transfection of pE2 in combination with a normal preadeno plasmid in PerC6 and in 293 leads to a strong increase of Ad DNA replication and to a more efficient production of complete infectious adenovirus particles.

30 Plasmid Construction

pE2 is based on the cloning vector pBI (CLONTECH) with the addition of two elements to allow episomal replication and selection in cell culture: (1) the EBV-OriP (EBV [nt] 7421-8042) region permitting plasmid replication in synchrony with the cell cycle when EBNA-1 is expressed and (2) the hygromycin-B phosphotransferase (HPH)-resistance gene allowing a positive selection of

transformed cells. The two transcriptional units for the adenoviral genes E2 a and b and E4-Orf6 were constructed and assembled in pE2 as described below.

The Ad5-Polymerase Clal/SphI fragment and the Ad5-pTP Acc65/EcoRV fragment were obtained from pVac-Pol and pVac-pTP (Stunnemberg et al. NAR 16:2431-2444, 1988). Both fragments were filled with Klenow and cloned into the SalI (filled) and EcoRV sites of pBI, respectively obtaining pBI-Pol/pTP.

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EBV-OriP element from pCEP4 (Invitrogen) was first inserted within two chicken β-globin insulator dimers by cloning it into *BamHI* site of pJC13-1 (Chung *et al.*, *Cell 74(3)*:505-14, 1993). HS4-OriP fragment from pJC13-OriP was then cloned inside pSA1mv (a plasmid containing tk-Hygro-B resistance gene expression cassette as well as Ad5 replication origin), the ITR's arranged as head-to-tail junction, obtained by PCR from pFG140 (Graham, *EMBO J. 3*:2917-2922, 1984) using the following primers: 5'-TCGAATCGATACGCGAACCTACGC-3' (SEQ. ID. NO. 16) and 5'-TCGACGTGTCGACTTCGAAGCGCACACCAAAAACGTC-3' (SEQ. ID. NO. 17), thus generating pMVHS4Orip. A DNA fragment from pMVHS4Orip, containing the insulated OriP, Ad5 ITR junction and tk-HygroB cassette, was then inserted into pBI-Pol/pTP vector restricted *Asel/AatII* generating pBI-Pol/pTPHS4.

To construct the second transcriptional unit expressing Ad5-Orf6 as well as Ad5-DBP, E4orf6 (Ad 5 [nt] 33193-34077) obtained by PCR was first inserted into pBI vector, generating pBI-Orf6. Subsequently, DBP coding DNA sequence (Ad 5 [nt] 22443-24032) was inserted into pBI-Orf6 obtaining the second bi-directional Tet-regulated expression vector (pBI-DBP/E4orf6). The original polyA signals present in pBI were substituted with BGH and SV40 polyA.

pBI-DBP/E4orf6 was then modified by inserting a DNA fragment containing the Adeno5-ITRs arranged in head-to-tail junction plus the hygromicin B resistance gene obtained from plasmid pSA-1mv. The new plasmid pBI-DBP/E4orf6shuttle was then used as donor plasmid to insert the second tet-regulated transcriptional unit into pBI-Pol/pTPHS4 by homologous recombination using *E. coli* strain BJ5183 obtaining pE2.

Cell lines, Transfections and Virus Amplification

PerC6 cells were cultured in Dulbecco's modified Eagle's Medium (DMEM) plus 10% fetal bovine serum (FBS), 10 mM MgCl₂, penicillin (100 U/ml), streptomycin (100 μ g/ml) and 2 mM glutamine.

All transfections were performed using Lipofectamine2000 (Invitrogen) as described by the manufacturer. 90% confluent PERC.6TM planted in 6-cm plates were transfected with 3.5 μg of Ad5/6NSOPTmut pre-adeno plasmids, digested with PacI, alone or in combination with 5 μg pE2 plus 1 μg pUHD52.1. pUHD52.1 is the expression vector for the reverse tet transactivator 2 (rtTA2) (Urlinger *et al.*, *Proc. Natl. Acad. Sci. U.S.A. 97(14)*:7963-7968, 2000). Upon transfection, cells were cultivated in the presence of 1 μg/ml of doxycycline to activate pE2 expression. 7 days post-transfection cells were harvested and cell lysate was obtained by three cycles of freeze-thaw. Two ml of cell lysate were used to infect a second 6-cm dish of PerC6. Infected cells were cultivated until a full CPE was observed then harvested. The virus was serially passaged five times as described above, then purified on CsCl gradient. The DNA structure of the purified virus was controlled by endonuclease digestion and agarose gel electrophoresis analysis and compared to the original pre-adeno plasmid restriction pattern.

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Example 11: Partial Optimizeation of HCV Polyprotein Encoding Nucleic acid

Partial optimization of HCV polyprotein encoding nucleic acid was performed to facilitate the production of adenovectors containing codons optimized for expression in a human host. The overall objective was to provide for increased expression due to codon optimization, while facilitating the production of an adenovector encoding HCV polyprotein.

Several difficulties were encountered in producing an adenovector encoding HCV polyprotein with codons optimized for expression in a human host. An adenovector containing an optimized sequence (SEQ. ID. NO. 3) was found to be more difficult to synthesize and rescue than an adenovector containing a non-optimized sequence (SEQ. ID. NO. 2).

The difficulties in producing an adenovector containing SEQ. ID. NO. 3 were attributed to a high GC content. A particularly problemetic region was the region at about position 3900 of NSOPTmut (SEQ. ID. NO. 3).

Alternative versions of optimized HCV encoding nucleic acid sequence were designed to facilitate its use in an adenovector. The alternative versions, compared to NSOPTmut, were designed to have a lower overall GC content, to reduce/avoid the presence of potentially problematic motifis of consecutive G's or C's, while maintaining a high level of codon optimization to allow improved expression of the encoded polyprotein and the individual cleavage products.

A starting point for the generation of a suboptimally codon-optimized sequence is the coding region of the NSOPTmut nucleotide sequence (bases 7 to 5961 of SEQ. ID. NO. 3). Values for codon usage frequencies (normalized to a total of 1.0 for each amino acid) were taken from the file human_high.cod available in the Wisconsin Package Version 10.3 (Accelrys Inc., a wholly owned subsidiary of Pharmacopeia, Inc).

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To reduce the local and overall GC content a table defining preferred codon substitutions for each amino acid was manually generated. For each amino acid the codon having 1) a lower GC content as compared to the most frequent codon and 2) a relativly high observed codon usage frequency (as defined in human_high.cod) was choosen as the replacement codon. For example for Arg the codon with the highest frequency is CGC. Out of the other five alternative codons encoding Arg (CGG, AGG, AGA, CGT, CGA) three (AGG, CGT, CGA) reduce the GC content by 1 base, one (AGA) by two bases and one (CGG) by 0 bases. Since the AGA codon is listed in human_high.cod as having a relatively low usage frequency (0.1), the codon substituting CGC was therefore choosen to be AGG with a relative frequency of 0.18. Similar criteria were applied in order to establish codon replacements for the other amino acids resulting in the list shown in Table 5. Parameters applied in the following optimization procedure were determined empirically such that the resulting sequence maintained a considerably improved codon usage (for each amino acid) and the GC content (overall and in form of local stretches of consecutive G's and/or C's) was decreased.

Two examples of partial optimized HCV encoding sequences are provided by SEQ. ID. NO. 10 and SEQ. ID. NO. 11. SEQ. ID. NO. 10 provides a HCV encoding sequence that is partially optimized throughout. SEQ. ID. NO. 11 provides an HCV encoding sequence fully optimized for codon usage with the exception of a region that was partially optimized.

Codon optimization was performed using the following procedure:

Step 1) The coding region of the input fully optimized NSOPTmut sequence was analyzed using a sliding window of 3 codons (9 bases) shifting the window by one codon after each cycle. Whenever a stretch containing 5 or more consecutive C's and/or G's was detected in the window the following replacement rule was applied: Let N indicate the number of codon replacements previously performed. If N is odd replace the middle codon in the window with the codon specified in Table 5, if N is even replace the third terminal codon in the window with the codon

specified in a codon optimization table such as human_high.cod. If Leu or Val is present at the second or third codon do not apply any replacement in order not to introduce Leu or Val codons with very low relative codon usage frequency (see, for example, human_high.cod). In the following cycle analysis of the shifted window was then applied to a sequence containing the replacements of the previous cycle.

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The alternating replacement of the middle and terminal codon in the 3 codon window was found empirically to give a more satisfying overall maintenance of optimized codon usage while also reducing GC content (as judged from the final sequence after the procedure). In general, however, the precise replacement strategy depends on the amino acid sequence encoded by the nucelotide sequence under analysis and will have to be determined empirically.

Step 2) The sequence containing all the codon replacements performed during step 1) was then subjected to an additional analysis using a sliding window of 21 codons (63 bases) in length: according to an adjustable parameter the overall GC content in the window was determined. If the GC content in the window was higher than 70% the following codon replacement strategy was applied: In the window replace the codons for the amino acids Asn, Asp, Cys, Glu, His, Ile, Lys, Phe, Tyr by the codons given in Table 5. Restriction of the replacement to this set of amino acids was motivated by the fact that a) the replacement codon still has an accetably high frequency of usage in human_high.cod and b) the average overall human codon usage in CUTG for the replacement codon is nearly as high as the most frequent codon. In the following cycle analysis of the shifted window is then applied to a sequence containing the replacements of the previous cycle.

The threshold 70% was determined empirically by compromising between an overall reduction in GC content and maintenance of a high codon optimization for the individual amino acids. As in step 1) the precise replacement strategy (choice of amino acids and GC content threshold value) will again depend on the amino acid sequence encoded by the nucleotide sequence under analysis and will have to be determined empirically.

Step 3) The sequence generated by steps 1) and 2) was then manually edited and additional codons were changed according to the following criteria:

Regions still having a GC content higher than 70% over a window of 21 codons were examined manually and a few codons were replaced again following the scheme given in Table 5.

Subsequent steps were performed to provide for useful restriction sites, remove possible open reading frames on the complementary strand, to add homologous recombinant regions, to add a Kozac signal, and to add a terminator. These steps are numbered 4-7

Step 4) The sequence generated in step 3 was examined for the absence of certain restriction sites (BgIII, PmeI and XbaI) and presence of only 1 StuI site to allow a subsequent cloning strategy using a subset of restriction enzymes. Two sites (one for BgIII and one for StuI) were removed from the sequence by replacing codons that were part of the respective recognition sites.

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Step 5) The sequence generated by steps 1) through 4) was then modified according to allow subsequent generation of a modified NSOPTmut sequence (by homologous recombination). In the sequence obtained from steps 1) through 4) the segment comprising base 3556 to 3755 and the segment comprising base 4456 to 4656 were replaced by the corresponding segments from NSOPTmut. The segment comprising bases 3556 to 4656 of SEQ. ID. NO. 10 can be used to replace the problematic region in NSOPTmut (around position 3900) by homologous recombination thus creating the variant of NSOPTmut having the sequence of SEQ. ID. NO. 11.

Step 6) Analysis of the sequence generated through steps 1) to 5) revealed a potential open reading frame spanning nearly the complete fragment on the complementary strand. Removal of all codons CTA and TTA (Leu) and TCA (Ser) from the sense strand effectively removed all stop codons in one of the reading frames on the complementary strand. Although the likelyhood for transcription of this complementary strand open reading frame and subsequent translation into protein is very small, in order to exclude a potential interference with the transcription and subsequent translation of the sequence encoded on the sense strand, TCA codons for Ser were introduced on the sense approximately every 500 bases. No changes were introduced in the segments introduced during step 5) to allow homologous recombination. The TCA codon for Ser was preferred over the CTA and TTA codons for Leu because of the higher relative frequency for TCA (0.05) as compared to CTA (0.02) and TTA (0.03) in human_high.cod. In addition, the average human codon usage from CUTG favored TCA (0.14 against 0.07 for CTA and TTA).

Step 7) In a final step GCCACC was added at the 5'end of the sequence to generate an optimized internal ribosome entry site (Kozak signal) and a TAAA stop sgnal was added at the 3'. To maintain the initiation of translation

properties of NSsuboptmut the first 8 codons of the coding region were kept identical to the NSOPTmut sequence. The resulting sequence was again checked for the absence of BglII, PmeI and XbaI recognition sites and the presence of only 1 StuI site.

The NSsuboptmut sequence (SEQ. ID. NO. 10) has an overall reduced

5 GC content (63.5%) as compared to NSOPTmut (70.3%) and maintains a well optimized level of codon usage optimization. Nucleotide sequence identity of NSsuboptmut is 77.2% with respect to NSmut.

Table 5: Definition of codon replacements performed during steps 1) and 2).

|--|

Amino Acid	Most frequent codon	Relative frequency	Reduction in GC content (bases)	Replacement codon	Relative frequency
Amino	Acids where the re	placement codon	reduces the codor	GC-content by 1	base
Ala	GCC	0.51	1	GCT	0.17
Arg	CGC	0.37	1	AGG	0.18
Asn	AAC	0.78	1	AAT	0.22
Asp	GAC	0.75	1	GAT	0.25
Cys	TGC	0.68	1	TGT	0.32
Glu	GAG	0.75	1	GAA	0.25
Gln	CAG	0.88	1	CAA	0.12
Gly	GGC	0.50	1	GGA	0.14
His	CAC	0.79	1	CAT	0.21
Ile	ATC	0.77	1	ATT	0.18
Lys	AAG	0.82	1	AAA	0.18
Phe	TTC	0.80	1	TTT	0.20
Pro	CCC	0.48	1	CCT	0.19
Ser	AGC	0.34	1	TCT	0.13
Thr	ACC	0.51	1	ACA	0.14
Туг	TAC	0.74	1	TAT	0.26
			alternative codon	<u> </u>	1
Met	ATG	1.00	0	ATG	1.00
Тгр	TGG	1.00	0	TGG	1.00

Amino Acid	s where the replaceme	nt codon has a very	low relative freq	uency. These amin	o acids were
	excl	uded from the repla	cement procedure	e	,
Leu	CTG	0.58	1	TTG	0.06
Val	GTG	0.64	1	GTT	0.07

Example 12: Virus Characterization

Adenovectors were characterized by: (a) measuring the physical particles/ml; (b) running a TaqMan PCR assay; and (c) checking protein expression after infection of HeLa cells.

a) Physical Particles Determination

CsCl purified virus was diluted 1/10 and 1/100 in 0.1% SDS PBS. As a control, buffer A105 was used. These dilutions were incubated 10 minutes at 55° C. After spinning the tubes briefly, O.D. at 260 nm was measured. The amount of viral particles was calculated as follows: 1 OD 260 nm = 1.1×10^{12} physical particles/ml. The results were typically between 5×10^{11} and 1×10^{12} physical particles/ml.

b) TaqMan PCR Assay

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TaqMan PCR assay was used for adenovectors genome quantification (Q-PCR particles/ml). TaqMan PCR assay was performed using the ABI Prism 7700-sequence detector. The reaction was performed in a final 50 μ l volume in the presence of oligonucleotides (at final 200 nM) and probe (at final 200 μ M) specific for the adenoviral backbone. The virus was diluted 1/10 in 0.1% SDS PBS and incubated 10 minutes at 55°C. After spinning the tube briefly, serial 1/10 dilutions (in water) were prepared. 10 μ l the 10⁻³, 10⁻⁵ and 10⁻⁷ dilutions were used as templates in the PCR assay.

The amount of particles present in each sample was calculated on the basis of a standard curve run in the same experiment. Typically results were between 1×10^{12} and 3×10^{12} Q-PCR particles/ml.

c) Expression of HCV Non-Structural Proteins

Expression of HCV NS proteins was tested by infection of HeLa cells. Cells were plated the day before the infection at 1.5 X 10⁶ cells/dish (10 cm ø Petri dishes). Different amounts of CsCl purified virus corresponding to m.o.i. of 50, 250

and 1250 pp/cell were diluted in medium (FCS free) up to a final volume of 5 ml. The diluted virus was added on the cells and incubated for 1 hour at 37° C in a CO₂ incubator (gently mixing every 20 minutes). 5 ml of 5% HS-DMEM was added and the cells were incubated at 37° C for 48 hours.

Cell extracts were prepared in 1% Triton/TEN buffer. The extracts were run on 10% SDS-acrylamide gel, blotted on nitrocellulose and assayed with antibodies directed against NS3, NS5a and NS5b in order to check the correct polyprotein cleavage. Mock-infected cells were used as a negative control. Results from representative experiments testing the Ad5-NS, MRKAd5-NSmut, MRKAd6-NSmut and MRKAd6-NSOPTmut are shown in Figure 14.

Example 13: Mice Immunization with Adenovectors Encoding Different NS Cassettes

The adenovectors Ad5-NS, MRKAd5-NSmut, MRKAd6-NSmut and

MRKAd6-NSOPTmut were injected in C57Black6 mice strains to evaluate their

potential to elicit anti-HCV immune responses. Groups of animals (N=9-10) were
injected intramuscularly with 109 pp of CsCl purified virus. Each animal received two
doses at three weeks interval.

Humoral immune response against the NS3 protein was measured in post dose two sera from C57Black6 immunized mice by ELISA on bacterially expressed NS3 protease domain. Antibodies specific for the tested antigen were detected with geometric mean titers (GMT) ranging from 100 to 46000 (Tables 6, 7, 8 and 9).

Table 6: Ad5-NS

											GMT
Mice n.	1	2	3	4	5	6	7	8	9	10	
Titer	50	253	50	50	50	2257	504	50	50	50	108

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Table 7: Ad5-NSmut

											GMT
Mice n.	11	12	13	14	15	16	17	18	19	20	
Titer	3162	78850	87241	6796	12134	3340	18473	13093	76167	49593	23645

Table 8: MRKAd6-NSmut

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	-			***	··						GMT
Mice	21	22	23	24	25	26	27	28	29	30	
n. Titer	125626	39751	40187	65834	60619	69933	21555	49348	29290	26859	46461

Table 9: MRKAd6-NSOPTmut

,,,, <u>,,,,,,,,</u>								GMT
Mice n.	31	32	33	34	35	36	37	
Titer	25430	3657	893	175	10442	49540	173	2785

T cell response in C57Black6 mice was analyzed by the quantitative 10

ELISPOT assay measuring the number of IFNγ secreting T cells in response to five pools (named from F to L+M) of 20mer peptides overlapping by ten residues encompassing the NS3-NS5B sequence. Specific CD8+ response induced in C57Black6 mice was analyzed by the same assay using a 20mer peptide

encompassing a CD8+ epitope for C57Black6 mice (pep1480). Cells secreting IFNy in an antigen specific-manner were detected using a standard ELIspot assay.

Spleen cells, splenocytes and peptides were produced and treated as described in Example 3, supra. Representative data from groups of C57Black6 mice (N=9-10) immunized with two injections of 109 viral particles of vectors Ad5-NS,

MRKAd5-NSmut and MRKAd6-NSmut are shown in Figure 15. 20

Example 14: Immunization of Rhesus macaques with Adenovectors

Rhesus macaques (N=3-4) were immunized by intramuscular injection of CsCl purified Ad5-NS, MRKAd5-NSmut, MRKAd6-NSmut or MRKAd6-

NSOPTmut virus. Each animal received two doses of 10^{11} or 10^{10} vp in the deltoid muscle at 0, and 4 weeks.

CMI was measured at different time points by a) IFN- γ ELISPOT (see Example 3, supra), b) IFN- γ ICS and c) bulk CTL assays. These assays measure HCV antigen-specific CD8+ and CD4+ T lymphocyte responses, and can be used for a variety of mammals, such as humans, rhesus monkeys, mice, and rats.

The use of a specific peptide or a pool of peptides can simplify antigen presentation in CTL cytotoxicity assays, interferon-gamma ELISPOT assays and interferon-gamma intracellular staining assays. Peptides based on the amino acid sequence of various HCV proteins (core, E2, NS3, NS4A, NS4B, NS5a, NS5b) were prepared for use in these assays to measure immune responses in HCV DNA and adenovirus vector vaccinated rhesus monkeys, as well as in HCV-infected humans. The individual peptides are overlapping 20-mers, offset by 10 amino acids. Large pools of peptides can be used to detect an overall response to HCV proteins while smaller pools and individual peptides may be used to define the epitope specificity of a response.

IFN-γICS

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For IFN- γ ICS, 2 x 106 PBMC in 1 ml R10 (RPMI medium, supplemented with 10% FCS) were stimulated with peptide pool antigens. Final concentration of each peptide was 2 μ g/ml. Cells were incubated for 1 hour in a CO₂ incubator at 37°C and then Brefeldin A was added to a final concentration of 10 μ g /ml to inhibit the secretion of soluble cytokines. Cells were incubated for additional 14-16 hours at 37°C.

Stimulation was done in the presence of co-stimulatory antibodies: CD28 and CD49d (anti-humanCD28 BD340975 and anti-humanCD49d BD340976). After incubation, cells were stained with fluorochrome-conjugated antibodies for surface antigens: anti-CD3, anti-CD4, anti-CD8 (CD3-APC Biosource APS0301, CD4-PE BD345769, CD8-PerCP BD345774).

To detect intracellular cytokines, cells were treated with FACS permeabilization buffer 2 (BD340973), 2x final concentration. Once fixed and permeabilized, cells were incubated with an antibody against human IFN-γ, IFN-γFITC (Biosource AHC4338).

Cells were resuspended in 1% formaldehyde in PBS and analyzed at FACS within 24 hours. Four color FACS analysis was performed on a FACSCalibur

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instrument (Becton Dickinson) equipped with two lasers. Acquisition was done gating on the lymphocyte population in the Forward versus Side Scatter plot coupled with the CD3, CD8 positive populations. At least 30,000 events of the gate were taken. The positive cells are expressed as number of IFN-γ expressing cells over 10⁶ lymphocytes.

IFN-γ ELISPOT and IFN-γ ICS data from immunized monkeys after one or two injections of 10^{10} or 10^{11} vp of the different adenovectors are reported in Figures 16A-16D, 17A, and 17B.

Bulk CTL Assays 10

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A distinguishing effector function of T lymphocytes is the ability of subsets of this cell population to directly lyse cells exhibiting appropriate MHCassociated antigenic peptides. This cytotoxic activity is most often associated with CD8+ T lymphocytes.

PBMC samples were infected with recombinant vaccine viruses expressing HCV antigens in vitro for approximately 14 days to provide antigen restimulation and expansion of memory T cells. Cytotoxicity against autologous B cell lines treated with peptide antigen pools was tested.

The lytic function of the culture is measured as a percentage of specific lysis resulted from chromium released from target cells during 4 hours incubation with CTL effector cells. Specific cytotoxicity is measured and compared to irrelevant antigen or excipient-treated B cell lines. This assay is semi-quantitative and is the preferred means for determining whether CTL responses were elicited by the vaccine. Data after two injections from monkeys immunized with 1011 vp/dose with adenovectors Ad5-NS, MRKAd5-NSmut and MRKAd6-NSmut are reported in 25 Figures 18A-18F.

Other embodiments are within the following claims. While several embodiments have been shown and described, various modifications may be made without departing from the spirit and scope of the present invention.

WHAT IS CLAIMED IS:

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 A nucleic acid comprising a nucleotide sequence encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide substantially similar to SEQ ID NO: 1, provided that said polypeptide has sufficient protease activity to process itself to produce an NS5B protein and said NS5B protein is enzymatically inactive.

- 2. The nucleic acid of claim 1, wherein said nucleotide sequence is substantially similar to the coding sequence of SEQ ID NO: 2.
- 3. The nucleic acid of claim 1, wherein said nucleotide sequence encodes for the polypeptide of SEQ ID NO: 1.
- 4. The nucleic acid of claim 3, wherein said nucleotide sequence is the coding sequence of either SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.
 - 5. The nucleic acid of claim 3, wherein said nucleotide sequence is the coding sequence of either SEQ ID NO: 2 or SEQ ID NO: 3.
 - 6. The nucleic acid of any one of claims 1-5, wherein said nucleic acid is an expression vector capable of expressing said polypeptide from said nucleotide sequence in a human cell.
- 7. A nucleic acid comprising a gene expression cassette able to express a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide substantially similar to SEQ ID NO: 1 in a human cell, provided that said polypeptide can process itself to produce an NS5B protein and said NS5B protein is enzymatically inactive, said expression cassette comprising:
 - a) a promoter transcriptionally coupled to a nucleotide sequence encoding said polypeptide;
 - b) a 5' ribosome binding site functionally coupled to said nucleotide sequence,

c) a terminator joined to the 3' end of said nucleotide sequence, and
d) a 3' polyadenylation signal functionally coupled to said nucleotide sequence.

- 5 8. The nucleic acid of claim 7, wherein said nucleotide sequence is substantially similar to either SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.
- 9. The nucleic acid of claim 8, wherein said nucleic acid is a shuttle vector further comprising a selectable marker, an origin of replication, a first adenovirus homology region and a second adenovirus homology region flanking said expression cassette, wherein said first homology region has at least about 100 base pairs substantially homologous to at least right end of a wild-type adenovirus region from about base pairs 1-425, and said second homology region has at least about 100 base pairs substantially homologous to at least the left end of a wild-type adenovirus region from about base pairs 3511-5792 of Ad5 or corresponding region of another adenovirus.
- 10. The nucleic acid of claim 9, wherein said nucleotide sequence 20 encodes for a polypeptide of SEQ ID NO: 1.
 - 11. The nucleic acid of claim 9, wherein said nucleotide sequence is SEQ ID NO: 2.
- 25 12. The nucleic acid of claim 9, wherein said nucleotide sequence is either SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.
 - 13. The nucleic acid of claim 8, wherein said nucleic acid is a plasmid suitable for administration into a human and further comprises a prokaryotic origin of replication and a gene coding for a selectable marker.

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14. The nucleic acid of claim 13, wherein said nucleotide sequence encodes for a polypeptide of SEQ ID NO: 1.

15. The nucleic acid of claim 14, wherein said nucleotide sequence is the coding sequence of either SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.

- 5 16. The nucleic acid of claim 14, wherein said nucleotide sequence is the coding sequence of SEQ ID NO: 2 or SEQ ID NO: 3.
 - 17. The nucleic acid of claim 14, wherein said promoter is the human intermediate early cytomegalovirus promoter (intron A), said 5' ribosome binding site consists of SEQ ID NO: 12, and said 3' polyadenylation is the bovine growth hormone (BGH) polyadenylation signal.

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- 18. The nucleic acid of claim 8, wherein said nucleic acid is a adenovirus genome plasmid comprising a selectable marker, an origin of replication, and a recombinant adenovector genome containing an E1 deletion, an E3 deletion, and said expression cassette.
 - 19. The nucleic acid of claim 8, wherein said nucleic acid is a adenovirus genome plasmid comprising a selectable marker, an origin of replication, and
 - a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;
 - b) said gene expression cassette in a E1 parallel or E1 anti-parallel orientation joined to said first region;
- 25 c) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said expression cassette;
 - d) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;
 - e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to said third region; and

f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to said fourth region.

- 5 20. The nucleic acid of claim 19, wherein said first region corresponds to Ad5, said second region corresponds to Ad5, said third region corresponds to Ad5, said fourth region corresponds to Ad5, and said fifth region corresponds to Ad5.
- 10 21. The nucleic acid of claim 20, wherein said promoter is the human intermediate early cytomegalovirus promoter, said 5' ribosome binding site consists of SEQ ID NO: 12, and said 3' polyadenylation is the BGH polyadenylation signal.
- 15 22. The nucleic acid of claim 21, wherein said expression cassette is in an E1 anti parallel orientation and said nucleotide sequence is either SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.
- The nucleic acid of claim 19, wherein said first region
 corresponds to Ad5 or Ad6, said second region corresponds to Ad5 or Ad6, said third region corresponds to Ad6, said fourth region corresponds to Ad6, and said fifth region corresponds to Ad5 or Ad6.
- 24. The nucleic acid of claim 23, wherein said promoter is the human intermediate early cytomegalovirus promoter, said 5' ribosome binding site consists of SEQ ID NO: 12, and said 3' polyadenylation is the BGH polyadenylation signal.
- 25. The nucleic acid of claim 24, wherein said expression cassette
 30 is in an E1 anti parallel orientation and said nucleotide sequence is either SEQ ID NO:
 2, SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.
- 26. The nucleic acid of claim 24, wherein said expression cassette is in an E1 anti parallel orientation and said nucleotide sequence is either SEQ ID NO:
 2 or SEQ ID NO: 3.

27. The nucleic acid of claim 8, wherein said nucleic acid is a adenovirus genome plasmid comprising an origin of replication, a selectable marker, and:

a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;

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- b) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said first region;
- c) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;
- d) said gene expression cassette in a E3 parallel or E3 anti-parallel orientation joined to said third region;
- e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to said gene expression cassette; and
- f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to said fourth region.
- 28. The nucleic acid of claim 27, wherein said first region corresponds to Ad5, said second region corresponds to Ad5, said third region corresponds to Ad5, said fourth region corresponds to Ad5, and said fifth region corresponds to Ad5.
- 29. The nucleic acid of claim 28, wherein said promoter is the human intermediate early cytomegalovirus promoter, said 5' ribosome binding site consists of SEQ ID NO: 12, and said 3' polyadenylation is the BGH polyadenylation signal.
- 30. The nucleic acid of claim 27, wherein said first region corresponds to Ad5 or Ad6, said second region corresponds to Ad5 of Ad6, said third region corresponds to Ad6, said fourth region corresponds to Ad6, and said fifth region corresponds to Ad5 or Ad6.

31. The nucleic acid of claim 30, wherein said promoter is the human intermediate early cytomegalovirus promoter, said 5' ribosome binding site consists of SEQ ID NO: 12, and said 3' polyadenylation is the BGH polyadenylation signal.

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- 32. The nucleic acid of claim 8, wherein said nucleic acid is a adenovector consisting of a nucleotide sequence substantially similar to of SEQ ID NO. 4 or a derivative thereof, wherein said derivative thereof has the HCV polyprotein encoding sequence present in SEQ ID NO: 4 replaced with the HCV polyprotein encoding sequence of either SEQ ID NO: 3, SEQ ID NO: 10 or SEQ ID NO: 11.
- 33. The nucleic acid of claim 8, wherein said nucleic acid is an adenovector having an adenovector genome containing an E1 deletion, an E3 deletion, and said expression cassette
 - 34. The nucleic acid of claim 8, wherein said nucleic acid is an adenovector consisting of:
 - a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;
 - b) said gene expression cassette in a E1 parallel or E1 anti-parallel orientation joined to said first region;
 - c) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said expression cassette;
 - d) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;
 - e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to said third region; and
- f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to said fourth region.

35. The nucleic acid of claim 34, wherein said first region corresponds to Ad5, said second region corresponds to Ad5, said third region corresponds to Ad5, said fourth region corresponds to Ad5, and said fifth region corresponds to Ad5.

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- 36. The nucleic acid of claim 35, wherein said promoter is the human intermediate early cytomegalovirus promoter, said 5' ribosome binding site consists of SEQ ID NO: 12, and said 3' polyadenylation is the BGH polyadenylation signal.
- 37. The nucleic acid of claim 36, wherein said expression cassette is in an E1 anti parallel orientation and said nucleotide sequence is either SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.

38. The nucleic acid of claim 34, wherein said first region corresponds to Ad5 or Ad6, said second region corresponds to Ad5 or Ad6, said third region corresponds to Ad6, said fourth region corresponds to Ad6, and said fifth region corresponds to Ad5 or Ad6.

39. The nucleic acid of claim 37, where said promoter is the human intermediate early cytomegalovirus promoter, said 5' ribosome binding site consists of SEQ ID NO: 12, and said 3' polyadenylation is the BGH polyadenylation signal.

- 40. The nucleic acid of claim 39, wherein said expression cassette is in an E1 anti parallel orientation and said nucleotide sequence is SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 10, or SEQ ID NO: 11.
- 41. The nucleic acid of claim 39, wherein said expression cassette is in an E1 anti parallel orientation and said nucleotide sequence is SEQ ID NO: 2 or SEQ ID NO: 3.
 - 42. The nucleic acid of claim 8, wherein said nucleic acid is an adenovector consisting of:

a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;

b) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said first region;

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- c) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;
- d) said gene expression cassette in a E3 parallel or E3 anti-parallel
 orientation joined to said third region;
 - e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to said gene expression cassette; and
- f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to said fourth region.
 - 43. The nucleic acid of claim 42, wherein said first region corresponds to Ad5, said second region corresponds to Ad5, said third region corresponds to Ad5, said fourth region corresponds to Ad5, and said fifth region corresponds to Ad5.
 - 44. The nucleic acid of claim 42, wherein said first region corresponds to Ad5 or Ad6, said second region corresponds to Ad5 or Ad6, said third region corresponds to Ad6, said fourth region corresponds to Ad6, and said fifth region corresponds to Ad5 or Ad6.
- ID NO. 4 or a derivative thereof, wherein said derivative thereof has the HCV polyprotein encoding sequence present in SEQ ID NO: 4 replaced with the HCV polyprotein encoding sequence of either SEQ ID NO: 3, SEQ ID NO: 10 or SEQ ID NO: 11.
 - 46. An adenovector produced by a process comprising the steps of:

a) producing an adenovirus genome plasmid by homologous recombination between the shuttle vector of claim 9 and a nucleic acid comprising; a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;

a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said first region;

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a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;

a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to said third region; and

a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to said fourth region; and

- b) rescuing said adenovector from said adenovirus plasmid.
- 47. A cultured recombinant cell comprising the nucleic acid of 20 claim 6.
 - 48. A cultured recombinant cell comprising the nucleic acid of any one of claims 9-46.
- 25 49. A method of making an adenovector comprising the steps of:
 - a) producing an adenovirus genome plasmid comprising a gene expression cassette by homologous recombination between the nucleic acid of claim 9 and a nucleic acid comprising;

a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;

a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said first region;

a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;

- a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to said third region; and
 - a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to the fourth region; and
 - b) rescuing said recombinant adenovirus from said recombinant adenovirus plasmid.
 - 50. A pharmaceutical composition comprising the nucleic acid of any one of claims 13-17 and 32-46 and pharmaceutically acceptable carrier.
 - 51. A method of treating a patient comprising the step of administering to said patient an effective amount of the nucleic acid of any one of claims 13-17 and 32-46.
- 20 52. The method of claim 51, wherein said patient is a human.
 - 53. The method of claim 52, wherein said patient is not infected with HCV.
- The method of claim 52, wherein said patient is infected with HCV.
 - 55. A recombinant nucleic acid comprising one or more Ad6 regions and a region not present in Ad6, wherein at least one Ad6 region is selected from the group consisting of: E1A, E1B, E2B, E2A, E4, L1, L2, L4, and L5.
 - 56. The recombinant nucleic acid of claim 55, wherein said region not present in Ad6, is an expression cassette coding for a polypeptide not found in Ad6.

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57. The recombinant nucleic acid of claim 56, wherein said recombinant nucleic acid is an adenovirus vector defective in at least E1 that is able to replicate when E1 is supplied *in trans*.

- 5 58. The recombinant nucleic acid of claim 57, wherein said vector consists of:
 - a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;
 - b) said gene expression cassette in an E1 parallel or E1 antiparallel orientation joined to said first region;
 - c) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said gene expression cassette;
 - d) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;
 - e) an optionally present fourth region from about base pair 28134 to about base pair 30817 corresponding to Ad5, or from about base pair 28157 to about 30789 corresponding to Ad6, joined to said third region;
 - f) a fifth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, wherein said fifth region is joined to said fourth region if said fourth region is present, or said fifth is joined to said third region if said fourth region is not present; and
 - g) a sixth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to said fourth region;

provided that at least one of said second, third, and fifth regions is from Ad6.

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- 59. The recombinant nucleic acid of claim 57, wherein said vector consists of:
- a) a first adenovirus region from about base pair 1 to about base pair 450 corresponding to either Ad5 or Ad6;

b) a second adenovirus region from about base pair 3511 to about base pair 5548 corresponding to Ad5 or from about base pair 3508 to about base pair 5541 corresponding to Ad6, joined to said first region;

- c) a third adenovirus region from about base pair 5549 to about base pair 28133 corresponding to Ad5 or from about base pair 5542 to about base pair 28156 corresponding to Ad6, joined to said second region;
 - d) said gene expression cassette in a E3 parallel or E3 anti-parallel orientation joined to said third region;
- e) a fourth adenovirus region from about base pair 30818 to about base pair 33966 corresponding to Ad5 or from about base pair 30789 to about base pair 33784 corresponding to Ad6, joined to said gene expression cassette; and
 - f) a fifth adenovirus region from about base pair 33967 to about base pair 35935 corresponding to Ad5 or from about base pair 33785 to about base pair 35759 corresponding to Ad6, joined to said fourth region;
- provided that at least one of said second, third, and fourth regions is from Ad6.

1	MAPITAYSQQ	TRGLLGCIIT	SLTGRDKNQV	EGEVQVVSTA	TQSFLATCVN
51	GVCWTVYHGA	GSKTLAGPKG	PITQMYTNVD	QDLVGWQAPP	GARSLTPCTC
101	GSSDLYLVTR	HADVIPVRRR	GDSRGSLLSP	RPVSYLKGSS	GGPLLCPSGH
151	AVGIFRAAVC	TRGVAKAVDF	VPVESMETTM	RSPVFTDNSS	PPAVPQSFQV
201	AHLHAPTGSG	KSTKVPAAYA	AQGYKVLVLN	PSVAATLGFG	AYMSKAHGID
251	PNIRTGVRTI	TTGAPVTYST	YGKFLADGGC	SGGAYDIIIC	DECHSTDSTT
301	ILGIGTVLDQ	AETAGARLVV	LATATPPGSV	TVPHPNIEEV	ALSNTGEIPF
351	YGKAIPIEAI	RGGRHLIFCH	SKKKCDELAA	KLSGLGINAV	AYYRGLDVSV
401	IPTIGDVVVV	ATDALMTGYT	GDFDSVIDCN	TCVTQTVDFS	LDPTFTIETT
451	TVPQDAVSRS	QRRGRTGRGR	RGIYRFVTPG	ERPSGMFDSS	VLCECYDAGC
501	AWYELTPAET	SVRLRAYLNT	PGLPVCQDHL	EFWESVFTGL	THIDAHFLSQ
551	${\tt TKQAGDNFPY}$	LVAYQATVCA	RAQAPPPSWD	QMWKCLIRLK	PTLHGPTPLL
601	YRLGAVQNEV	TLTHPITKYI	MACMSADLEV	VTSTWVLVGG	VLAALAAYCL
651	TTGSVVIVGR	IILSGRPAIV	PDREFLYQEF	DEMEECASHL	PYIEQGMQLA
701	EQFKQKALGL	LQTATKQAEA	AAPVVESKWR	ALETFWAKHM	WNFISGIQYL
75 1	AGLSTLPGNP	AIASLMAFTA	SITSPLTTQS	TLLFNILGGW	VAAQLAPPSA
801	ASAFVGAGIA	GAAVGSIGLG	KVLVDILAGY	GAGVAGALVA	FKVMSGEMPS
851	TEDLVNLLPA	ILSPGALVVG	VVCAAILRRH	VGPGEGAVQW	MNRLIAFASR
901	GNHVSPTHYV	PESDAAARVT	QILSSLTITQ	LLKRLHQWIN	EDCSTPCSGS
951	WLRDVWDWIC	TVLTDFKTWL	QSKLLPQLPG	VPFFSCQRGY	KGVWRGDGIM
1001	QTTCPCGAQI	TGHVKNGSMR	IVGPKTCSNT	WHGTFPINAY	TTGPCTPSPA
1051	PNYSRALWRV	AAEEYVEVTR	VGDFHYVTGM	TTDNVKCPCQ	VPAPEFFTEV
1101	DGVRLHRYAP	ACRPLLREEV	${\tt TFQVGLNQYL}$	VGSQLPCEPE	PDVAVLTSML
1151	TDPSHITAET	AKRRLARGSP	PSLASSSASQ	LSAPSLKATC	TTHHVSPDAD
1201	LIEANLLWRQ	EMGGNITRVE	SENKVVVLDS	FDPLRAEEDE	REVSVPAEIL
1251	RKSKKFPAAM	PIWARPDYNP	PLLESWKDPD	YVPPVVHGCP	LPPIKAPPIP
1301	PPRRKRTVVL	TESSVSSALA	ELATKTFGSS	ESSAVDSGTA	TALPDQASDD
1351	GDKGSDVESY	SSMPPLEGEP	GDPDLSDGSW	STVSEEASED	VVCCSMSYTW
1401	TGALITPCAA	EESKLPINAL	SNSLLRHHNM	VYATTSRSAG	LRQKKVTFDR
1451	LQVLDDHYRD	VLKEMKAKAS	TVKAKLLSVE	EACKLTPPHS	AKSKFGYGAK
1501	DVRNLSSKAV	NHIHSVWKDL	LEDTVTPIDT	TIMAKNEVFC	VQPEKGGRKP
1551	ARLIVFPDLG	VRVCEKMALY	DVVSTLPQVV	MGSSYGFQYS	PGQRVEFLVN
1601	TWKSKKNPMG	FSYDTRCFDS	TVTENDIRVE	ESIYQCCDLA	PEARQAIKSL
1651	TERLYIGGPL	TNSKGQNCGY	RRCRASGVLT	TSCGNTLTCY	LKASAACRAA

FIG. 1A

1701	KLQDCTMLVN	AAGLVVICES	AGTQEDAASL	RVFTEAMTRY	SAPPGDPPQP
1751	EYDLELITSC	SSNVSVAHDA	SGKRVYYLTR	DPTTPLARAA	WETARHTPVN
1801	SWLGNIIMYA	PTLWARMILM	THFFSILLAQ	EQLEKALDCQ	IYGACYSIEP
1851	LDLPQIIERL	HGLSAFSLHS	YSPGEINRVA	SCLRKLGVPP	LRVWRHRARS
1901	VRARLLSQGG	RAATCGKYLF	NWAVKTKLKL	TPIPAASQLD	LSGWFVAGYS
1951	GGDIYHSLSR	ARPRWFMLCL	LLLSVGVGIY	LLPNR	

1	GCCACCATGG	CGCCCATCAC	GGCCTACTCC	CAACAGACGC	GGGGCCTACT
51	TGGTTGCATC	ATCACTAGCC	TTACAGGCCG	GGACAAGAAC	CAGGTCGAGG
101	GAGAGGTTCA	GGTGGTTTCC	ACCGCAACAC	AATCCTTCCT	GGCGACCTGC
151	GTCAACGGCG	TGTGTTGGAC	CGTTTACCAT	GGTGCTGGCT	CAAAGACCTT
201	AGCCGGCCCA	AAGGGCCAA	TCACCCAGAT	GTACACTAAT	GTGGACCAGG
251	ACCTCGTCGG	CTGGCAGGCG	CCCCCGGGG	CGCGTTCCTT	GACACCATGC
301	ACCTGTGGCA	GCTCAGACCT	TTACTTGGTC	ACGAGACATG	CTGACGTCAT
351	TCCGGTGCGC	CGGCGGGCG	ACAGTAGGGG	GAGCCTGCTC	TCCCCCAGGC
401	CTGTCTCCTA	CTTGAAGGGC	TCTTCGGGTG	GTCCACTGCT	CTGCCCTTCG
451	GGGCACGCTG	TGGGCATCTT	CCGGGCTGCC	GTATGCACCC	GGGGGGTTGC
501	GAAGGCGGTG	GACTTTGTGC	CCGTAGAGTC	CATGGAAACT	ACTATGCGGT
551	CTCCGGTCTT	CACGGACAAC	TCATCCCCCC	CGGCCGTACC	GCAGTCATTT
601	CAAGTGGCCC	ACCTACACGC	TCCCACTGGC	AGCGGCAAGA	GTACTAAAGT
651	GCCGGCTGCA	TATGCAGCCC	AAGGGTACAA	GGTGCTCGTC	CTCAATCCGT
701	CCGTTGCCGC	TACCTTAGGG	TTTGGGGCGT	ATATGTCTAA	GGCACACGGT
751	ATTGACCCCA	ACATCAGAAC	TGGGGTAAGG	ACCATTACCA	CAGGCGCCCC
801	CGTCACATAC	TCTACCTATG	GCAAGTTTCT	TGCCGATGGT	GGTTGCTCTG
851	GGGGCGCTTA	TGACATCATA	ATATGTGATG	AGTGCCATTC	AACTGACTCG
901	ACTACAATCT	TGGGCATCGG	CACAGTCCTG	GACCAAGCGG	AGACGGCTGG
951	AGCGCGGCTT	GTCGTGCTCG	CCACCGCTAC	GCCTCCGGGA	TCGGTCACCG
1001	TGCCACACCC	AAACATCGAG	GAGGTGGCCC	TGTCTAATAC	TGGAGAGATC
1051	CCCTTCTATG	GCAAAGCCAT	CCCCATTGAA	GCCATCAGGG	GGGGAAGGCA
1101	TCTCATTTTC	TGTCATTCCA	AGAAGAAGTG	CGACGAGCTC	GCCGCAAAGC
1151	TGTCAGGCCT	CGGAATCAAC	GCTGTGGCGT	ATTACCGGGG	GCTCGATGTG
1201	TCCGTCATAC	CAACTATCGG	AGACGTCGTT	GTCGTGGCAA	CAGACGCTCT
1251	GATGACGGGC	TATACGGGCG	ACTTTGACTC	AGTGATCGAC	TGTAACACAT
1301	GTGTCACCCA	GACAGTCGAC	TTCAGCTTGG	ATCCCACCTT	CACCATTGAG
1351	ACGACGACCG	TGCCTCAAGA	CGCAGTGTCG	CGCTCGCAGC	GGCGGGGTAG
1401	GACTGGCAGG	GGTAGGAGAG	GCATCTACAG	GTTTGTGACT	CCGGGAGAAC
1451	GGCCCTCGGG	CATGTTCGAT	TCCTCGGTCC	TGTGTGAGTG	CTATGACGCG
1501	GGCTGTGCTT	GGTACGAGCT	CACCCCCGCC	GAGACCTCGG	TTAGGTTGCG
1551	GGCCTACCTG	AACACACCAG	GGTTGCCCGT	TTGCCAGGAC	CACCTGGAGT
1601	TCTGGGAGAG	TGTCTTCACA	GGCCTCACCC	ACATAGATGC	ACACTTCTTG
1651	TCCCAGACCA	AGCAGGCAGG	AGACAACTTC	CCCTACCTGG	TAGCATACCA

FIG. 2A

					maaa.ma
1701		TGCGCCAGGG			
1751		TCTCATACGG			
1801		GGCTGGGAGC			
1851		TACATCATGG			
1901	CTAGCACCTG	GGTGCTGGTG	GGCGGAGTCC	TTGCAGCTCT	GGCCGCGTAT
1951	TGCCTGACAA	CAGGCAGTGT	GGTCATTGTG	GGTAGGATTA	TCTTGTCCGG
2001	GAGGCCGGCT	ATTGTTCCCG	ACAGGGAGTT	TCTCTACCAG	GAGTTCGATG
2051	AAATGGAAGA	GTGCGCCTCG	CACCTCCCTT	ACATCGAGCA	GGGAATGCAG
2101	CTCGCCGAGC	AATTCAAGCA	GAAAGCGCTC	GGGTTACTGC	AAACAGCCAC
2151	CAAACAAGCG	GAGGCTGCTG	CTCCCGTGGT	GGAGTCCAAG	TGGCGAGCCC
2201	TTGAGACATT	CTGGGCGAAG	CACATGTGGA	ATTTCATCAG	CGGGATACAG
2251	TACTTAGCAG	GCTTATCCAC	TCTGCCTGGG	AACCCCGCAA	TAGCATCATT
2301	GATGGCATTC	ACAGCCTCTA	TCACCAGCCC	GCTCACCACC	CAAAGTACCC
2351	TCCTGTTTAA	CATCTTGGGG	GGGTGGGTGG	CTGCCCAACT	CGCCCCCCC
2401	AGCGCCGCTT	CGGCTTTCGT	GGGCGCCGGC	ATCGCCGGTG	CGGCTGTTGG
2451	CAGCATAGGC	CTTGGGAAGG	TGCTTGTGGA	CATTCTGGCG	GGTTATGGAG
2501	CAGGAGTGGC	CGGCGCGCTC	GTGGCCTTCA	AGGTCATGAG	CGGCGAGATG
2551	CCCTCCACCG	AGGACCTGGT	CAATCTACTT	CCTGCCATCC	TCTCTCCTGG
2601	CGCCCTGGTC	GTCGGGGTCG	TGTGTGCAGC	AATACTGCGT	CGACACGTGG
2651	GTCCGGGAGA	GGGGGCTGTG	CAGTGGATGA	ACCGGCTGAT	AGCGTTCGCC
2701	TCGCGGGGTA	ATCATGTTTC	CCCCACGCAC	TATGTGCCTG	AGAGCGACGC
2751	CGCAGCGCGT	GTTACTCAGA	TCCTCTCCAG	CCTTACCATC	ACTCAGCTGC
2801	TGAAAAGGCT	CCACCAGTGG	ATTAATGAAG	ACTGCTCCAC	ACCGTGTTCC
2851	GGCTCGTGGC	TAAGGGATGT	TTGGGACTGG	ATATGCACGG	TGTTGACTGA
2901	CTTCAAGACC	TGGCTCCAGT	CCAAGCTCCT	GCCGCAGCTA	CCGGGAGTCC
2951	CTTTTTTCTC	GTGCCAACGC	GGGTACAAGG	GAGTCTGGCG	GGGAGACGGC
3001	ATCATGCAAA	CCACCTGCCC	ATGTGGAGCA	CAGATCACCG	GACATGTCAA
3051	AAACGGTTCC	ATGAGGATCG	TCGGGCCTAA	GACCTGCAGC	AACACGTGGC
3101	ATGGAACATT	CCCCATCAAC	GCATACACCA	CGGGCCCCTG	CACACCCTCT
3151	CCAGCGCCAA	ACTATTCTAG	GGCGCTGTGG	CGGGTGGCCG	CTGAGGAGTA
3201	CGTGGAGGTC	ACGCGGGTGG	GGGATTTCCA	CTACGTGACG	GGCATGACCA
3251	CTGACAACGT	AAAGTGCCCA	TGCCAGGTTC	CGGCTCCTGA	ATTCTTCACG
3301	GAGGTGGACG	GAGTGCGGTT	GCACAGGTAC	GCTCCGGCGT	GCAGGCCTCT
3351	CCTACGGGAG	GAGGTTACAT	TCCAGGTCGG	GCTCAACCAA	TACCTGGTTG

FIG. 2B

3401	GGTCACAGCT	ACCATGCGAG	CCCGAACCGG	ATGTAGCAGT	GCTCACTTCC
3451	ATGCTCACCG	ACCCCTCCCA	CATCACAGCA	GAAACGGCTA	AGCGTAGGTT
3501	GGCCAGGGGG	TCTCCCCCT	CCTTGGCCAG	CTCTTCAGCT	AGCCAGTTGT
3551	CTGCGCCTTC	CTTGAAGGCG	ACATGCACTA	CCCACCATGT	CTCTCCGGAC
3601	GCTGACCTCA	TCGAGGCCAA	CCTCCTGTGG	CGGCAGGAGA	TGGGCGGGAA
3651	CATCACCCGC	GTGGAGTCGG	AGAACAAGGT	GGTAGTCCTG	GACTCTTTCG
3701	ACCCGCTTCG	AGCGGAGGAG	GATGAGAGGG	AAGTATCCGT	TCCGGCGGAG
3751	ATCCTGCGGA	AATCCAAGAA	GTTCCCCGCA	GCGATGCCCA	TCTGGGCGCG
3801	CCCGGATTAC	AACCCTCCAC	TGTTAGAGTC	CTGGAAGGAC	CCGGACTACG
3851	TCCCTCCGGT	GGTGCACGGG	${\tt TGCCCGTTGC}$	CACCTATCAA	GGCCCCTCCA
3901	ATACCACCTC	CACGGAGAAA	GAGGACGGTT	GTCCTAACAG	AGTCCTCCGT
3951	GTCTTCTGCC	TTAGCGGAGC	TCGCTACTAA	GACCTTCGGC	AGCTCCGAAT
4001	CATCGGCCGT	CGACAGCGGC	ACGGCGACCG	CCCTTCCTGA	CCAGGCCTCC
4051	GACGACGGTG	ACAAAGGATC	CGACGTTGAG	TCGTACTCCT	CCATGCCCCC
4101	CCTTGAGGGG	GAACCGGGGG	ACCCCGATCT	CAGTGACGGG	TCTTGGTCTA
4151	CCGTGAGCGA	GGAAGCTAGT	GAGGATGTCG	TCTGCTGCTC	AATGTCCTAC
4201	ACATGGACAG	${\tt GCGCCTTGAT}$	CACGCCATGC	GCTGCGGAGG	AAAGCAAGCT
4251	GCCCATCAAC	GCGTTGAGCA	ACTCTTTGCT	GCGCCACCAT	AACATGGTTT
4301	ATGCCACAAC	ATCTCGCAGC	GCAGGCCTGC	GGCAGAAGAA	GGTCACCTTT
4351	GACAGACTGC	AAGTCCTGGA	CGACCACTAC	CGGGACGTGC	TCAAGGAGAT
4401	GAAGGCGAAG	GCGTCCACAG	TTAAGGCTAA	ACTCCTATCC	GTAGAGGAAG
4451	CCTGCAAGCT	GACGCCCCCA	CATTCGGCCA	AATCCAAGTT	TGGCTATGGG
4501	GCAAAGGACG	TCCGGAACCT	ATCCAGCAAG	GCCGTTAACC	ACATCCACTC
4551	CGTGTGGAAG	GACTTGCTGG	AAGACACTGT	GACACCAATT	GACACCACCA
4601	TCATGGCAAA	AAATGAGGTT	TTCTGTGTCC	AACCAGAGAA	AGGAGGCCGT
4651	AAGCCAGCCC	GCCTTATCGT	ATTCCCAGAT	CTGGGAGTCC	GTGTATGCGA
4701	GAAGATGGCC	CTCTATGATG	TGGTCTCCAC	CCTTCCTCAG	GTCGTGATGG
4751	GCTCCTCATA	CGGATTCCAG	TACTCTCCTG	GGCAGCGAGT	CGAGTTCCTG
4801	GTGAATACCT	GGAAATCAAA	GAAAAACCCC	ATGGGCTTTT	CATATGACAC
4851	TCGCTGTTTC	GACTCAACGG	TCACCGAGAA	CGACATCCGT	GTTGAGGAGT
4901	CAATTTACCA	ATGTTGTGAC	TTGGCCCCCG	AAGCCAGACA	GGCCATAAAA
4951	TCGCTCACAG	AGCGGCTTTA	TATCGGGGGT	CCTCTGACTA	ATTCAAAAGG
5001	GCAGAACTGC	GGTTATCGCC	GGTGCCGCGC	GAGCGGCGTG	CTGACGACTA
5051	GCTGCGGTAA	CACCCTCACA	TGTTACTTGA	AGGCCTCTGC	AGCCTGTCGA

5101	GCTGCGAAGC	TCCAGGACTG	CACGATGCTC	GTGAACGCCG	CCGGCCTTGT
5151	CGTTATCTGT	GAAAGCGCGG	GAACCCAAGA	GGACGCGGCG	AGCCTACGAG
5201	TCTTCACGGA	GGCTATGACT	AGGTACTCTG	CCCCCCCGG	GGACCCGCCC
5251	CAACCAGAAT	ACGACTTGGA	GCTGATAACA	${\tt TCATGTTCCT}$	CCAATGTGTC
5301	GGTCGCCCAC	GATGCATCAG	GCAAAAGGGT	GTACTACCTC	ACCCGTGATC
5351	CCACCACCCC	CCTCGCACGG	GCTGCGTGGG	AAACAGCTAG	ACACACTCCA
5401	GTTAACTCCT	GGCTAGGCAA	CATTATCATG	TATGCGCCCA	CTTTGTGGGC
5451	AAGGATGATT	CTGATGACTC	ACTTCTTCTC	CATCCTTCTA	GCACAGGAGC
5501	AACTTGAAAA	AGCCCTGGAC	TGCCAGATCT	ACGGGGCCTG	TTACTCCATT
5551	GAGCCACTTG	ACCTACCTCA	GATCATTGAA	CGACTCCATG	GCCTTAGCGC
5601	ATTTTCACTC	CATAGTTACT	CTCCAGGTGA	GATCAATAGG	GTGGCTTCAT
5651	GCCTCAGGAA	ACTTGGGGTA	CCACCCTTGC	GAGTCTGGAG	ACATCGGGCC
5701	AGGAGCGTCC	GCGCTAGGCT	ACTGTCCCAG	GGGGGGAGGG	CCGCCACTTG
5751	TGGCAAGTAC	CTCTTCAACT	GGGCAGTGAA	GACCAAACTC	AAACTCACTC
5801	CAATCCCGGC	TGCGTCCCAG	CTGGACTTGT	CCGGCTGGTT	CGTTGCTGGT
5851	TACAGCGGGG	GAGACATATA	TCACAGCCTG	TCTCGTGCCC	GACCCCGCTG
5901	GTTCATGCTG	TGCCTACTCC	TACTTTCTGT	AGGGGTAGGC	ATCTACCTGC
5951	TCCCCAACCG	ATAAA			

1	GCCACCATGG	CCCCCATCAC	CGCCTACAGC	CAGCAGACCC	GCGGCCTGCT
51	GGGCTGCATC	ATCACCAGCC	TGACCGGCCG	CGACAAGAAC	CAGGTGGAGG
101	GCGAGGTGCA	GGTGGTGAGC	ACCGCCACCC	AGAGCTTCCT	GGCCACCTGC
151	GTGAACGGCG	TGTGCTGGAC	CGTGTACCAC	GGCGCCGGCA	GCAAGACCCT
201	GGCCGGCCCC	AAGGGCCCCA	TCACCCAGAT	GTACACCAAC	GTGGACCAGG
251	ACCTGGTGGG	CTGGCAGGCC	CCCCCGGCG	CCCGCAGCCT	GACCCCCTGC
301	ACCTGCGGCA	GCAGCGACCT	GTACCTGGTG	ACCCGCCACG	CCGACGTGAT
351	CCCCGTGCGC	CGCCGCGGCG	ACAGCCGCGG	CAGCCTGCTG	AGCCCCCGCC
401	CCGTGAGCTA	CCTGAAGGGC	AGCAGCGGCG	GCCCCTGCT	GTGCCCCAGC
451	GGCCACGCCG	TGGGCATCTT	CCGCGCCGCC	GTGTGCACCC	GCGGCGTGGC
501	CAAGGCCGTG	GACTTCGTGC	CCGTGGAGAG	CATGGAGACC	ACCATGCGCA
551	GCCCCGTGTT	CACCGACAAC	AGCAGCCCCC	CCGCCGTGCC	CCAGAGCTTC
601	CAGGTGGCCC	ACCTGCACGC	CCCCACCGGC	AGCGGCAAGA	GCACCAAGGT
651	GCCCGCCGCC	TACGCCGCCC	AGGGCTACAA	GGTGCTGGTG	CTGAACCCCA
701	GCGTGGCCGC	CACCCTGGGC	${\tt TTCGGCGCCT}$	ACATGAGCAA	GGCCCACGGC
751	ATCGACCCCA	ACATCCGCAC	CGGCGTGCGC	ACCATCACCA	CCGGCGCCCC
801	CGTGACCTAC	AGCACCTACG	GCAAGTTCCT	GGCCGACGGC	GGCTGCAGCG
851	GCGGCGCCTA	CGACATCATC	ATCTGCGACG	AGTGCCACAG	CACCGACAGC
901	ACCACCATCC	TGGGCATCGG	CACCGTGCTG	GACCAGGCCG	AGACCGCCGG
951	CGCCCGCCTG	GTGGTGCTGG	CCACCGCCAC	CCCCCCGGC	AGCGTGACCG
1001	TGCCCCACCC	CAACATCGAG	GAGGTGGCCC	TGAGCAACAC	CGGCGAGATC
1051	CCCTTCTACG	GCAAGGCCAT	CCCCATCGAG	GCCATCCGCG	GCGGCCGCCA
1101	CCTGATCTTC	TGCCACAGCA	AGAAGAAGTG	CGACGAGCTG	GCCGCCAAGC
1151	TGAGCGGCCT	GGGCATCAAC	GCCGTGGCCT	ACTACCGCGG	CCTGGACGTG
1201	AGCGTGATCC	CCACCATCGG	CGACGTGGTG	GTGGTGGCCA	CCGACGCCCT
1251	GATGACCGGC	TACACCGGCG	ACTTCGACAG	CGTGATCGAC	TGCAACACCT
1301	GCGTGACCCA	GACCGTGGAC	TTCAGCCTGG	ACCCCACCTT	CACCATCGAG
1351	ACCACCACCG	TGCCCCAGGA	CGCCGTGAGC	CGCAGCCAGC	GCCGCGGCCG
1401	CACCGGCCGC	GGCCGCCGCG	GCATCTACCG	CTTCGTGACC	CCCGGCGAGC
1451	GCCCAGCGG	CATGTTCGAC	AGCAGCGTGC	TGTGCGAGTG	CTACGACGCC
1501	GGCTGCGCCT	GGTACGAGCT	GACCCCCGCC	GAGACCAGCG	TGCGCCTGCG
1551	CGCCTACCTG	AACACCCCCG	GCCTGCCCGT	GTGCCAGGAC	CACCTGGAGT
1601	TCTGGGAGAG	CGTGTTCACC	GGCCTGACCC	ACATCGACGC	CCACTTCCTG
1651	AGCCAGACCA	AGCAGGCCGG	CGACAACTTC	CCCTACCTGG	TGGCCTACCA

FIG. 3A

					maaa, aa, aa
1701			CCCAGGCCCC		
1751			CTGAAGCCCA		
1801			CGTGCAGAAC		
1851	CATCACCAAG	TACATCATGG	CCTGCATGAG	CGCCGACCTG	GAGGTGGTGA
1901	CCAGCACCTG	GGTGCTGGTG	GGCGGCGTGC	TGGCCGCCCT	GGCCGCCTAC
1951	TGCCTGACCA	CCGGCAGCGT	GGTGATCGTG	GGCCGCATCA	TCCTGAGCGG
2001	CCGCCCCGCC	ATCGTGCCCG	ACCGCGAGTT	CCTGTACCAG	GAGTTCGACG
2051	AGATGGAGGA	GTGCGCCAGC	CACCTGCCCT	ACATCGAGCA	GGGCATGCAG
2101	CTGGCCGAGC	AGTTCAAGCA	GAAGGCCCTG	GGCCTGCTGC	AGACCGCCAC
2151	CAAGCAGGCC	GAGGCCGCCG	$\mathtt{CCCCGTGGT}$	GGAGAGCAAG	TGGCGCGCCC
2201	TGGAGACCTT	${\tt CTGGGCCAAG}$	CACATGTGGA	ACTTCATCAG	CGGCATCCAG
2251	TACCTGGCCG	GCCTGAGCAC	CCTGCCCGGC	AACCCCGCCA	TCGCCAGCCT
2301	GATGGCCTTC	ACCGCCAGCA	TCACCAGCCC	CCTGACCACC	CAGAGCACCC
2351	TGCTGTTCAA	CATCCTGGGC	GGCTGGGTGG	CCGCCCAGCT	GGCCCCCCC
2401	AGCGCCGCCA	GCGCCTTCGT	GGGCGCCGGC	ATCGCCGGCG	CCGCCGTGGG
2451	CAGCATCGGC	CTGGGCAAGG	TGCTGGTGGA	CATCCTGGCC	GGCTACGGCG
2501	CCGGCGTGGC	CGGCGCCCTG	GTGGCCTTCA	AGGTGATGAG	CGGCGAGATG
2551	CCCAGCACCG	AGGACCTGGT	GAACCTGCTG	CCCGCCATCC	TGAGCCCCGG
2601	CGCCCTGGTG	GTGGGCGTGG	TGTGCGCCGC	CATCCTGCGC	CGCCACGTGG
2651	GCCCGGCGA	GGGCGCCGTG	CAGTGGATGA	ACCGCCTGAT	CGCCTTCGCC
2701	AGCCGCGGCA	ACCACGTGAG	CCCCACCCAC	TACGTGCCCG	AGAGCGACGC
2751	CGCCGCCCGC	GTGACCCAGA	TCCTGAGCAG	CCTGACCATC	ACCCAGCTGC
2801	TGAAGCGCCT	GCACCAGTGG	ATCAACGAGG	ACTGCAGCAC	CCCCTGCAGC
2851	GGCAGCTGGC	TGCGCGACGT	GTGGGACTGG	ATCTGCACCG	TGCTGACCGA
2901	CTTCAAGACC	TGGCTGCAGA	GCAAGCTGCT	GCCCCAGCTG	CCCGGCGTGC
2951	CCTTCTTCAG	CTGCCAGCGC	GGCTACAAGG	GCGTGTGGCG	CGGCGACGGC
3001	ATCATGCAGA	CCACCTGCCC	CTGCGGCGCC	CAGATCACCG	GCCACGTGAA
3051	GAACGGCAGC	ATGCGCATCG	TGGGCCCCAA	GACCTGCAGC	AACACCTGGC
3101	ACGGCACCTT	CCCCATCAAC	GCCTACACCA	CCGGCCCCTG	CACCCCAGC
3151	CCCGCCCCCA	ACTACAGCCG	CGCCCTGTGG	CGCGTGGCCG	CCGAGGAGTA
3201	CGTGGAGGTG	ACCCGCGTGG	GCGACTTCCA	CTACGTGACC	GGCATGACCA
3251	CCGACAACGT	GAAGTGCCCC	TGCCAGGTGC	CCGCCCCCGA	GTTCTTCACC
3301	GAGGTGGACG	GCGTGCGCCT	GCACCGCTAC	GCCCCGCCT	GCCGCCCCT
3351	GCTGCGCGAG	GAGGTGACCT	TCCAGGTGGG	CCTGAACCAG	TACCTGGTGG

3401	GCAGCCAGCT	GCCCTGCGAG	CCCGAGCCCG	ACGTGGCCGT	GCTGACCAGC
3451	ATGCTGACCG	ACCCCAGCCA	CATCACCGCC	GAGACCGCCA	AGCGCCGCCT
3501	GGCCCGCGGC	AGCCCCCCA	GCCTGGCCAG	CAGCAGCGCC	AGCCAGCTGA
3551	GCGCCCCAG	CCTGAAGGCC	ACCTGCACCA	CCCACCACGT	GAGCCCCGAC
3601	GCCGACCTGA	TCGAGGCCAA	CCTGCTGTGG	CGCCAGGAGA	TGGGCGGCAA
3651	CATCACCCGC	GTGGAGAGCG	AGAACAAGGT	GGTGGTGCTG	GACAGCTTCG
3701	ACCCCTGCG	CGCCGAGGAG	GACGAGCGCG	AGGTGAGCGT	GCCCGCCGAG
3751	ATCCTGCGCA	AGAGCAAGAA	GTTCCCCGCC	GCCATGCCCA	TCTGGGCCCG
3801	CCCCGACTAC	AACCCCCCC	TGCTGGAGAG	CTGGAAGGAC	CCCGACTACG
3851	TGCCCCCGT	GGTGCACGGC	TGCCCCCTGC	CCCCCATCAA	GCCCCCCC
3901	ATCCCCCCC	CCCGCCGCAA	GCGCACCGTG	GTGCTGACCG	AGAGCAGCGT
3951	GAGCAGCGCC	CTGGCCGAGC	TGGCCACCAA	GACCTTCGGC	AGCAGCGAGA
4001	GCAGCGCCGT	GGACAGCGGC	ACCGCCACCG	CCCTGCCCGA	CCAGGCCAGC
4051	GACGACGGCG	ACAAGGGCAG	CGACGTGGAG	AGCTACAGCA	GCATGCCCCC
4101	CCTGGAGGGC	GAGCCCGGCG	ACCCCGACCT	GAGCGACGGC	AGCTGGAGCA
4151	CCGTGAGCGA	GGAGGCCAGC	GAGGACGTGG	TGTGCTGCAG	CATGAGCTAC
4201	ACCTGGACCG	GCGCCCTGAT	CACCCCTGC	GCCGCCGAGG	AGAGCAAGCT
4251	GCCCATCAAC	GCCCTGAGCA	ACAGCCTGCT	GCGCCACCAC	AACATGGTGT
4301	ACGCCACCAC	CAGCCGCAGC	GCCGGCCTGC	GCCAGAAGAA	GGTGACCTTC
4351	GACCGCCTGC	AGGTGCTGGA	CGACCACTAC	CGCGACGTGC	TGAAGGAGAT
4401	GAAGGCCAAG	GCCAGCACCG	TGAAGGCCAA	GCTGCTGAGC	GTGGAGGAGG
4451	CCTGCAAGCT	GACCCCCCC	CACAGCGCCA	AGAGCAAGTT	CGGCTACGGC
4501	GCCAAGGACG	TGCGCAACCT	GAGCAGCAAG	GCCGTGAACC	ACATCCACAG
4551	CGTGTGGAAG	GACCTGCTGG	AGGACACCGT	GACCCCCATC	GACACCACCA
4601	TCATGGCCAA	GAACGAGGTG	TTCTGCGTGC	AGCCCGAGAA	GGGCGGCCGC
4651	AAGCCCGCCC	GCCTGATCGT	GTTCCCCGAC	CTGGGCGTGC	GCGTGTGCGA
4701	GAAGATGGCC	CTGTACGACG	TGGTGAGCAC	CCTGCCCCAG	GTGGTGATGG
4751	GCAGCAGCTA	CGGCTTCCAG	TACAGCCCCG	GCCAGCGCGT	GGAGTTCCTG
4801	GTGAACACCT	GGAAGAGCAA	GAAGAACCCC	ATGGGCTTCA	GCTACGACAC
4851	CCGCTGCTTC	GACAGCACCG	TGACCGAGAA	CGACATCCGC	GTGGAGGAGA
4901	GCATCTACCA	GTGCTGCGAC	CTGGCCCCCG	AGGCCCGCCA	GGCCATCAAG
4951	AGCCTGACCG	AGCGCCTGTA	CATCGGCGGC	CCCCTGACCA	ACAGCAAGGG
5001	CCAGAACTGC	GGCTACCGCC	GCTGCCGCGC	CAGCGGCGTG	CTGACCACCA
5051	GCTGCGGCAA	CACCCTGACC	TGCTACCTGA	AGGCCAGCGC	CGCCTGCCGC

5101	GCCGCCAAGC	TGCAGGACTG	CACCATGCTG	GTGAACGCCG	CCGGCCTGGT
5151	GGTGATCTGC	GAGAGCGCCG	GCACCCAGGA	GGACGCCGCC	AGCCTGCGCG
5201	TGTTCACCGA	GGCCATGACC	CGCTACAGCG	CCCCCCCGG	CGACCCCCCC
5251	CAGCCCGAGT	ACGACCTGGA	GCTGATCACC	AGCTGCAGCA	GCAACGTGAG
5301	CGTGGCCCAC	GACGCCAGCG	GCAAGCGCGT	GTACTACCTG	ACCCGCGACC
5351	CCACCACCCC	CCTGGCCCGC	GCCGCCTGGG	AGACCGCCCG	CCACACCCCC
5401	GTGAACAGCT	GGCTGGGCAA	CATCATCATG	TACGCCCCCA	CCCTGTGGGC
5451	CCGCATGATC	CTGATGACCC	ACTTCTTCAG	CATCCTGCTG	GCCCAGGAGC
5501	AGCTGGAGAA	GGCCCTGGAC	TGCCAGATCT	ACGGCGCCTG	CTACAGCATC
5551	GAGCCCCTGG	ACCTGCCCCA	GATCATCGAG	CGCCTGCACG	GCCTGAGCGC
5601	CTTCAGCCTG	CACAGCTACA	GCCCCGGCGA	GATCAACCGC	GTGGCCAGCT
5651	GCCTGCGCAA	GCTGGGCGTG	CCCCCCTGC	GCGTGTGGCG	CCACCGCGCC
5701	CGCAGCGTGC	GCGCCCGCCT	GCTGAGCCAG	GGCGGCCGCG	CCGCCACCTG
5751	CGGCAAGTAC	CTGTTCAACT	GGGCCGTGAA	GACCAAGCTG	AAGCTGACCC
5801	CCATCCCCGC	CGCCAGCCAG	CTGGACCTGA	GCGGCTGGTT	CGTGGCCGGC
5851	TACAGCGGCG	GCGACATCTA	CCACAGCCTG	AGCCGCGCCC	GCCCCCGCTG
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5951	TGCCCAACCG	CTAAA			



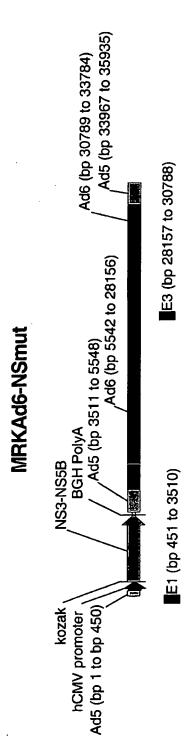


FIG. 4A

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241	taaatttggg	cotaaccoao	taagatttgg	ccattttcgc	gggaaaactg	aataagagga
301	agtgaaatct	gaataatttt	gtgttactca	tagogogtaa	tatttgtcta	gggccgcggg
361	gactttgacc	gtttacgtgg	agactcgccc	aggtgttttt	ctcaggtgtt	ttccgcgttc
121	cgggtcaaag	ttaacatttt	attattatag	acaaccacaa	tccattgcat	acgttgtatc
4Ω1	catatcataa	tatgtacatt	tatattggct	catotccaac	attaccgcca	tgttgacatt
5/1	gattattgac	tagttattaa	tagtaatcaa	ttacqqqqtc	attagttcat	agcccatata
601	tggagttccg	cottacataa	cttacggtaa	atggcccgcc	tggctgaccg	cccaacgacc
661	cccgcccatt	gacgtcaata	atgacgtatg	ttcccatagt	aacgccaata	gggactttcc
721	attgacgtca	atagatagaa	tatttacoot	aaactgccca	cttggcagta	catcaagtgt
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961	actcacgggg	atttccaaqt	ctccacccca	ttgacgtcaa	tgggagtttg	ttttggcacc
1021	aaaatcaacg	ggactttcca	aaatgtcgta	acaactccgc	cccattgacg	caaatgggcg
1021	gtaggcgtgt	acggt.gggag	otctatataa	gcagageteg	tttagtgaac	cgtcagatcg
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1321	gcaacacaat	cettectage	gacctgcgtc	aacggcgtgt	gttggaccgt	ttaccatggt
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1501	gaccaggacc	tcatcaacta	gcaggcgccc	cccggggcgc	gttccttgac	accatgcacc
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2101	tgctctgggg	gcgcttatga	catcataata	tgtgatgagt	gccattcaac	tgactcgact
2161	acaatcttgg	gcatcggcac	agtcctggac	caagcggaga	cggctggagc	gcggcttgtc
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	gagagggaag					
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24721	ccacgctgga	agccatgctc	agaaatgaca	ccaacgacca	gtcctttaat	gactaccttt
24781	ccgccgccaa	catgctatat	cccatacccg	ccaacgccac	caacgtgccc	atctccatcc
24841	catcgcgcaa	ctgggcagca	tttcgcggtt	gggccttcac	acgcttgaag	acaaaggaaa
24901	cccttccct	gggatcaggc	tacgaccctt	actacaccta	ctctggctcc	ataccatacc
24961	ttgacggaac	cttctatctt	aatcacacct	ttaagaaggt	ggccattact	tttgactctt
25021	ctgttagctg	gccgggcaac	gaccgcctgc	ttactcccaa	tgagtttgag	attaagcgct
25081	cagttgacgg	ggagggctat	aacgtagctc	agtgcaacat	gacaaaggac	tggttcctag
25141	tgcagatgtt	ggccaactac	aatattggct	accagggctt	ctacattcca	gaaagctaca
25201	aagaccgcat	gtactcgttc	ttcagaaact	tccagcccat	gagccggcaa	gtggtggacg
25261	atactaaata	caaagattat	cagcaggttg	gaattatcca	ccagcataac	aactcaggct
25321	tcgtaggcta	cctcqctccc	accatgcgcg	agggacaagc	ttaccccgct	aatgttccct
25381	acccactaat	aggcaaaacc	gcggttgata	gtattaccca	gaaaaagttt	ctttgcgacc
25441	gcaccctgtg	gcgcatcccc	ttctccagta	actttatgtc	catgggtgcg	ctcacagacc
25501	tgggccaaaa	ccttctctac	gcaaactccg	cccacgcgct	agacatgacc	tttgaggtgg
25561	atcccatgga	cgagcccacc	cttctttatg	ttttgtttga	agtctttgac	gtggtccgtg
25621	tgcaccagcc	gcaccgcggc	gtcatcgaga	ccgtgtacct	gcgcacgccc	ttctcggccg
25681	gcaacgccac	aacataaaga	agcaagcaac	atcaacaaca	gctgccgcca	tgggctccag
25741	tgagcaggaa	ctgaaagcca	ttgtcaaaga	tcttggttgt	gggccatatt	ttttgggcac
25801	ctatgacaag	cacttcccaq	gctttgtttc	cccacacaag	ctcgcctgcg	ccatagttaa
25861	cacggccggt	cacaagacta	ggggcgtaca	ctggatggcc	tttgcctgga	accegegete
25921	aaaaacatgc	tacctctttq	agccctttgg	cttttctgac	caacgtctca	agcaggttta
25981	ccagtttgag	tacgagtcac	tcctgcgccg	tagcgccatt	gcctcttccc	ccgaccgctg
26041	tataacgctg	gaaaagtcca	cccaaagcgt	gcaggggccc	aactcggccg	cctgtggcct
26101	attctgctgc	atgtttctcc	acgcctttgc	caactggccc	caaactccca	tggatcacaa
26161	cccaccatg	aaccttatta	ccggggtacc	caactccatg	cttaacagtc	cccaggtaca
26221	gcccaccctg	cgccgcaacc	aggaacagct	ctacagcttc	ctggagcgcc	actcgcccta
26281	cttccqcaqc	cacagtgcgc	aaattaggag	cgccacttct	ttttgtcact	tgaaaaacat
26341	gtaaaaataa	tgtactagga	gacactttca	ataaaggcaa	atgtttttat	ttgtacactc
26401	tcgggtgatt	atttacccc	accettgeeg	tctgcgccgt	ttaaaaatca	aaggggttct
	222-3	=		-		

FIG. 41

26461	accacacate	gctatgcgcc	actggcaggg	acacgttgcg	atactggtgt	ttagtgctcc
26521	acttaaactc	aggcacaacc	atccgcggca	gctcggtgaa	gttttcactc	cacaggctgc
26581	gcaccatcac	caacgcgttt	agcaggtcgg	gcgccgatat	cttgaagtcg	cagttggggc
26361	staggesta	cgcgcgcgag	rtgcgataca	cagggttaca	gcactggaac	actatcagcg
26701	ccacataata	cacgctggcc	agcacgctct	totcogagat	cagatccgcg	tccaggtcct
20/01	ccgggrggrg	cagggcgaac	ggagtcaact	ttggtagctg	ccttcccaaa	aagggtgcat
20/01	cegegreget	tgagttgcac	tracaccata	gtggcatcag	aaggtgaccg	tgcccagtct
20021	geeeayyeer	atacagcgcc	tgcatgaaag	ccttgatctg	cttaaaaqcc	acctgagect
20001	ttacacetta	agagaagaac	atoccocaao	acttoccooa	aaactgattg	gccggacagg
20341	coordinate	cacgcagcac	cttacatcaa	tottogagat	ctgcaccaca	tttcggcccc
27001	eegegteatg	cacgatettg	accttactag	actoctcctt	cadcacacac	tgcccgtttt
27001	accognicit	atccatttca	atcacatact	ccttatttat	cataatoctc	ccgtgtagac
2/121	egetegteac	gccttcgatc	trancoranc	ggtgcagcca	caacgcgcag	cccqtqqqct
2/181	acttaagete	gtaggttacc	tctgcgcago	actocagota	cacctacaaa	aatcgcccca
2/241	egeggegeee	aaaggtcttg	ttactaataa	aggreagetg	caacccgcgg	tactcctcat
2/301	teategreae	cttgcatacg	accaccadaa	cttccacttq	gtcaggcagt	agcttgaagt
2736I	ttagccaggu	atcgttatcc	acataataat	totccatcaa	cacacacaca	gcctccatgc
27421	ttgcctttag	atcgttatcc	atgraggagg	tcaccactt	tatcaccata	ctttcacttt
27481	ccttctccca	cgcagacacg	ateggeagge	ccaycygycc	accaccaca	actoggtcgt
27541	ccgcttcact	ggactcttcc	ELECCICAL	gtattegtat	accedegee	agcaccagta
27601	cttcattcag	ccgccgcacc	gtgegettae	catattatat	ttcttcctca	ctatccacaa
27661	ggttgctgaa	acccaccatt	tgtagcgcca	taccecce	acacttettt	ttctttttaa
27721	tcacctctgg	ggatggcggg	egeteggget	rgggagaggg	actagatata	cacaacacca
27781	acgcaatggc	caaatccgcc	gtcgaggtcg	argactaga	acceptant	agccgctttt
27841	gcgcatcttg	tgacgagtct	tettegteet	cygactcyay	acyccycccc	tccatcatta
27901	ttgggggcgc	gcggggaggc	ggcggcgacg	gcgacgggga	ttagaggtac	tectetteee
27961	gtggacgtcg	cgccgcaccg	egteegeget	egggggtggt		22222222
28021	gactggccat	ttccttctcc	tataggcaga	aaaagatcat	ggagtcagtc	gagaaggagg
28081	acagcctaac	cgccccttt	gagttcgcca	ccaccgcctc	cacegatgee	gccaacgcgc
28141	ctaccacctt	ccccgtcgag	gcacccccgc	ttgaggagga	ggaagtgatt	accyaycayy
28201	acccaggttt	tgtaagcgaa	gacgacgaag	atcgctcagt	accaacagag	gataaaaagc
28261	aagaccagga	cgacgcagag	gcaaacgagg	aacaagtcgg	geggggggae	caaaggcacg
28321	gcgactacct	agatgtggga	gacgacgtgc	tgttgaagca	tetgeagege	caguguguua
28381	ttatctgcga	cgcgttgcaa	gagcgcagcg	atgtgcccct	cgccatageg	gatgteagee
28441	ttgcctacga	acgccacctg	ttctcaccgc	gcgtaccccc	caaacgccaa	gaaaacgyca
28501	catgcgagcc	caacccgcgc	ctcaacttct	accccgtatt	tgccgtgcca	gaggtgettg
28561	ccacctatca	catcttttc	caaaactgca	agatacccct	atcctgccgt	gccaaccgca
28621	gccgagcgga	caagcagctg	gccttgcggc	agggcgctgt	catacctgat	ategeetege
28681	togacgaagt	gccaaaaatc	tttgagggtc	ttggacgcga	cgagaagcgc	gcggcaaacg
28741	ctctgcaaca	agaaaacagc	gaaaatgaaa	gtcactgtgg	agtgctggtg	gaacttgagg
28801	gtgagaacgc	acacctaacc	gtgctgaaac	gcagcatcga	ggtcacccac	tttgcctacc
28861	coocacttaa	cctaccccc	aaggttatga	gcacagtcat	gagcgagctg	ategtgegee
28921	gtgcacgacc	cctggagagg	gatgcaaact	tgcaagaaca	aaccgaggag	ggcctacccg
28981	cagttggcga	tgagcagetg	acacactage	ttgagacgcg	cgagcctgcc	gacttggagg
29041	agcgacgcaa	actaatqatq	gccgcagtgc	ttgttaccgt	ggagcttgag	tgcatgcage
29101	gattetttac	tgacccggag	atgcagcgca	agctagagga	aacgttgcac	tacaccttcc
29161	gccagggta	catacaccaa	gcctgcaaaa	tttccaacgt	ggagctctgc	aacctggtct
29221	cctaccttgg	aattttgcac	gaaaaccgcc	ttgggcaaaa	cgtgcttcat	tccacgctca
29281	ададсаадас	gcgccgcgac	tacgtccgcg	actgcgttta	cttatttctg	tgctacacct
29341	aacaaacaac	cataggcgtg	tagcagcagt	gcctggagga	gcgcaacctg	aaggagctgc
29401	agaagetget	aaagcaaaac	ttgaaggacc	tatggacggc	cttcaacgag	cgctccgtgg
29461	concocacct	ggcggacatt	atcttccccg	aacgcctgct	taaaaccctg	caacagggtc
29521	taccagactt	caccagtcaa	agcatgttgc	aaaactttag	gaactttatc	ctagagegee
29581	caggaattct	gcccgccacc	tactatacac	ttcctagcga	ctttgtgccc	attaagtacc
29641	gtgaatgccc	tecaccactt	tggggtcact	gctaccttct	gcagctagcc	aactaccttg
29701	cctaccactc	cgacatcatg	gaagacgtga	gcggtgacgg	cctactggag	tgtcactgtc
		J				

29761	gctgcaacct	atgcaccccg	caccgctccc	tggtctgcaa	ttcacaactg	cttagcgaaa
29821	gtcaaattat	cggtaccttt	gagctgcagg	gtccctcgcc	tgacgaaaag	tccgcggctc
29881	cggggttgaa	actcactccg	gggctgtgga	cgtcggctta	ccttcgcaaa	tttgtacctg
29941	aggactacca	cgcccacgag	attaggttct	acgaagacca	atcccgcccg	ccaaatgcgg
30001	agcttaccgc	ctgcgtcatt	acccagggcc	acatccttgg	ccaattgcaa	gccattaaca
30061	aagcccgcca	agagtttctg	ctacgaaagg	gacggggggt	ttacttggac	ccccagtccg
30121	gcgaggagct	caacccaatc	ccccgccgc	cgcagcccta	tcagcagccg	cgggcccttg
30181	cttcccagga	tggcacccaa	aaagaagctg	cagctgccgc	cgccgccacc	cacggacgag
		gggacagtca				
		gcctagacga				
30361	tcaccctcgg	tcgcattccc	ctcgccggcg	ccccagaaat	cggcaaccgt	tcccagcatt
30421	gctacaacct	ccgctcctca	ggcgccgccg	gcactgcccg	ttcgccgacc	caaccgtaga
30481	tgggacacca	ctggaaccag	ggccggtaag	tctaagcagc	cgccgccgtt	agcccaagag
30541	caacaacagc	gccaaggcta	ccgctcgtgg	cgcgtgcaca	agaacgccat	agttgcttgc
30601	ttgcaagact	gtgggggcaa	catctccttc	gcccgccgct	ttcttctcta	ccatcacggc
		cccgtaacat				
30721	ggcggcagcg	gcagcaacag	cagcggccac	gcagaagcaa	aggcgaccgg	atagcaagac
30781	tctgacaaag	cccaagaaat	ccacagcggc	ggcagcagca	ggaggaggag	cactgcgtct
		gaacccgtat				
30901	tgctatattt	caacagagca	ggggccaaga	acaagagctg	aaaataaaaa	acaggtctct
		acccgcagct				
		gaggctctct				
		caaatttaag				
31141	agcacctgtc	gtcagcgcca	ttatgagcaa	ggaaattccc	acgccctaca	tgtggagtta
31201	ccagccacaa	atgggacttg	cggctggagc	tgcccaagac	tactcaaccc	gaataaacta
31261	catgagcgcg	ggaccccaca	tgatatcccg	ggtcaacgga	atccgcgccc	accgaaaccg
31321	aattctcctc	gaacaggcgg	ctattaccac	cacacctcgt	aataacctta	atccccgtag
31381	ttggcccgct	gccctggtgt	accaggaaag	tcccgctccc	accactgtgg	tacttcccag
31441	agacgcccag	gccgaagttc	agatgactaa	ctcaggggcg	cagcttgcgg	gcggctttcg
31501	tcacagggtg	cggtcgcccg	ggcagggtat	aactcacctg	aaaatcagag	ggcgaggtat
31561	tcagctcaac	gacgagtcgg	tgagctcctc	tcttggtctc	cgtccggacg	ggacatttca
		gctggccgct				
		gagccgcgct				
		tacttcaacc				
		gacgcggtaa				
		ctgcgcctga				
31921	cggctccggt	gagttttgtt	actttgaatt	gcccgaagag	catatcgagg	gcccggcgca
31981	cggcgtccgg	ctcaccaccc	aggtagagct	tacacgtagc	ctgattcggg	agtttaccaa
32041	gcgccccctg	ctagtggagc	gggagcgggg	tccctgtgtt	ctgaccgtgg	tttgcaactg
32101	tcctaaccct	ggattacatc	aagatcttat	tccattcaac	taacaataaa	cacacaataa
32161	attacttact	taaaatcagt	cagcaaatct	ttgtccagct	tattcagcat	cacctccttt
32221	ccctcctccc	aactctggta	tttcagcagc	cttttagctg	cgaactttct	ccaaagtcta
32281	aatgggatgt	caaattcctc	atgttcttgt	ccctccgcac	ccactatctt	catattgttg
32341	cagatgaaac	gcgccagacc	gtctgaagac	accttcaacc	ctgtgtaccc	atatgacacg
32401	gaaaccggcc	ctccaactgt	gcctttcctt	acccctccct	ttgtgtcgcc	aaatgggttc
		ccccggagt				
32521	ggcatgcttg	cgctaaaaat	gggcagcggc	ctgtccctgg	atcaggcagg	caaccttaca
		tcactgtttc				
		cccttacagt				
32701	gtggtctctg	acaacactct	taccatgcaa	tcacaagcac	cgctaaccgt	gcaagactca
32761	aaacttagca	ttgctaccaa	agagccactt	acagtgttag	atggaaaact	ggccctgcag
32821	acatcagccc	ccctctctgc	cactgataac	aacgccctca	ctatcactgc	ctcacctcct
32881	cttactactg	caaatggtag	tctggctgtt	accatggaaa	acccacttta	caacaacaat
32941	ggaaaacttg	ggctcaaaat	tggcggtcct	ttgcaagtgg	ccaccgactc	acatgcacta
33001	acactaggta	ctggtcaggg	ggttgcagtt	cataacaatt	tgctacatac	aaaagttaca

33061	ggcgcaatag	ggtttgatac	atctggcaac	atggaactta	aaactggaga	tggcctctat
33121	gragatageg	ccggtcctaa	ccaaaaacta	catattaatc	taaataccac	aaaaggcctt
33121	actiticaca	acaccgcaat	aacaattaac	gctggaaaag	ggttggaatt	tgaaacagac
332/11	tecteaaaca	gaaatcccat	aaaaacaaaa	attogatcag	gcatacaata	taataccaat
33301	agagetatag	ttgcaaaact	tggaacaggc	ctcagttttg	acageteegg	agccataaca
33361	atagacaaca	taaacaatga	cagacttact	ctttggacaa	caccagaccc	atccccaaat
33301	tacagaatta	cttcagataa	agactgcaag	ctaactctqq	cgctaacaaa	atgtggcagt
22421	casattttaa	gcactgtttc	agetttggca	gtatcaggta	atatogcctc	catcaatgga
22541	actotaacca	gtgtaaactt	ggttcttaga	tttgatgaca	acqqaqtqct	tatgtcaaat
33501	tcatcactoo	acaaacagta	ttggaacttt	agaaacgggg	actccactaa	cggtcaacca
22661	tacacttato	ctgttgggtt	tatgccaaac	ctaaaagctt	acccaaaaac	tcaaagtaaa
22721	actornana	gtaatattgt	tagccaggtg	tatcttaatq	gtgacaagtc	taaaccattg
22701	actycaaaaa	ttacgctaaa	tagaacagat	gaaaccaacc	aagtaagcaa	atactcaata
22041	tasttasatt	ggtcctggaa	cagtagacaa	tacactaatq	acaaatttgc	caccaattcc
33001	tatacettet	cctacattgc	ccargaataa	agaatcgtga	acctgttgca	tgttatgttt
33301	cacacccccc	atttttcaat	tacagaaaat	ttcaagtcat	ttttcattca	gtagtatagc
33301	caacgtgttt	catagettat	actaatcacc	gtaccttaat	caaactcaca	gaaccctagt
34021	atteaacete	ccacctccct	cccaacacac	agagtacaca	atcetttete	cccggctggc
34001	atteaacety	atcatatcat	adataacada	catattetta	ggtgttatat	tccacacggt
34141	cttadatage	gccaaacgct	catcagtgat	gttaataaac	tececaaaca	gctcgcttaa
34201	ettestetes	ctgtccagct	actagaccac	aggetgetgt	ccaacttgcg	gttgctcaac
34201	grecargrey	ggagaagtcc	accetacat	aggetjetje	tcataatcqt	gcatcaggat
34321	gggcggcgaa	tgctgcagca	acgcccacat	aaactoctoc	caccaccact	ccgtcctgca
34381	agggeggegg	atggcagtgg	teteeteace	dateegeege	accoccoca	gcataaggcg
34441	ggaatacaac	cgggcacagc	accececage	gatgateegt	aagtcagcac	agtaactgca
34501	cettgteete	acaatattgt	ttaaaatccc	acanthreaan	acactatate	caaagctcat
34561	gcacagtacc	acagaaccca	cataaccetc	ataccacaa	cocaggtaga	ttaagtggcg
34621	ggcggggacc	acacacctgg	acataaacat	tacctcttt	agcatattat	aattcaccac
34681	acccctcata	catataaacc	totoattaaa	categococca	tccaccacca	tcctaaacca
34/41	ctcccggtac	acctgcccgc	caactataca	ctacaaaaaa	ccaaaactaa	aacaatgaca
34801	gctggccaaa	caggactcgt	eaccatogat	catcatocto	gtcatgatat	caatgttggc
34861	gtggagagee	cacacgtgca	tacacttcct	caccatgece	agetectece	gcgtcagaac
34921	acaacacagg	ggaacaaccc	attecteast	caggattaat	ccacactec	agggaagacc
34981	catateceag	ctcacgttgt	accectgaac	agegeaaae	tcaaacaaca	gcggatgatc
35041	tegeacgtaa	gtagcgcggg	tttctctctc	agegeeacat	agacgatccc	tactgtacgg
35101	ctccagtatg	gtagegeggg	atastattaa	testastate	ataccaaata	gaacgccgga
35161	agtgcgccga	gacaaccgag	accycyctyy	tacagagaga	acaecadat	ctacatetee
35221	cgtagtcata	tttcctgaag cttagatcgc	totatataat	agttataata	tatccactct	ctcaaagcat
35281	ggtctcgccg	cttagatege	aattatatat	agetycagea	atacaccact	accetaataa
35341	ccaggcgccc	cctggcttcg cgcagaataa	ggttttatgt	gaacectec	acattcattc	tacaaatcac
35401	catccaccac	agcgggaaga	getacaccca	ccatatttt	tttttattc	caaaagatta
35461	acacgggagg	caaaatgaag	atctattaac	taaacacact	ccctccat	gacataatca
35521	tccaaaacct	ccaaagaaca	accuactaag	tttataaaat	ottocacaat	ggcttccaaa
35581	aactctacag	ccaaagaaca	gacaacygca	taaagac	accettcace	graatctcc
35641	aggcaaacgg	ccctcacgtc	ttannanta	cccaataat	teteatetea	ccaccttctc
35701	tctataaaca	ttccagcacc	cccaaccatg	cccaaacaac	ttataaaaat	ctactccaga
35761	aatatatctc	taagcaaatc	ccgaacacca	agtetggtea	cogcaadaac	ctgctccaga
35821	gcgccctcca	cetteageet	caagcagcga	accargaccy	caaaaactca	ggttcctcac
35881	agacctgtat	aagattcaaa	ageggaacat	taacaaaat	accecyatec	cgtaggtccc
35941	ttcgcagggc	cagctgaaca	taatcgtgca	ggtetgeacg	gaccaycycy	gccacttccc
36001	cgccaggaac	catgacaaaa	gaacccacac	cgattatgac	acgeatacte	ggagctatgc
36061	taaccagcgt	agccccgatg	caagettgtt	gcatgggcgg	cyacacaada	tgcaaggtgc
36121	tgctcaaaaa	atcaggcaaa	gcctcgcgca	aaaaagaaag	cacategtag	tcatgctcat
36181	gcagataaag	gcaggtaagc	tccggaacca	ccacagaaaa	agacaccatt	tttctctcaa
36241	acatgtctgc	gggtttctgc	ataaacacaa	aataaaataa	caaaaaaaca	tttaaacatt
36301	agaagcctgt	cttacaacag	gaaaaacaac	ccttataagc	ataagacgga	ctacggccat

FIG. 4L

36361	gccggcgtga	ccgtaaaaaa	actggtcacc	gtgattaaaa	agcaccaccg	acagctcctc
36421	ggtcatgtcc	ggagtcataa	tgtaagactc	ggtaaacaca	tcaggttgat	tcacatcggt
36481	cagtgctaaa	aagcgaccga	aatagcccgg	gggaatacat	acccgcaggc	gtagagacaa
36541	cattacagcc	cccataggag	gtataacaaa	attaatagga	gagaaaaaca	cataaacacc
36601	tgaaaaaccc	tcctgcctag	gcaaaatagc	accctcccgc	tccagaacaa	catacagcgc
					aaagaaaacc	
						caagtgcaga
					gtccacaaaa	
					aacccacaac	
36901	cgtcacttcc	gttttcccac	gttacgtcac	ttcccatttt	aagaaaacta	caattcccaa
36961	cacatacaag	ttactccgcc	ctaaaaccta	cgtcacccgc	cccgttccca	cgccccgcgc
37021	cacgtcacaa	actccacccc	ctcattatca	tattggcttc	aatccaaaat	aaggtatatt
37081	attgatgatg					

10		30	50	
			CGGGGCCTACTTG(
MetAlaPı	roIleThrAlaTy		ArgGlyLeuLeuGl	LYCYSIIeIIeThr 20
		10		20
	70	90		110
AGCCTTAG		AGAACCAGGTCGAG	GGAGAGGTTCAGG	rggtttccaccgca
SerLeuTh	nrGlyArgAspLy	/sAsnGlnValGlu	uGlyGluValGlnVa	
		30		40
		150		170
	130 		GTGTGTTGGACCG	
			yValCysTrpThrV	
		50		60
				020
	190	210	* * **********************************	230 ACACTAATCTCGAC
			AATCACCCAGATGT	
				yrThrAsnValAsp
	•	70		80
	250	270		290
				CACCATGCACCTGT
				hrProCysThrCys
GIIMSDD	euvaigiyiipa	90	,	100
	310	330		350
				CGGTGCGCCGGCGG
				++
GlySerS	erAspLeuTyrL	euvaiTnrarghi 110	SAIAASPVAILLEP	roValArgArgArg 120
		110		
	370	390		410
GGCGACA	GTAGGGGGAGCC'	TGCTCTCCCCAG	GCCTGTCTCCTACT	TGAAGGGCTCTTCG
				+
GlyAspS	erArgGlySerL		gProValSerTyrL	euLysGlySerSer
		130		140

FIG. 5A

470	450	430
ATCTTCCGGGCTGCCGTATGC		
TlePheArgAlaAlaValCys 160	SerGlyHisAlaValGly1 150	GlyGlyProLeuLeuCysPr
100	130	
530	510	490
GAGTCCATGGAAACTACTATG	GTGGACTTTGTGCCCGTAG	
++		
GluSerMetGluThrThrMet		ThrArgGlyValAlaLysAl
180	170	
590	570	550
GTACCGCAGTCATTTCAAGTG		
++	+	
ValProGlnSerPheGlnVal	AsnSerSerProProAlaV	ArgSerProValPheThrAs
200	190	
650	630	610
AAAGTGCCGGCTGCATATGCA		
++		
LysValProAlaAlaTyrAla	SlySerGlyLysSerThrL	AlaHisLeuHisAlaProTh
220	210	
84.0		
710	690	670
GCCGCTACCTTAGGGTTTGGG ++		
AlaAlaThrLeuGlyPheGly		
240	230	
770	750	730
AGAACTGGGGTAAGGACCATT ++		
ArgThrGlyValArgThrIle		
260	250	AIGIT THE COCIDY DILLUIT
830	810	790
TTTCTTGCCGATGGTGGTTGC		
++	·	·
PheLeuAlaAspGlyGlyCys	yrSerThrTyrGlyLysPl 270	ThrThrGlyAlaProValTh

850	870	890
		CCATTCAACTGACTCGACTACA
		-++ sHisSerThrAspSerThrThr
SerGlyGlyAlaTyrAsp	290	300
	22.0	
910	930	950
		GGCTGGAGCGCGGCTTGTCGTG
		-++ rAlaGlyAlaArgLeuValVal
lierendilitedilim	310	320
970	990	1010
		ACACCCAAACATCGAGGAGGTG
		-++ oHisProAsnIleGluGluVal
Leualainialainipio	330	340
1030	1050	1070
		AGCCATCCCCATTGAAGCCATC
		-++
AlaLeuSerAsnThrGly		sAlaIleProIleGluAlaIle
	350	300
1090	1110	1130
	CATTTTCTGTCATTCCAAGAA	GAAGTGCGACGAGCTCGCCGCA
		-++
ArgGlyGlyArgHisLe		sLysCysAspGluLeuAlaAla
	370	380
1150	1170	1190
		CCGGGGGCTCGATGTGTCCGTC
		-++
LysLeuSerGlyLeuGly	yIleAsnAlaValAlaTyrTy	rArgGlyLeuAspValSerVal
	390	400
1210	1230	1250
		ACGCTCTGATGACGGGCTATACG
		++
IleProThrIleGlyAs	pValValValValAlaThrAs	spAlaLeuMetThrGlyTyrThr
	410	420

1270	1290	1310
		TCACCCAGACAGTCGACTTCAGC
		++
GlyAspPheAspSerVal		alThrGlnThrValAspPheSer
	430	440
1330	1350	1370
		TCAAGACGCAGTGTCGCGCTCG
		++
LeuAspProThrPheThr	:IleGluThrThrThrValP:	roGlnAspAlaValSerArgSer
	450	460
1390		1430
		rctacaggtttgtgactccggga
GlnArgArgGlyArgThr	GiyargGiyargargGiyii 470	leTyrArgPheValThrProGly 480
	470	400
1450	1470	1490
		STGAGTGCTATGACGCGGGCTGT
		-++
GluArgProSerGlyMet	PheAspSerSerValLeuCy	sGluCysTyrAspAlaGlyCys
	490	500
1510	1530	1550
		GGTTGCGGGCCTACCTGAACACA
		++ rgLeuArgAlaTyrLeuAsnThr
AlampiyiGlubeuini	510	520
	340	, , , , , , , , , , , , , , , , , , ,
1570 .	1590	1610
		GGAGAGTGTCTTCACAGGCCTC
	-+	-++
ProGlyLeuProValCys	GlnAspHisLeuGluPheTr	pGluSerValPheThrGlyLeu
	530	540
1630	1650	1670
		AGGCAGGAGACAACTTCCCCTAC
		nAlaGlyAspAsnPheProTyr
INTUISTIEWSPATEUIS	550	560

FIG. 5D

730	173	1710	1690
		CAAGCCACGGTGTGCGCCA	
		+	
	rgAlaGlnAlaProProF	GlnAlaThrValCysAlaA	LeuValAlaTyr
580		570	
790	179	1770	1750
		TGTCTCATACGGCTGAAAC	
		CysLeulleArgLeuLys	
600		590	
850		1830	1810
		GCCGTCCAAAATGAGGTC	
		+	
ernrLysTyrffe 620	hrLeuThrHisProlle	AlaValGlnAsnGluVal	TyrArgLeuGly
620		610	
910	191	1890	1870
GCTGGTGGGCGGA	TCACTAGCACCTGGGTG(TCGGCTGACCTGGAGGTC	
		+	
		SerAlaAspLeuGluValV	
640		630	
.970		1950	1930
		CTGGCCGCGTATTGCCTG	
660	nriniglyServalval.	LeuAlaAlaTyrCysLeu 650	ValLeuAlaAla
		630	
2030	20:	2010	1990
CTACCAGGAGTTC	CCGACAGGGAGTTTCTC'	GGGAGGCCGGCTATTGTT	
		+	
		GlyArgProAlaIleVal	
680		670	
2090	•	2070	2050
		GAGTGCGCCTCGCACCTC	
lymetGInLeuAla 700		GluCysAlaSerHisLeu 690	AspGluMetGlu

FIG. 5E

2110	2130	2150
		CAGCCACCAAACAAGCGGAGGCT
		+
GluGinPheLysGinLy	SAIaLeuGIYLeuLeuGITT 710	nrAlaThrLysGlnAlaGluAla 720
	710	720
2170	2190	2210
		AGACATTCTGGGCGAAGCACATG
		luThrPheTrpAlaLysHisMet
	730	740
2230		2270
		TATCCACTCTGCCTGGGAACCCC
		++ euSerThrLeuProGlyAsnPro
TipasnPheileSelGi	ylleginlylleuxlaglybe 750	ruser milleuri ogryksmrio 760
	730	
2290	2310	2330
GCAATAGCATCATTGAT	GGCATTCACAGCCTCTATCAC	CAGCCCGCTCACCACCCAAAGT
		++
AlaIleAlaSerLeuMe	•	rSerProLeuThrThrGlnSer
	770	780
2350	2370	2390
		CCAACTCGCCCCCCCCAGCGCC
		-++
ThrLeuLeuPheAsnIle	eLeuGlyGlyTrpValAlaAl	.aGlnLeuAlaProProSerAla
	790	800
	2430	
		TGTTGGCAGCATAGGCCTTGGG
•		aValGlySerIleGlyLeuGly
Alaberatur nevaror	810	820
		·
2470	2490	2510
AAGGTGCTTGTGGACAT	rctggcgggttatggagcagg	AGTGGCCGGCGCGCTCGTGGCC
•	+	
LysValLeuValAspIle		yValAlaGlyAlaLeuValAla
	830	840

2530	2550	2570
TTCAAGGTCATGAGCGGCGA	GATGCCCTCCACCGAG	GACCTGGTCAATCTACTTCCTGCC
		+
		AspLeuValAsnLeuLeuProAla
PhebysvaimecselGiyGi		860
	850	000
2590	2610	2630
		TGTGCAGCAATACTGCGTCGACAC
		++
IleLeuSerProGlyAlaLe	uValValGlyValVal	CysAlaAlaIleLeuArgArgHis
	870	880
2650	2670	2690
	YPCTCCACTCGATGAAC	CGGCTGATAGCGTTCGCCTCGCGG
		++
		ArgLeuIleAlaPheAlaSerArg
ValGlyProGlyGluGlyAl		
	890	900
2710	2730	2750
		AGCGACGCCGCAGCGCGTGTTACT
		+
GlyAsnHisValSerProTh	nrHisTyrValProGlu	SerAspAlaAlaAlaArgValThr
	910	920
2770	2790	2810
— · · · ·		AAAAGGCTCCACCAGTGGATTAAT
		+
GlnIleLeuSerSerLeuTh		LysArgLeuHisGlnTrpIleAsn
	930	940
2830	2850	2870
GAAGACTGCTCCACACCGTC	STTCCGGCTCGTGGCTA	AGGGATGTTTGGGACTGGATATGC
	+	+
		ArgAspValTrpAspTrpIleCys
Grunspey soci initizes	950	960
	,,,,	
2000	2010	2930
2890	2910	
		AAGCTCCTGCCGCAGCTACCGGGA
		+
ThrValLeuThrAspPheLy	ysThrTrpLeuGlnSer	LysLeuLeuProGlnLeuProGly
	970	980

FIG. 5G

2950	2970	2990
		CTGGCGGGGAGACGGCATCATG
		++ alTrpArgGlyAspGlyIleMet
Valiable and and an	990	1000
3010	3030 .	3050 ATGTCAAAAACGGTTCCATGAGG
		++
GlnThrThrCysProCys0	GlyAlaGlnIleThrGlyHi	isValLysAsnGlySerMetArg
	1010	1020
2070	3090	3110
3070 ATCGTCGGGCCTAAGACCT		GAACATTCCCCATCAACGCATAC
		++
IleValGlyProLysThrO		yThrPheProIleAsnAlaTyr
	1030	1040
3130	3150	3170
		ATTCTAGGGCGCTGTGGCGGGTG
		++
ThrThrGlyProCysThrE		rSerArgAlaLeuTrpArgVal 1060
	1050	1000
3190	3210	3230
		ATTTCCACTACGTGACGGGCATG
		-++
AlaAlaGluGluTyrvalG	siuvaitniargvaigiyas 1070	pPheHisTyrValThrGlyMet 1080
3250	3270	3290
		TCCTGAATTCTTCACGGAGGTG
		aProGluPhePheThrGluVal
IIII IIII ASPASII VALLY SO	1090	1100
3310	3330	3350
	AGGTACGCTCCGGCGTGCAG	GCCTCTCCTACGGGAGGAGGTT
		gProLeuLeuArgGluGluVal
• • · · · · · · · · · · · · · · · · ·	1110	1120

3370	3390	3410
		ACAGCTACCATGCGAGCCCGAA
		erGlnLeuProCysGluProGlu
_	1130	1140
3430	3450	3470
		CTCCCACATCACAGCAGAAACG
		oSerHisIleThrAlaGluThr
-	1150	1160
3490	3510	3530
		GGCCAGCTCTTCAGCTAGCCAG
		euAlaSerSerSerAlaSerGln
	1170	1180
3550	3570	3590
TTGTCTGCGCCTTCCTTG		ACCATGTCTCTCCGGACGCTGAC
		++ LsHisValSerProAspAlaAsp
Dedgetwiat roperace	1190	1200
3610	3630	3650
CTCATCGAGGCCAACCTC	CTGTGGCGCAGGAGATGG	GCGGGAACATCACCCGCGTGGAG
		++ lyGlyAsnIleThrArgValGlu
LeuileGlualdasilee	1210	1220
3.670	3690	3710
3670 TCGGAGAACAAGGTGGTA		CGCTTCGAGCGGAGGAGGATGAG
SerGluAsnLysValVal	1230	roLeuArgAlaGluGluAspGlu 1240
3730	3750	3770
		CCAAGAAGTTCCCCGCAGCGATG
		erLysLysPheProAlaAlaMet
	1250	1260

FIG. 51

3790	3810	3830
CCCATCTGGGCGCGCCCGG	ATTACAACCCTCCACTGT	PAGAGTCCTGGAAGGACCCGGAC
	+	++
ProIleTrpAlaArgProA	.spTyrAsnProProLeuLe	euGluSerTrpLysAspProAsp
	1270	1280
3850	3870	3890
		CTATCAAGGCCCCTCCAATACCA
		++
TyrValProProValValH		colleLysAlaProProllePro
	1290	1300
	3930	3950
		CTCCGTGTCTTCTGCCTTAGCG
ProproArgArgLysArgT	nrvalvalleuThrGluSe 1310	erSerValSerSerAlaLeuAla 1320
	1310	1320
3970	3990	4010
		GGCCGTCGACAGCGGCACGGCG
		-+
		erAlaValAspSerGlyThrAla
•	1330	1340
4030	4050	4070
ACCGCCCTTCCTGACCAGG	CCTCCGACGACGGTGACAA	AGGATCCGACGTTGAGTCGTAC
	+	-++
ThrAlaLeuProAspGlnA	laSerAspAspGlyAspLy	sGlySerAspValGluSerTyr
	1350	1360
4090	4110	4130
		CGATCTCAGTGACGGGTCTTGG
		-++
SerSerMetProProLeuG		oAspLeuSerAspGlySerTrp
	1370	1380
4150	4170	4190
		CTGCTCAATGTCCTACACATGG
	t	
		sCysSerMetSerTyrThrTrp
Jel IIII valdetotuotun.	1300	1400

FIG. 5J

4210	4230	4250
ACAGGCGCCTTGATCACG	CCATGCGCTGCGGAGGAAAG	CAAGCTGCCCATCAACGCGTTG
		-++
ThrGlyAlaLeuIleThr		rLysLeuProIleAsnAlaLeu
	1410	1420
40.00	4290	4310
4270		CACAACATCTCGCAGCGCAGGC
AGCAACTCTTTGCTGCGC	CACCATAACATGGTTTATGC	-+
		aThrThrSerArgSerAlaGly
Delimenta della mara	1430	1440
		•
4330	4350	4370
CTGCGGCAGAAGAAGGTC	:ACCTTTGACAGACTGCAAGT	CCTGGACGACCACTACCGGGAC
		_+
LeuArgGlnLysLysVal		lLeuAspAspHisTyrArgAsp
	1450	1400
4200	4410	4430
4390		GGCTAAACTCCTATCCGTAGAG
GIGCICAAGGAGAIGAAG	+	-++
		rsAlaLysLeuLeuSerValGlu
•	1470	1480
4450	4470	4490
		CCAAGTTTGGCTATGGGGCAAAG
GluAlaCysLysLeuTh		erLysPheGlyTyrGlyAlaLys 1500
	1490	2500
4510	4530	4550
		rccactccgtgtggaaggacttg
	+	++
AspValArgAsnLeuSe:	rSerLysAlaValAsnHisI	leHisSerValTrpLysAspLeu
	1510	1520
4570	4590	4610
		TGGCAAAAAATGAGGTTTTCTGT
		++
LeuGluAspThrValTh		etAlaLysAsnGluValPheCys 1540
	1530	1340

FIG. 5K

4630	4650	4670
		TATCGTATTCCCAGATCTGGGA
		eulleValPheProAspLeuGly
, mioint 1001 mil 100 mil	1550	1560
		4500
4690 GTCCCTCTATGCGAGAAG	4710 ATGCCCCTCTATGATGTGGT	4730 CTCCACCCTTCCTCAGGTCGTG
		-++
ValArgValCysGluLys		lSerThrLeuProGlnValVal
	1570	1580
4750	4770	4790
		GCGAGTCGAGTTCCTGGTGAAT
		-+++
MetGlySerSerlyrGly	1590	nArgValGluPheLeuValAsn 1600
4810		4850
		TGACACTCGCTGTTTCGACTCA
		rAspThrArgCysPheAspSer
	.1610	1620
4870	4890	4910
		TTACCAATGTTGTGACTTGGCC
		-++
ThrValThrGluAsnAsp		eTyrGlnCysCysAspLeuAla
	1630	1640
4930	4950	4970
CCCGAAGCCAGACAGGCC	ATAAAATCGCTCACAGAGCG	GCTTTATATCGGGGGTCCTCTG
		-++ gLeuTyrIleGlyGlyProLeu
PIOGIUATAALYGINATA	1650	1660
4990	5010	5030
	AACTGCGGTTATCGCCGGTG -+	CCGCGCGAGCGGCGTGCTGACG
		sArgAlaSerGlyValLeuThr
	1670	1690

FIG. 5L

5050	5070	5090
		CCTCTGCAGCCTGTCGAGCTGCG
		++ laSerAlaAlaCysArgAlaAla
Intocto, Bot, ibutility	1690	1700
* == ·	5130	5150
		ACCTTGTCGTTATCTGTGAAAGC
		spLeuValValIleCysGluSer
	1710	1720
5170	5190	5210
		TCACGGAGGCTATGACTAGGTAC
		heThrGluAlaMetThrArgTyr
_	1730	1740
5230	5250	5270
		ACTTGGAGCTGATAACATCATGT
		spLeuGluLeuIleThrSerCys
•	1750	1760
5290	5310	5330
		AAAGGGTGTACTACCTCACCCGT
		ysArgValTyrTyrLeuThrArg
	1770	1780
5350	5370	5390
		CAGCTAGACACACTCCAGTTAAC
		hrAlaArgHisThrProValAsn
nopi i o i i i i i i i i i i i i i i i i	1790	1800
5410	5430	5450
		TGTGGGCAAGGATGATTCTGATG
		++ euTrpAlaArgMetIleLeuMet
SerrpheuGlyAsnilel	1810	eulipalaalgmetilebeumet 1820

FIG. 5M

5470	5490	5510
		rtgaaaaagccctggactgccag ++
		euGluLysAlaLeuAspCysGln
	1830	1840
5530	5550	5570
		PACCTCAGATCATTGAACGACTC
		euProGlnIleIleGluArgLeu
	1850	1860
5590	5610	5630
		CAGGTGAGATCAATAGGGTGGCT
		++ roGlyGluIleAsnArgValAla
	1870	1880
5650	5670	5690
		CTGGAGACATCGGGCCAGGAGC
		-++ :lTrpArgHisArgAlaArgSer
	1890	1900
5710	5730	5750
		CACTTGTGGCAAGTACCTCTTC
		aThrCysGlyLysTyrLeuPhe
•	1910	1920
5770	5790	5810
		CCCGGCTGCGTCCCAGCTGGAC
		eProAlaAlaSerGlnLeuAsp
	1930	1940
5830	5850	5870
		CATATATCACAGCCTGTCTCGT
		-++ pIleTyrHisSerLeuSerArg
<u>-</u> -	1950	1960

5890	5910	5930
GCCCGACCCCGCTGGTT	CATGCTGTGCCTACTCTACTT	CTGTAGGGGTAGGCATCTAC
	+	·+
AlaArgProArgTrpPl	neMetLeuCysLeuLeuLeuLeuS	SerValGlyValGlyIleTyr
• •	1970	1980
5950 5955		
CTGCTCCCCAACCGA	(SEQ. ID. NO. 5)	
LeuLeuProAsnArg	(SEQ. ID. NO. 6)	
1985		

1	TCGCGCGTTT	CGGTGATGAC	GGTGAAAACC	TCTGACACAT	GCAGCTCCCG
51	GAGACGGTCA	CAGCTTGTCT	GTAAGCGGAT	GCCGGGAGCA	GACAAGCCCG
101	TCAGGGCGCG	TCAGCGGGTG	TTGGCGGGTG	TCGGGGCTGG	CTTAACTATG
151	CGGCATCAGA	GCAGATTGTA	CTGAGAGTGC	ACCATATGCG	GTGTGAAATA
201	CCGCACAGAT	GCGTAAGGAG	AAAATACCGC	ATCAGATTGG	CTATTGGCCA
251	TTGCATACGT	TGTATCCATA	TCATAATATG	TACATTTATA	TTGGCTCATG
301	TCCAACATTA	CCGCCATGTT	GACATTGATT	ATTGACTAGT	TATTAATAGT
351	AATCAATTAC	GGGGTCATTA	GTTCATAGCC	CATATATGGA	GTTCCGCGTT
401	ACATAACTTA	CGGTAAATGG	CCCGCCTGGC	TGACCGCCCA	ACGACCCCCG
451	CCCATTGACG	TCAATAATGA	CGTATGTTCC	CATAGTAACG	CCAATAGGGA
501	CTTTCCATTG	ACGTCAATGG	GTGGAGTATT	TACGGTAAAC	TGCCCACTTG
551	GCAGTACATC	AAGTGTATCA	TATGCCAAGT	ACGCCCCCTA	TTGACGTCAA
601	TGACGGTAAA	TGGCCCGCCT	GGCATTATGC	CCAGTACATG	ACCTTATGGG
651	ACTTTCCTAC	TTGGCAGTAC	ATCTACGTAT	TAGTCATCGC	${\tt TATTACCATG}$
701	GTGATGCGGT	TTTGGCAGTA	CATCAATGGG	CGTGGATAGC	GGTTTGACTC
751	ACGGGGATTT	CCAAGTCTCC	ACCCCATTGA	CGTCAATGGG	AGTTTGTTTT
801	GGCACCAAAA	TCAACGGGAC	TTTCCAAAAT	GTCGTAACAA	CTCCGCCCCA
851	TTGACGCAAA	TGGGCGGTAG	GCGTGTACGG	TGGGAGGTCT	ATATAAGCAG
901	AGCTCGTTTA	GTGAACCGTC	AGATCGCCTG	GAGACGCCAT	CCACGCTGTT
951	TTGACCTCCA	TAGAAGACAC	CGGGACCGAT	CCAGCCTCCG	CGGCCGGGAA
1001	CGGTGCATTG	GAACGCGGAT	TCCCCGTGCC	AAGAGTGACG	TAAGTACCGC
1051	CTATAGACTC	TATAGGCACA	CCCCTTTGGC	TCTTATGCAT	GCTATACTGT
1101	TTTTGGCTTG	GGGCCTATAC	ACCCCCGCTT	CCTTATGCTA	TAGGTGATGG
1151	TATAGCTTAG	CCTATAGGTG	TGGGTTATTG	ACCATTATTG	ACCACTCCCC
1201	TATTGGTGAC	GATACTTTCC	ATTACTAATC	CATAACATGG	CTCTTTGCCA
1251	CAACTATCTC	TATTGGCTAT	ATGCCAATAC	TCTGTCCTTC	AGAGACTGAC
1301	ACGGACTCTG	TATTTTTACA	GGATGGGGTC	CCATTTATTA	TTTACAAATT
1351	CACATATACA	ACAACGCCGT	CCCCGTGCC	CGCAGTTTTT	ATTAAACATA
1401	GCGTGGGATC	TCCACGCGAA	TCTCGGGTAC	GTGTTCCGGA	CATGGGCTCT
1451	TCTCCGGTAG	CGGCGGAGCT	TCCACATCCG	AGCCCTGGTC	CCATGCCTCC
1501	AGCGGCTCAT	GGTCGCTCGG	CAGCTCCTTG	CTCCTAACAG	TGGAGGCCAG
1551	ACTTAGGCAC	AGCACAATGC	CCACCACCAC	CAGTGTGCCG	CACAAGGCCG
1601	TGGCGGTAGG	GTATGTGTCT	GAAAATGAGC	GTGGAGATTG	GGCTCGCACG
1651	GCTGACGCAG	ATGGAAGACT	TAAGGCAGCG	GCAGAAGAAG	ATGCAGGCAG
1701	CTGAGTTGTT	GTATTCTGAT	AAGAGTCAGA	GGTAACTCCC	GTTGCGGTGC
1751	TGTTAACGGT	GGAGGGCAGT	GTAGTCTGAG	CAGTACTCGT	TGCTGCCGCG
1801	CGCGCCACCA	GACATAATAG	CTGACAGACT	AACAGACTGT	TCCTTTCCAT
1851	GGGTCTTTTC	TGCAGTCACC	GTCCTTAGAT	CTAGGTACCA	GATATCAGAA
1901	TTCAGTCGAC	AGCGGCCGCG	ATCTGCTGTG	CCTTCTAGTT	GCCAGCCATC
1951	TGTTGTTTGC	CCCTCCCCG	TGCCTTCCTT	GACCCTGGAA	GGTGCCACTC
2001	CCACTGTCCT	TTCCTAATAA	AATGAGGAAA	TTGCATCGCA	TTGTCTGAGT
2051	AGGTGTCATT	CTATTCTGGG	GGGTGGGGTG	GGGCAGGACA	GCAAGGGGGA

FIG. 6A

2101	-			GGATGCGGTG	
2151				CGGTTCCTCC	
2201				CACCCTGTCC	
2251				TCATAGCTCA	
2301				AGCGGTCTCT	
2351				GAGTGGGAAG	
2401				AAATGCCTCC	
2451				CCGCTTCCTC	
2501	TCGCTGCGCT	CGGTCGTTCG	GCTGCGGCGA	GCGGTATCAG	CTCACTCAAA
2551				GGATAACGCA	
2601	TGTGAGCAAA	AGGCCAGCAA	AAGGCCAGGA	ACCGTAAAAA	GGCCGCGTTG
2651				GACGAGCATC	
2701	ACGCTCAAGT	CAGAGGTGGC	GAAACCCGAC	AGGACTATAA	AGATACCAGG
2751	CGTTTCCCCC	TGGAAGCTCC	CTCGTGCGCT	CTCCTGTTCC	GACCCTGCCG
2801	CTTACCGGAT	ACCTGTCCGC	CTTTCTCCCT	TCGGGAAGCG	TGGCGCTTTC
2851	TCATAGCTCA	CGCTGTAGGT	ATCTCAGTTC	GGTGTAGGTC	GTTCGCTCCA
2901	AGCTGGGCTG	TGTGCACGAA	CCCCCGTTC	AGCCCGACCG	CTGCGCCTTA
2951	TCCGGTAACT	ATCGTCTTGA	GTCCAACCCG	GTAAGACACG	ACTTATCGCC
3001	ACTGGCAGCA	GCCACTGGTA	ACAGGATTAG	CAGAGCGAGG	TATGTAGGCG
3051	GTGCTACAGA	GTTCTTGAAG	TGGTGGCCTA	ACTACGGCTA	CACTAGAAGA
3101	ACAGTATTTG	GTATCTGCGC	TCTGCTGAAG	CCAGTTACCT	TCGGAAAAAG
3151	AGTTGGTAGC	TCTTGATCCG	GCAAACAAAC	CACCGCTGGT	AGCGGTGGTT
3201	TTTTTGTTTG	CAAGCAGCAG	ATTACGCGCA	GAAAAAAAGG	ATCTCAAGAA
3251	GATCCTTTGA	TCTTTTCTAC	GGGGTCTGAC	GCTCAGTGGA	ACGAAAACTC
3301	ACGTTAAGGG	ATTTTGGTCA	TGAGATTATC	AAAAAGGATC	TTCACCTAGA
3351	TCCTTTTAAA	TTAAAAATGA	AGTTTTAAAT	CAATCTAAAG	TATATATGAG
3401	TAAACTTGGT	CTGACAGTTA	CCAATGCTTA	ATCAGTGAGG	CACCTATCTC
3451	AGCGATCTGT	CTATTTCGTT	CATCCATAGT	TGCCTGACTC	GGGGGGGGG
3501	GGCGCTGAGG	TCTGCCTCGT	GAAGAAGGTG	TTGCTGACTC	ATACCAGGCC
3551	TGAATCGCCC	CATCATCCAG	CCAGAAAGTG	AGGGAGCCAC	GGTTGATGAG
3601	AGCTTTGTTG	TAGGTGGACC	AGTTGGTGAT	TTTGAACTTT	TGCTTTGCCA
3651	CGGAACGGTC	TGCGTTGTCG	GGAAGATGCG	TGATCTGATC	CTTCAACTCA
3701	GCAAAAGTTC	GATTTATTCA	ACAAAGCCGC	CGTCCCGTCA	AGTCAGCGTA
3751					AGAAAAACTC
3801	ATCGAGCATC	AAATGAAACT	GCAATTTATT	CATATCAGGA	TTATCAATAC
3851	CATATTTTTG	AAAAAGCCGT	TTCTGTAATG	AAGGAGAAAA	CTCACCGAGG
3901					TTCCGACTCG
3951					ATAAGGTTAT
4001					GAATGGCAAA
4051					CATTACGCTC
					CGTGATTGCG
					ATTACAAACA

FIG. 6B

4201	GGAATCGAAT	GCAACCGGCG	CAGGAACACT	GCCAGCGCAT	CAACAATATT
4251	TTCACCTGAA	TCAGGATATT	CTTCTAATAC	CTGGAATGCT	GTTTTCCCGG
4301	GGATCGCAGT	GGTGAGTAAC	CATGCATCAT	CAGGAGTACG	GATAAAATGC
4351	TTGATGGTCG	GAAGAGGCAT	AAATTCCGTC	AGCCAGTTTA	GTCTGACCAT
4401	CTCATCTGTA	ACATCATTGG	CAACGCTACC	TTTGCCATGT	TTCAGAAACA
4451	ACTCTGGCGC	ATCGGGCTTC	CCATACAATC	GATAGATTGT	CGCACCTGAT
4501	TGCCCGACAT	TATCGCGAGC	CCATTTATAC	CCATATAAAT	CAGCATCCAT
4551	GTTGGAATTT	AATCGCGGCC	TCGAGCAAGA	CGTTTCCCGT	TGAATATGGC
4601	TCATAACACC	${\tt CCTTGTATTA}$	CTGTTTATGT	AAGCAGACAG	TTTTATTGTT
4651	CATGATGATA	TATTTTTATC	${\tt TTGTGCAATG}$	TAACATCAGA	GATTTTGAGA
4701	CACAACGTGG	CTTTCCCCCC	CCCCCATTA	TTGAAGCATT	TATCAGGGTT
4751	ATTGTCTCAT	GAGCGGATAC	ATATTTGAAT	GTATTTAGAA	AAATAAACAA
.4801	ATAGGGGTTC	CGCGCACATT	TCCCCGAAAA	GTGCCACCTG	ACGTCTAAGA
4851	AACCATTATT	ATCATGACAT	TAACCTATAA	AAATAGGCGT	ATCACGAGGC
4901	CCTTTCGTC				

1	CATCATCAAT	AATATACCTT	ATTTTGGATT	GAAGCCAATA	TGATAATGAG	GGGGTGGAGT
61	TTGTGACGTG	GCGCGGGGCG	TGGGAACGGG	GCGGGTGACG	TAGTAGTGTG	GCGGAAGTGT
121	GATGTTGTAA	GTGTGGCGGA	ACACATGTAA	GCGCCGGATG	TGGTAAAAGT	GACGTTTTTG
181	GTGTGCGCCG	GTGTACACGG	GAAGTGACAA	TTTTCGCGCG	GTTTTAGGCG	GATGTTGTAG
241	TAAATTTGGG	CGTAACCAAG	TAATATTTGG	CCATTTTCGC	GGGAAAACTG	AATAAGAGGA
301	AGTGAAATCT	GAATAATTCT	GTGTTACTCA	TAGCGCGTAA	TATTTGTCTA	GGGCCGCGGG
361	GACTTTGACC	GTTTACGTGG	AGACTCGCCC	AGGTGTTTTT	CTCAGGTGTT	TTCCGCGTTC
421	CGGGTCAAAG	TTGGCGTTTT	ATTATTATAG	${\tt TCAGCTGACG}$	CGCAGTGTAT	TTATACCCGG
481	TGAGTTCCTC	AAGAGGCCAC	TCTTGAGTGC	CAGCGAGTAG	AGTTTTCTCC	TCCGAGCCGC
541	TCCGACACCG	GGACTGAAAA	TGAGACATAT	${\tt TATCTGCCAC}$	${\tt GGAGGTGTTA}$	TTACCGAAGA
601	AATGGCCGCC	AGTCTTTTGG	ACCAGCTGAT	CGAAGAGGTA	CTGGCTGATA	ATCTTCCACC
661	TCCTAGCCAT	TTTGAACCAC	CTACCCTTCA	CGAACTGTAT	GATTTAGACG	TGACGGCCCC
721	CGAAGATCCC	AACGAGGAGG	CGGTTTCGCA	GATTTTTCCC	GAGTCTGTAA	TGTTGGCGGT
					GGTTCTCCGG	
841	CCTTTCCCGG	CAGCCCGAGC	AGCCGGAGCA	GAGAGCCTTG	GGTCCGGTTT	CTATGCCAAA
					GGCTTTCCAC	
					GAGCACCCCG	
					GATATTATGT	
					AAAATTATGG	
					TTTACAGTTT	
					AACCTGAGCC	
					TGGTGCCTGC	
					CGGATAGCTG	
					TGTGCCCCAT	
					TCGAGGACTT	
					CATAAGGTGT	
					ATGTAAGTTT	
					GGCTTAAAGG	
					CTTGGGAGTG	
					GTACCTCTTG	
					TTAAGGAGGA	
					ATTCTTTGAA	
					TTTCCACACC	
					GGAGCGAAGA	
					GGAGAGCGGT	
					TAATACCGAC	
					GCCCATGGAA	
					ACTGTTTCCA	
						GAGCGGGGG
					CTTAATGACC	
						GATCTGCTGG
2461	CGCAGAAGTA	TTCCATAGAG	CAGCTGACCA	CTTACTGGCT	GCAGCCAGGG	GATGATTTTG

FIG. 7A

2521	AGGAGGCTAT	TAGGGTATAT	GCAAAGGTGG	CACTTAGGCC	AGATTGCAAG	TACAAGATTA
2581	GCAAACTTGT	AAATATCAGG	AATTGTTGCT	ACATTTCTGG	GAACGGGGCC	GAGGTGGAGA
2641	TAGATACGGA	GGATAGGGTG	GCCTTTAGAT	GTAGCATGAT	AAATATGTGG	CCGGGGGTGC
2701	TTGGCATGGA	CGGGGTGGTT	ATTATGAATG	TGAGGTTTAC	TGGTCCCAAT	TTTAGCGGTA
2761	CGGTTTTCCT	GGCCAATACC	AATCTTATCC	TACACGGTGT	AAGCTTCTAT	GGGTTTAACA
2821	ATACCTGTGT	GGAAGCCTGG	ACCGATGTAA	GGGTTCGGGG	CTGTGCCTTT	TACTGCTGCT
2881	GGAAGGGGGT	GGTGTGTCGC	CCCAAAAGCA	GGGCTTCAAT	TAAGAAATGC	CTGTTTGAAA
2941	GGTGTACCTT	GGGTATCCTG	TCTGAGGGTA	ACTCCAGGGT	GCGCCACAAT	GTGGCCTCCG
3001	ACTGTGGTTG	CTTTATGCTA	GTGAAAAGCG	TGGCTGTGAT	TAAGCATAAC	ATGGTGTGTG
3061	GCAACTGCGA	GGACAGGGCC	TCTCAGATGC	${\tt TGACCTGCTC}$	GGACGGCAAC	TGTCACTTGC
3121	TGAAGACCAT	TCACGTAGCC	AGCCACTCTC	GCAAGGCCTG	GCCAGTGTTT	GAGCACAACA
3181	TACTGACCCG	CTGTTCCTTG	CATTTGGGTA	ACAGGAGGG	GGTGTTCCTA	CCTTACCAAT
3241	GCAATTTGAG	TCACACTAAG	ATATTGCTTG	AGCCCGAGAG	CATGTCCAAG	GTGAACCTGA
3301	ACGGGGTGTT	TGACATGACC	ATGAAGATCT	GGAAGGTGCT	GAGGTACGAT	GAGACCCGCA
3361	CCAGGTGCAG	ACCCTGCGAG	TGTGGCGGTA	AACATATTAG	GAACCAGCCT	GTGATGCTGG
3421	ATGTGACCGA	GGAGCTGAGG	CCCGATCACT	TGGTGCTGGC	CTGCACCCGC	GCTGAGTTTG
3481	GCTCTAGCGA	TGAAGATACA	GATTGAGGTA	CTGAAATGTG	TGGGCGTGGC	TTAAGGGTGG
3541	GAAAGAATAT	ATAAGGTGGG	GGTCTCATGT	AGTTTTGTAT	CTGTTTTGCA	GCAGCCGCCG
3601	CCATGAGCGC	CAACTCGTTT	GATGGAAGCA	TTGTGAGCTC	ATATTTGACA	ACGCGCATGC
3661	CCCCATGGGC	CGGGGTGCGT	CAGAATGTGA	TGGGCTCCAG	CATTGATGGT	CGCCCCGTCC
3721	TGCCCGCAAA	CTCTACTACC	TTGACCTACG	AGACCGTGTC	TGGAACGCCG	TTGGAGACTG
3781	CAGCCTCCGC	CGCCGCTTCA	GCCGCTGCAG	CCACCGCCCG	CGGGATTGTG	ACTGACTTTG
3841	CTTTCCTGAG	CCCGCTTGCA	AGCAGTGCAG	CTTCCCGTTC	ATCCGCCCGC	GATGACAAGT
3901	TGACGGCTCT	TTTGGCACAA	TTGGATTCTT	TGACCCGGGA	ACTTAATGTC	GTTTCTCAGC
3961	AGCTGTTGGA	TCTGCGCCAG	CAGGTTTCTG	CCCTGAAGGC	TTCCTCCCCT	CCCAATGCGG
4021	TTTAAAACAT	AAATAAAAAC	CAGACTCTGT	TTGGATTTGG	ATCAAGCAAG	TGTCTTGCTG
4081	TCTTTATTTA	GGGGTTTTGC	GCGCGCGGTA	GGCCCGGGAC	CAGCGGTCTC	GGTCGTTGAG
		ATTTTTTCCA				
		TCTCTGGGGT				
		ATCCAGTCGT				
		ATTGCCAGGG				
		ATACGTGGGG				
		ATATCCCTCC				
		GGAAATTTGT				
		CCTCCAAGAT				
		TGGGCGAAGA				
		TAGGCCATTT				
		GGCCCAGGGG				
		GGGATCATGT			•	
		TGGGAAGAAA				
		ACACCTATTA				
4981	ATCCCTGAGC	AGGGGGCCA	CTTCGTTAAG	CATGTCCCTG	ACTTGCATGT	TTTCCCTGAC

FIG. 7B

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5041	CAAATCCGCC	AGAAGGCGCT	CGCCGCCCAG	CGATAGCAGT	TCTTGCAAGG	AAGCAAAGTT
5101	TTTCAACGGT	TTGAGGCCGT	CCGCCGTAGG	CATGCTTTTG	AGCGTTTGAC	CAAGCAGTTC
				TACGGCATCT		
				GGCAGTAGTC		
				CTCGTCAGCG		
				GTGCGCTTGA		
				GCCAGGTAGC		
				CGCAGCTTGC		
				AGCTTGGGCG		
				ACGGTCTCGC		
				CCCCCATGCT		
				GTGACGAAAA		
5761	ACAGACTTGA	GAGGCCTGTC	CTCGAGCGGT	GTTCCGCGGT	CCTCCTCGTA	TAGAAACTCG
5821	GACCACTCTG	AGACGAAGGC	TCGCGTCCAG	GCCAGCACGA	AGGAGGCTAA	GTGGGAGGGG
				CGCTCCAGGG		
				TAGGTGTAGG		
				CGTTCGTCCT		
				TCCCTCTCAA		
				GATTTGATAT		
				GAAAAGACAA		
				AGCAACTTGG		
				GCGATGTTTA		
				CGCTCGTCGG		
				CTGGTGGCTA		
				GAGCAGAATG		
				AAGACCCCGG		
				GCCTGCTGCC		
				GGCATGGGGT		
				TCTCTGAGTA		
				TAATCGTATA		
				TGCTCTGCTC		
6901	ATGGCATGTG	AGTTGGATGA	TATGGTTGGA	CGCTGGAAGA	CGTTGAAGCT	GGCGTCTGTG
6961	AGACCTACCG	CGTCACGCAC	GAAGGAGGCG	TAGGAGTCGC	GCAGCTTGTT	GACCAGCTCG
7021	GCGGTGACCT	GCACGTCTAG	GGCGCAGTAG	TCCAGGGTTT	CCTTGATGAT	GTCATACTTA
						GTCTTTCCAG
						GTAGAACTGG
						CTGCGCGGCC
						GAGGTACTGG
						CGTGCGCTTT
						TCCCGCGCGA
						GTTAATTACC
7501	TGGGCGGCGA	GCACGATCTC	GTCAAAGCCG	TTGATGTTGT	GGCCCACAAT	GTAAAGTTCC

FIG. 7C

7561	AAGAAGCGCG	GGATGCCCTT	GATGGAAGGC	AATTTTTAA	GTTCCTCGTA	GGTGAGCTCT
	TCAGGGGAGC					
	ACGAATGAGC					
	AACTGGCGAC					
	TCCCAGCGGT					
7861	TCTCCGCCGA	ACTTCATGAC	CAGCATGAAG	GGCACGAGCT	GCTTCCCAAA	GGCCCCCATC
7921	CAAGTATAGG	TCTCTACATC	GTAGGTGACA	AAGAGACGCT	CGGTGCGAGG	ATGCGAGCCG
7981	ATCGGGAAGA	ACTGGATCTC	CCGCCACCAG	TTGGAGGAGT	GGCTGTTGAT	GTGGTGAAAG
8041	TAGAAGTCCC	TGCGACGGGC	CGAACACTCG	TGCTGGCTTT	TGTAAAAACG	TGCGCAGTAC
8101	TGGCAGCGGT	GCACGGGCTG	TACATCCTGC	ACGAGGTTGA	CCTGACGACC	GCGCACAAGG
8161	AAGCAGAGTG	GGAATTTGAG	CCCCTCGCCT	GGCGGGTTTG	GCTGGTGGTC	TTCTACTTCG
8221	GCTGCTTGTC	CTTGACCGTC	TGGCTGCTCG	AGGGGAGTTA	CGGTGGATCG	GACCACCACG
8281	CCGCGCGAGC	CCAAAGTCCA	GATGTCCGCG	CGCGGCGGTC	GGAGCTTGAT	GACAACATCG
8341	CGCAGATGGG	AGCTGTCCAT	GGTCTGGAGC	TCCCGCGGGG	TCAGGTCAGG	CGGGAGCTCC
8401	TGCAGGTTTA	CCTCGCATAG	CCGGGTCAGG	GCGCGGCTA	GGTCCAGGTG	ATACCTGATT
8461	TCCAGGGGCT	GGTTGGTGGC	GGCGTCGATG	GCTTGCAAGA	GGCCGCATCC	CCGCGGCGCG
8521	ACTACGGTAC	CGCGCGGCGG	GCGGTGGGCC	GCGGGGGTGT	CCTTGGATGA	TGCATCTAAA
8581	AGCGGTGACG	CGGGCGGCC	CCCGGAGGTA	GGGGGGCTC	GGGACCCGCC	GGGAGAGGGG
	GCAGGGGCAC				•	
8701	CGAACGCGAC	GACGCGGCGG	TTGATCTCCT	GAATCTGGCG	CCTCTGCGTG	AAGACGACGG
8761	GCCCGGTGAG	CTTGAACCTG	AAAGAGAGTT	CGACAGAATC	AATTTCGGTG	TCGTTGACGG
8821	CGGCCTGGCG	CAAAATCTCC	TGCACGTCTC	CTGAGTTGTC	TTGATAGGCG	ATCTCGGCCA
8881	TGAACTGCTC	GATCTCTTCC	TCCTGGAGAT	CTCCGCGTCC	GGCTCGCTCC	ACGGTGGCGG
8941	CGAGGTCGTT	GGAGATGCGG	GCCATGAGCT	GCGAGAAGGC	GTTGAGGCCT	CCCTCGTTCC
9001	AGACGCGGCT	GTAGACCACG	CCCCCTTCGG	CATCGCGGGC	GCGCATGACC	ACCTGCGCGA
	GATTGAGCTC					
	TGAGGGTGGT					
	ATTCGTTGAT					
9241	AGTTGAAAAA	CTGGGAGTTG	CGCGCCGACA	CGGTTAACTC	CTCCTCCAGA	AGACGGATGA
	GCTCGGCGAC					
	CAATCTCCTC					
	GGACACGGCG					
	CGCGGCGACG					
	AGACGCCGCC					
	CGGCGCTAAC					
	GCGAGTCCGC					
	CGCAAGGTAG					
	CGGAGGTGCT					
	GAAGCACCAT					
	CTTCGTTTTG					
	CTTCTTCTTC					
10021	AGTTTGGCCG	TAGGTGGCGC	CCTCTTCCTC	CCATGCGTGT	GACCCCGAAG	CCCCTCATCG

FIG. 7D

10081	GCTGAAGCAG	GGCCAGGTCG	GCGACAACGC	GCTCGGCTAA	TATGGCCTGC	TGCACCTGCG
10141	TGAGGGTAGA	CTGGAAGTCG	TCCATGTCCA	CAAAGCGGTG	GTATGCGCCC	GTGTTGATGG
10201	TGTAAGTGCA	GTTGGCCATA	ACGGACCAGT	TAACGGTCTG	GTGACCCGGC	TGCGAGAGCT
10261	CGGTGTACCT	GAGACGCGAG	TAAGCCCTTG	AGTCAAAGAC	GTAGTCGTTG	CAAGTCCGCA
10321	CCAGGTACTG	GTATCCCACC	AAAAAGTGCG	GCGGCGGCTG	GCGGTAGAGG	GGCCAGCGTA
10381	GGGTGGCCGG	GGCTCCGGGG	GCGAGGTCTT	CCAACATAAG	GCGATGATAT	CCGTAGATGT
10441	ACCTGGACAT	CCAGGTGATG	CCGGCGGCGG	TGGTGGAGGC	GCGCGGAAAG	TCACGGACGC
10501	GGTTCCAGAT	GTTGCGCAGC	GGCAAAAAGT	GCTCCATGGT	CGGGACGCTC	TGGCCGGTCA
10561	GGCGCGCGCA	GTCGTTGACG	CTCTAGACCG	TGCAAAAGGA	GAGCCTGTAA	GCGGGCACTC
10621	TTCCGTGGTC	TGGTGGATAA	ATTCGCAAGG	GTATCATGGC	GGACGACCGG	GGTTCGAACC
10681	CCGGATCCGG	CCGTCCGCCG	TGATCCATGC	GGTTACCGCC	CGCGTGTCGA	ACCCAGGTGT
10741	GCGACGTCAG	ACAACGGGGG	AGCGCTCCTT	TTGGCTTCCT	TCCAGGCGCG	GCGGATGCTG
10801	CGCTAGCTTT	TTTGGCCACT	GGCCGCGCGC	GGCGTAAGCG	GTTAGGCTGG	AAAGCGAAAG
10861	CATTAAGTGG	CTCGCTCCCT	GTAGCCGGAG	GGTTATTTTC	CAAGGGTTGA	GTCGCGGGAC
10921	CCCCGGTTCG	AGTCTCGGGC	CGGCCGGACT	GCGGCGAACG	GGGGTTTGCC	TCCCCGTCAT
10981	GCAAGACCCC	GCTTGCAAAT	TCCTCCGGAA	ACAGGGACGA	GCCCCTTTTT	TGCTTTTCCC
11041	AGATGCATCC	GGTGCTGCGG	CAGATGCGCC	CCCCTCCTCA	GCAGCGGCAA	GAGCAAGAGC
11101	AGCGGCAGAC	ATGCAGGGCA	CCCTCCCCTT	CTCCTACCGC	GTCAGGAGGG	GCAACATCCG
11161	CGGCTGACGC	GGCGGCAGAT	GGTGATTACG	AACCCCCGCG	GCGCCGGACC	CGGCACTACT
11221	TGGACTTGGA	GGAGGGCGAG	GGCCTGGCGC	GGCTAGGAGC	GCCCTCTCCT	GAGCGACACC
11281	CAAGGGTGCA	GCTGAAGCGT	GACACGCGCG	AGGCGTACGT	GCCGCGGCAG	AACCTGTTTC
11341	GCGACCGCGA	GGGAGAGGAG	CCCGAGGAGA	TGCGGGATCG	AAAGTTCCAT	GCAGGGCGCG
11401	AGTTGCGGCA	TGGCCTGAAC	CGCGAGCGGT	TGCTGCGCGA	GGAGGACTTT	GAGCCCGACG
11461	CGCGGACCGG	GATTAGTCCC	GCGCGCGCAC	ACGTGGCGGC	CGCCGACCTG	GTAACCGCGT
11521	ACGAGCAGAC	GGTGAACCAG	GAGATTAACT	TTCAAAAAAG	CTTTAACAAC	CACGTGCGCA
11581	CGCTTGTGGC	GCGCGAGGAG	GTGGCTATAG	GACTGATGCA	TCTGTGGGAC	TTTGTAAGCG
11641	CGCTGGAGCA	AAACCCAAAT	AGCAAGCCGC	TCATGGCGCA	GCTGTTCCTT	ATAGTGCAGC
11701	ACAGCAGGGA	CAACGAGGCA	TTCAGGGATG	CGCTGCTAAA	CATAGTAGAG	CCCGAGGGCC
11761	GCTGGCTGCT	CGATTTGATA	AACATTCTGC	AGAGCATAGT	GGTGCAGGAG	CGCAGCTTGA
11821	GCCTGGCTGA	CAAGGTGGCC	GCCATTAACT	ATTCCATGCT	CAGTCTGGGC	AAGTTTTACG
11881	CCCGCAAGAT	ATACCATACC	CCTTACGTTC	CCATAGACAA	GGAGGTAAAG	ATCGAGGGGT
11941	TCTACATGCG	CATGGCGCTG	AAGGTGCTTA	CCTTGAGCGA	CGACCTGGGC	GTTTATCGCA
12001	ACGAGCGCAT	CCACAAGGCC	GTGAGCGTGA	GCCGGCGGCG	CGAGCTCAGC	GACCGCGAGC
12061	TGATGCACAG	CCTGCAAAGG	GCCCTGGCTG	GCACGGGCAG	CGGCGATAGA	GAGGCCGAGT
12121	CCTACTTTGA	CGCGGGCGCT	GACCTGCGCT	GGGCCCCAAG	CCGACGCGCC	CTGGAGGCAG
12181	CTGGGGCCGG	ACCTGGGCTG	GCGGTGGCAC	CCGCGCGCGC	TGGCAACGTC	GGCGGCGTGG
12241	AGGAATATGA	CGAGGACGAT	GAGTACGAGC	CAGAGGACGG	CGAGTACTAA	GCGGTGATGT
12301	TTCTGATCAG	ATGATGCAAG	ACGCAACGGA	CCCGGCGGTG	CGGGCGGCGC	TGCAGAGCCA
12361	GCCGTCCGGC	CTTAACTCCA	CGGACGACTG	GCGCCAGGTC	ATGGACCGCA	TCATGTCGCT
12421	GACTGCGCGC	AACCCTGACG	CGTTCCGGCA	GCAGCCGCAG	GCCAACCGGC	TCTCCGCAAT
12481	TCTGGAAGCG	GTGGTCCCGG	CGCGCGCAAA	CCCCACGCAC	GAGAAGGTG	TGGCGATCGT
12541	AAACGCGCTG	GCCGAAAACA	GGGCCATCCG	GCCCGATGAC	GCCGGCCTGC	TCTACGACGC

FIG. 7E

		_				
	GCTGCTTCAG					
	GGTGGGGGAT					
	GGGCTCCATG					
	ACAGGAGGAC					
	AAGTGAGGTG					
	GACCGTAAAC					
	CACAGGCGAC					
	GCTAATAGCG					
13081	GCTGACACTG	TACCGCGAGG	CCATAGGTCA	GGCGCATGTG	GACGAGCATA	CTTTCCAGGA
13141	GATTACAAGT	GTTAGCCGCG	CGCTGGGGCA	GGAGGACACG	GGCAGCCTGG	AGGCAACCCT
13201	GAACTACCTG	CTGACCAACC	GGCGGCAAAA	AATCCCCTCG	TTGCACAGTT	TAAACAGCGA
13261	GGAGGAGCGC	ATTTTGCGCT	ATGTGCAGCA	GAGCGTGAGC	CTTAACCTGA	TGCGCGACGG
13321	GGTAACGCCC	AGCGTGGCGC	TGGACATGAC	CGCGCGCAAC	ATGGAACCGG	GCATGTATGC
13381	CTCAAACCGG	CCGTTTATCA	ATCGCCTAAT	GGACTACTTG	CATCGCGCGG	CCGCCGTGAA
13441	CCCCGAGTAT	TTCACCAATG	CCATCTTGAA	CCCGCACTGG	CTACCGCCCC	CTGGTTTCTA
13501	CACCGGGGGA	TTCGAGGTGC	CCGAGGGTAA	${\tt CGATGGATTC}$	${\tt CTCTGGGACG}$	ACATAGACGA
13561	CAGCGTGTTT	TCCCCGCAAC	CGCAGACCCT	${\tt GCTAGAGTTG}$	CAACAACGCG	AGCAGGCAGA
13621	GGCGGCGCTG	CGAAAGGAAA	GCTTCCGCAG	GCCAAGCAGC	${\tt TTGTCCGATC}$	TAGGCGCTGC
13681	GGCCCCGCGG	TCAGATGCTA	GTAGCCCATT	TCCAAGCTTG	ATAGGGTCTC	TTACCAGCAC
13741	TCGCACCACC	CGCCCGCGCC	TGCTGGGCGA	GGAGGAGTAC	CTAAACAACT	CGCTGCTGCA
13801	GCCGCAGCGC	GAAAAGAACC	TGCCTCCGGC	GTTTCCCAAC	AACGGGATAG	AGAGCCTAGT
13861	GGACAAGATG	ÂGTAGATGGA	AGACGTATGC	GCAGGAGCAC	AGGGATGTGC	CCGGCCCGCG
13921	CCCGCCCACC	CGTCGTCAAA	GGCACGACCG	TCAGCGGGGT	CTGGTGTGGG	AGGACGATGA
13981	CTCGGCAGAC	GACAGCAGCG	TCTTGGATTT	GGGAGGGAGT	GGCAACCCGT	TTGCACACCT
14041	TCGCCCCAGG	CTGGGGAGAA	TGTTTTAAAA	AAAGCATGAT	GCAAAATAAA	AAACTCACCA
14101	AGGCCATGGC	ACCGAGCGTT	GGTTTTCTTG	TATTCCCCTT	AGTATGCGGC	GCGCGGCGAT
14161	GTATGAGGAA	GGTCCTCCTC	CCTCCTACGA	GAGCGTGGTG	AGCGCGGCGC	CAGTGGCGGC
14221	GGCGCTGGGT	TCACCCTTCG	ATGCTCCCCT	GGACCCGCCG	TTCGTGCCTC	CGCGGTACCT
-	GCGGCCTACC					
14341	CACCCGTGTG	TACCTTGTGG	ACAACAAGTC	AACGGATGTG	GCATCCCTGA	ACTACCAGAA
	CGACCACAGC					
	AAGCACACAG					
	CCTGCATACC					
	GGTGATGGTG					
	GGAGTTCACG					
	CGCGATCGTG					
	CGGGGTAAAG					
	CATGCCTGGG					
	CGGGGTGGAC					
	CTTCCAGGAG					
	GTTGGATGTG					
	CGCAGGCGGC					
T200T	CGCAGGCGGC	GGCAACAACA	C. GGCAGCGG	COCOGIRIONG		20001100100

FIG. 7F

						_
		CCGGTGGAGG				
		GAGAAGCGCG				
		CAACCCGAGG				
		AAGAAACGCA				
		TACCTTGCAT				
15421	CCTGCTTTGC	ACTCCTGACG	TAACCTGCGG	CTCGGAGCAG	GTATACTGGT	CGTTGCCCGA
15481	CATGATGCAA	GACCCCGTGA	${\tt CCTTCCGCTC}$	CACGCGCCAG	ATCAGCAACT	TTCCGGTGGT
		CTGTTGCCCG				
15601	CCAGCTCATC	CGCCAGTTTA	CCTCTCTGAC	CCACGTGTTC	AATCGCTTTC	CCGAGAACCA
15661	GATTTTGGCG	CGCCCGCCAG	CCCCCACCAT	CACCACCGTC	AGTGAAAACG	TTCCTGCTCT
15721	CACAGATCAC	GGGACGCTAC	CGCTGCGCAA	CAGCATCGGA	GGAGTCCAGC	GAGTGACCAT
15781	TACTGACGCC	AGACGCCGCA	CCTGCCCCTA	CGTTTACAAG	GCCCTGGGCA	TAGTCTCGCC
15841	GCGCGTCCTA	TCGAGCCGCA	CTTTTTGAGC	AAGCATGTCC	ATCCTTATAT	CGCCCAGCAA
15901	TAACACAGGC	TGGGGCCTGC	GCTTCCCAAG	CAAGATGTTT	GGCGGGGCCA	AGAAGCGCTC
15961	CGACCAACAC	CCAGTGCGCG	TGCGCGGGCA	CTACCGCGCG	CCCTGGGGCG	CGCACAAACG
16021	CGGCCGCACT	GGGCGCACCA	CCGTCGATGA	CGCCATCGAC	GCGGTGGTGG	AGGAGGCGCG
		CCCACGCCGC				
16141	GCGCGGAGCC	CGGCGCTACG	CTAAAATGAA	GAGACGGCGG	AGGCGCGTAG	CACGTCGCCA
		CCCGGCACTG				
16261	TCGCACCGGC	CGACGGGCGG	CCATGCGAGC	CGCTCGAAGG	CTGGCCGCGG	GTATTGTCAC
16321	TGTGCCCCCC	AGGTCCAGGC	GACGAGCGGC	CGCCGCAGCA	GCCGCGGCCA	TTAGTGCTAT
16381	GACTCAGGGT	CGCAGGGGCA	ACGTGTACTG	GGTGCGCGAC	TCGGTTAGCG	GCCTGCGCGT
16441	GCCCGTGCGC	ACCCGCCCCC	CGCGCAACTA	GATTGCAATA	AAAAACTACT	TAGACTCGTA
16501	CTGTTGTATG	TATCCAGCGG	CGGCGCGCG	CATCGAAGCT	ATGTCCAAGC	GCAAAATCAA
16561	AGAAGAGATG	CTCCAGGTCA	TCGCGCCGGA	GATCTATGGC	CCCCGAAGA	AGGAAGAGCA
16621	GGATTACAAG	CCCCGAAAGC	TAAAGCGGGT	CAAAAAGAAA	AAGAAAGATG	ATGATGATGA
16681	TGAACTTGAC	GACGAGGTGG	AACTGTTGCA	CGCGACCGCG	CCCAGGCGAC	GGGTACAGTG
16741	GAAAGGTCGA	CGCGTAAGAC	GTGTTTTGCG	ACCCGGCACC	ACCGTAGTCT	TTACGCCCGG
16801	TGAGCGCTCC	ACCCGCACCT	ACAAGCGCGT	GTATGATGAG	GTGTACGGCG	ACGAGGACCT
16861	GCTTGAGCAG	GCCAACGAGC	GCCTCGGGGA	GTTTGCCTAC	GGAAAGCGGC	ATAAGGACAT
16921	GCTGGCGTTG	CCGCTGGACG	AGGGCAACCC	AACACCTAGC	CTAAAGCCCG	TGACACTGCA
16981	GCAGGTGCTG	CCCGCGCTTG	CACCGTCCGA	AGAAAAGCGC	GGCCTAAAGC	GCGAGTCTGG
17041	TGACTTGGCA	CCCACCGTGC	AGCTGATGGT	ACCCAAGCGT	CAGCGACTGG	AAGATGTCTT
17101	GGAAAAAATG	ACCGTGGAGC	CTGGGCTGGA	GCCCGAGGTC	CGCGTGCGGC	CAATCAAGCA
17161	GGTGGCACCG	GGACTGGGCG	TGCAGACCGT	GGACGTTCAG	ATACCCACCA	CCAGTAGCAC
17221	TAGTATTGCC	ACTGCCACAG	AGGGCATGGA	GACACAAACG	TCCCCGGTTG	CCTCGGCGGT
17281	GGCAGATGCC	GCGGTGCAGG	CGGCCGCTGC	GGCCGCGTCC	AAGACCTCTA	CGGAGGTGCA
						CAAGGAAGTA
		AGCGCGCTAC				
						GCCGAACCAC
						TTTCCGTGCG
17581	CAGGGTGGCT	CGCGAAGGAG	GCAGGACCCT	GGTGCTGCCA	ACAGCGCGCT	ACCACCCCAG

FIG. 7G

				maa.a.m.m.	CCCCCC A CCC	CCCCCCTCCC
	CATCGTTTAA					
	TTTCCCGGTG					
	CCTGACGGGC					
	GCGCGGCGGT					
	CGGAATTGCA					
	GAAAAATCAA					
	. AATGGAAGAC					
	AAACTGGCAA					
	. GTGGAGCGGC					
	. CAGCAGCACA					
	. GGTAGATGGC					
	AAATAAGATT					
	GGAGACAGTG					
	TCTGGTGACG					
	CACCACCCGT					
18541	GCTGGACCTG	CCTCCCCCG	CCGACACCCA	GCAGAAACCT	GTGCTGCCAG	GCCCGTCCGC
18601	CGTTGTTGTA	ACCCGTCCTA	GCCGCGCGTC	CCTGCGCCGC	GCCGCCAGCG	GTCCGCGATC
18661	GTTGCGGCCC	GTAGCCAGTG	GCAACTGGCA	AAGCACACTG	AACAGCATCG	TGGGTTTGGG
18721	GGTGCAATCC	CTGAAGCGCC	GACGATGCTT	CTGATAGCTA	ACGTGTCGTA	TGTGTGTCAT
18781	GTATGCGTCC	ATGTCGCCGC	CAGAGGAGCT	GCTGAGCCGC	CGCGCGCCCG	CTTTCCAAGA
18841	TGGCTACCCC	TTCGATGATG	CCGCAGTGGT	CTTACATGCA	CATCTCGGGC	CAGGACGCCT
18901	CGGAGTACCT	GAGCCCCGGG	${\tt CTGGTGCAGT}$	TCGCCCGCGC	CACCGAGACG	TACTTCAGCC
18961	TGAATAACAA	GTTTAGAAAC	CCCACGGTGG	CGCCTACGCA	CGACGTGACC	ACAGACCGGT
19021	CTCAGCGTTT	GACGCTGCGG	TTCATCCCCG	TGGACCGCGA	GGATACTGCG	TACTCGTACA
19081	AGGCGCGGTT	CACCCTAGCT	GTGGGTGATA	ACCGTGTGCT	AGACATGGCT	TCCACGTACT
19141	TTGACATCCG	CGGCGTGCTG	GACAGGGGCC	CTACTTTTAA	GCCCTACTCT	GGCACTGCCT
19201	ACAACGCACT	GGCCCCCAAG	GGTGCCCCCA	ACTCGTGCGA	GTGGGAACAA	AATGAAACTG
19261	CACAAGTGGA	TGCTCAAGAA	CTTGACGAAG	AGGAGAATGA	AGCCAATGAA	GCTCAGGCGC
19321	GAGAACAGGA	ACAAGCTAAG	AAAACCCATG	TATATGCCCA	GGCTCCACTG	TCCGGAATAA
19381	AAATAACTAA	AGAAGGTCTA	CAAATAGGAA	CTGCCGACGC	CACAGTAGCA	GGTGCCGGCA
19441	AAGAAATTTT	CGCAGACAAA	ACTTTTCAAC	CTGAACCACA	AGTAGGAGAA	TCTCAATGGA
19501	ACGAAGCGGA	TGCCACAGCA	GCTGGTGGAA	GGGTTCTTAA	AAAGACAACT	CCCATGAAAC
19561	CCTGCTATGG	CTCATACGCT	AGACCCACCA	ATTCCAACGG	CGGACAGGGC	GTTATGGTTG
19621	AACAAAATGG	TAAATTGGAA	AGTCAAGTCG	AAATGCAATT	TTTTTCCACA	TCCACAAATG
19681	CCACAAATGA	AGTTAACAAT	ATACAACCAA	CAGTTGTATT	GTACAGCGAA	GATGTAAACA
19741	TGGAAACTCC	AGATACTCAT	СТТТСТТАТА	ААССТААААТ	GGGGGATAAA	AATGCCAAAG
19801	TCATGCTTGG	ACAACAAGCA	ATGCCAAACA	GACCAAATTA	CATTGCTTTT	AGAGACAATT
19861	TTATTGGTCT	CATGTATTAC	AACAGCACAG	GTAACATGGG	TGTCCTTGCT	GGTCAGGCAT
19921	CGCAGTTGAA	CGCTGTTGTA	GATTTGCAAG	ACAGAAACAC	AGAGCTGTCC	TACCAGCTTT
19981	TGCTTGATTC	AATTGGCGAC	AGAACAAGAT	ACTTTTCAAT	GTGGAATCAA	GCTGTTGACA
	GCTATGATCC					
	ATTGCTTTCC					

FIG. 7H

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		GGACCAAGGC				
		GGTGGGAAAT				
		TTACTCCAAT				
		AATATCTGAC				
		TGTAGACTGC				
20461	ACAACGTTAA	TCCCTTTAAC	CACCACCGCA	ATGCGGGCCT	GCGTTACCGC	TCCATGTTGT
20521	TGGGAAACGG	CCGCTACGTG	CCCTTTCACA	TTCAGGTGCC	CCAAAAGTTT	TTTGCCATTA
		CCTCCTGCCA				
20641	ACATGGTTCT	GCAGAGCTCT	CTGGGAAACG	ACCTTAGAGT	TGACGGGGCT	AGCATTAAGT
20701	TTGACAGCAT	TTGTCTTTAC	GCCACCTTCT	TCCCCATGGC	CCACAACACG	GCCTCCACGC
		GCTCAGAAAT				
		ATATCCCATA				
20881	GCAACTGGGC	AGCATTTCGC	GGTTGGGCCT	TCACACGCTT	GAAGACAAAG	GAAACCCCTT
20941	CCCTGGGATC	AGGCTACGAC	CCTTACTACA	CCTACTCTGG	CTCCATACCA	TACCTTGACG
		TCTTAATCAC				
		CAACGACCGC				
		CTATAACGTA				
		CTACAATATT				
		GTTCTTCAGA				
		TTATCAGCAG				
21361	GCTACCTCGC	TCCCACCATG	CGCGAGGGAC	AAGCTTACCC	CGCTAATGTT	CCCTACCCAC
		AACCGCGGTT				
		CCCCTTCTCC				
		CTACGCAAAC				
21601	TGGACGAGCC	CACCCTTCTT	TATGTTTTGT	TTGAAGTCTT	TGACGTGGTC	CGTGTGCACC
		CGGCGTCATC				
		AAGAAGCAAG				
		GCCATTGTCA				
		CCAGGCTTTG				
		ACTGGGGGCG				
		TTTGAGCCCT				
		TCACTCCTGC				
22081	GCTGGAAAAG	TCCACCCAAA	GCGTGCAGGG	GCCCAACTCG	GCCGCCTGTG	GCCTATTCTG
22141	CTGCATGTTT	CTCCACGCCT	TTGCCAACTG	GCCCCAAACT	CCCATGGATC	ACAACCCCAC
22201	CATGAACCTT	ATTACCGGGG	TACCCAACTC	CATGCTTAAC	AGTCCCCAGG	TACAGCCCAC
22261	CCTGCGCCGC	AACCAGGAAC	AGCTCTACAG	CTTCCTGGAG	CGCCACTCGC	CCTACTTCCG
22321	CAGCCACAGT	GCGCAAATTA	GGAGCGCCAC	TTCTTTTTGT	CACTTGAAAA	ACATGTAAAA
22381	ATAATGTACT	AGGAGACACT	TTCAATAAAG	GCAAATGTTT	TTATTTGTAC	ACTCTCGGGT
22441	GATTATTTAC	CCCCACCCTT	GCCGTCTGCG	CCGTTTAAAA	ATCAAAGGGG	TTCTGCCGCG
22501	CATCGCTATG	CGCCACTGGC	AGGGACACGT	TGCGATACTG	GTGTTTAGTG	CTCCACTTAA
22561	ACTCAGGCAC	AACCATCCGC	GGCAGCTCGG	TGAAGTTTTC	ACTCCACAGG	CTGCGCACCA
22621	TCACCAACGO	GTTTAGCAGG	TCGGGCGCCG	ATATCTTGAA	GTCGCAGTTG	GGGCCTCCGC

FIG. 7I

22681	CCTGCGCGCG	CGAGTTGCGA	TACACAGGGT	TACAGCACTG	GAACACTATC	AGCGCCGGGT
22741	GGTGCACGCT	GGCCAGCACG	CTCTTGTCGG	AGATCAGATC	CGCGTCCAGG	TCCTCCGCGT
22801	TGCTCAGGGC	GAACGGAGTC	AACTTTGGTA	GCTGCCTTCC	CAAAAAGGGT	GCATGCCCAG
22861	GCTTTGAGTT	GCACTCGCAC	CGTAGTGGCA	TCAGAAGGTG	ACCGTGCCCA	GTCTGGGCGT
22921	TAGGATACAG	CGCCTGCATG	AAAGCCTTGA	TCTGCTTAAA	AGCCACCTGA	GCCTTTGCGC
22981	CTTCAGAGAA	GAACATGCCG	CAAGACTTGC	CGGAAAACTG	ATTGGCCGGA	CAGGCCGCGT
23041	CATGCACGCA	GCACCTTGCG	TCGGTGTTGG	AGATCTGCAC	CACATTTCGG	CCCCACCGGT
23101	TCTTCACGAT	CTTGGCCTTG	CTAGACTGCT	CCTTCAGCGC	GCGCTGCCCG	TTTTCGCTCG
23161	TCACATCCAT	TTCAATCACG	TGCTCCTTAT	TTATCATAAT	GCTCCCGTGT	AGACACTTAA
23221	GCTCGCCTTC	GATCTCAGCG	CAGCGGTGCA	GCCACAACGC	GCAGCCCGTG	GGCTCGTGGT
23281	GCTTGTAGGT	TACCTCTGCA	AACGACTGCA	GGTACGCCTG	CAGGAATCGC	CCCATCATCG
23341	TCACAAAGGT	CTTGTTGCTG	GTGAAGGTCA	GCTGCAACCC	GCGGTGCTCC	TCGTTTAGCC
23401	AGGTCTTGCA	TACGGCCGCC	AGAGCTTCCA	CTTGGTCAGG	CAGTAGCTTG	AAGTTTGCCT
23461	TTAGATCGTT	ATCCACGTGG	TACTTGTCCA	TCAACGCGCG	CGCAGCCTCC	ATGCCCTTCT
23521	CCCACGCAGA	CACGATCGGC	AGGCTCAGCG	GGTTTATCAC	CGTGCTTTCA	CTTTCCGCTT
23581	CACTGGACTC	TTCCTTTTCC	TCTTGCATCC	GCATACCCCG	CGCCACTGGG	TCGTCTTCAT
23641	TCAGCCGCCG	CACCGTGCGC	TTACCTCCCT	TGCCGTGCTT	GATTAGCACC	GGTGGGTTGC
23701	TGAAACCCAC	CATTTGTAGC	GCCACATCTT	CTCTTTCTTC	CTCGCTGTCC	ACGATCACCT
23761	CTGGGGATGG	CGGGCGCTCG	GGCTTGGGAG	AGGGGCGCTT	CTTTTTCTTT	TTGGACGCAA
23821	TGGCCAAATC	CGCCGTCGAG	GTCGATGGCC	GCGGGCTGGG	TGTGCGCGGC	ACCAGCGCAT
23881	CTTGTGACGA	GTCTTCTTCG	TCCTCGGACT	CGAGACGCCG	CCTCAGCCGC	TTTTTTGGGG
23941	GCGCGCGGG	AGGCGGCGGC	GACGGCGACG	GGGACGAGAC	GTCCTCCATG	GTTGGTGGAC
24001	GTCGCGCCGC	ACCGCGTCCG	CGCTCGGGGG	TGGTTTCGCG	CTGCTCCTCT	TCCCGACTGG
24061	CCATTTCCTT	CTCCTATAGG	CAGAAAAAGA	TCATGGAGTC	AGTCGAGAAG	GAGGACAGCC
24121	TAACCGCCCC	CTTTGAGTTC	GCCACCACCG	CCTCCACCGA	TGCCGCCAAC	GCGCCTACCA
24181	CCTTCCCCGT	CGAGGCACCC	CCGCTTGAGG	AGGAGGAAGT	GATTATCGAG	CAGGACCCAG
24241	GTTTTGTAAG	CGAAGACGAC	GAAGATCGCT	CAGTACCAAC	AGAGGATAAA	AAGCAAGACC
24301	AGGACGACGC	AGAGGCAAAC	GAGGAACAAG	TCGGGCGGGG	GGACCAAAGG	CATGGCGACT
24361	ACCTAGATGT	GGGAGACGAC	GTGCTGTTGA	AGCATCTGCA	GCGCCAGTGC	GCCATTATCT
24421	GCGACGCGTT	GCAAGAGCGC	AGCGATGTGC	CCCTCGCCAT	AGCGGATGTC	AGCCTTGCCT
24481	ACGAACGCCA	CCTGTTCTCA	CCGCGCGTAC	CCCCCAAACG	CCAAGAAAAC	GGCACATGCG
24541	AGCCCAACCC	GCGCCTCAAC	TTCTACCCCG	TATTTGCCGT	GCCAGAGGTG	CTTGCCACCT
24601	ATCACATCTT	TTTCCAAAAC	TGCAAGATAC	CCCTATCCTG	CCGTGCCAAC	CGCAGCCGAG
24661	CGGACAAGCA	GCTGGCCTTG	CGGCAGGGCG	CTGTCATACC	TGATATCGCC	TCGCTCGACG
24721	AAGTGCCAAA	AATCTTTGAG	GGTCTTGGAC	GCGACGAGAA	GCGCGCGCA	AACGCTCTGC
24781	AACAAGAAAA	CAGCGAAAAT	GAAAGTCACT	GTGGAGTGCT	GGTGGAACTT	GAGGGTGACA
24841	ACGCGCGCCT	AGCCGTGCTG	AAACGCAGCA	TCGAGGTCAC	CCACTTTGCC	TACCCGGCAC
24901	TTAACCTACC	CCCCAAGGTT	ATGAGCACAG	TCATGAGCGA	GCTGATCGTG	CGCCGTGCAC
24961	GACCCCTGGA	GAGGGATGCA	AACTTGCAAG	AACAAACCGA	GGAGGGCCTA	CCCGCAGTTG
25021	GCGATGAGCA	GCTGGCGCGC	TGGCTTGAGA	CGCGCGAGCC	TGCCGACTTG	GAGGAGCGAC
25081	GCAAGCTAAT	GATGGCCGCA	GTGCTTGTTA	CCGTGGAGCT	TGAGTGCATG	CAGCGGTTCT
25141	TTGCTGACCC	GGAGATGCAG	CGCAAGCTAG	AGGAAACGTT	GCACTACACC	TTTCGCCAGG

FIG. 7J

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25201	GCTACGTGCG	CCAGGCCTGC	ААААТТТССА	ACGTGGAGCT	CTGCAACCTG	GTCTCCTACC
25261	TTGGAATTTT	GCACGAAAAC	CGCCTTGGGC	AAAACGTGCT	TCATTCCACG	CTCAAGGGCG
25321	AGGCGCGCCG	CGACTACGTC	CGCGACTGCG	TTTACTTATT	TCTGTGCTAC	ACCTGGCAAA
25381	CGGCCATGGG	CGTGTGGCAG	CAGTGCCTGG	AGGAGCGCAA	CCTGAAGGAG	CTGCAGAAGC
		AAACTTGAAG				
		CATTATCTTC				
25561	ACTTCACCAG	TCAAAGCATG	TTGCAAAACT	TTAGGAACTT	TATCCTAGAG	CGTTCAGGAA
25621	TTCTGCCCGC	CACCTGCTGT	GCGCTTCCTA	GCGACTTTGT	GCCCATTAAG	TACCGTGAAT
25681	GCCTCCGCC	GCTTTGGGGT	CACTGCTACC	TTCTGCAGCT	AGCCAACTAC	CTTGCCTACC
25741	ACTCCGACAT	CATGGAAGAC	GTGAGCGGTG	ACGGCCTACT	GGAGTGTCAC	TGTCGCTGCA
		CCCGCACCGC				
25861	TTATCGGTAC	CTTTGAGCTG	CAGGGTCCCT	CGCCTGACGA	AAAGTCCGCG	GCTCCGGGGT
25921	TGAAACTCAC	TCCGGGGCTG	TGGACGTCGG	${\tt CTTACCTTCG}$	CAAATTTGTA	CCTGAGGACT
25981	ACCACGCCCA	CGAGATTAGG	TTCTACGAAG	ACCAATCCCG	CCCGCCAAAT	GCGGAGCTTA
26041	CCGCCTGCGT	CATTACCCAG	GGCCACATCC	${\tt TTGGCCAATT}$	GCAAGCCATT	AACAAAGCCC
		TCTGCTACGA				
26161	AGCTCAACCC	AATCCCCCCG	CCGCCGCAGC	CCTATCAGCA	GCCGCGGGCC	CTTGCTTCCC
26221	AGGATGGCAC	CCAAAAAGAA	GCTGCAGCTG	CCGCCGCCGC	CACCCACGGA	CGAGGAGGAA
26281	TACTGGGACA	GTCAGGCAGA	GGAGGTTTTG	GACGAGGAGG	AGGAGATGAT	GGAAGACTGG
26341	GACAGCCTAG	ACGAGGAAGC	TTCCGAGGCC	GAAGAGGTGT	CAGACGAAAC	ACCGTCACCC
26401	TCGGTCGCAT	TCCCCTCGCC	GGCGCCCCAG	AAATCGGCAA	CCGTTCCCAG	CATTGCTACA
26461	ACCTCCGCTC	CTCAGGCGCC	GCCGGCACTG	CCCGTTCGCC	GACCCAACCG	TAGATGGGAC
26521	ACCACTGGAA	CCAGGGCCGG	TAAGTCTAAG	CAGCCGCCGC	CGTTAGCCCA	AGAGCAACAA
26581	CAGCGCCAAG	GCTACCGCTC	GTGGCGCGTG	CACAAGAACG	CCATAGTTGC	TTGCTTGCAA
26641	GACTGTGGGG	GCAACATCTC	CTTCGCCCGC	CGCTTTCTTC	TCTACCATCA	CGGCGTGGCC
26701	TTCCCCCGTA	ACATCCTGCA	TTACTACCGT	CATCTCTACA	GCCCCTACTG	CACCGGCGGC
26761	AGCGGCAGCA	ACAGCAGCGG	CCACGCAGAA	GCAAAGGCGA	CCGGATAGCA	AGACTCTGAC
26821	AAAGCCCAAG	AAATCCACAG	CGGCGGCAGC	AGCAGGAGGA	GGAGCACTGC	GTCTGGCGCC
26881	CAACGAACCC	GTATCGACCC	GCGAGCTTAG	AAACAGGATT	TTTCCCACTC	TGTATGCTAT
26941	ATTTCAACAG	AGCAGGGGCC	AAGAACAAGA	GCTGAAAATA	AAAAACAGGT	CTCTGCGCTC
						CGCTGGAAGA
27061	CGCGGAGGCT	CTCTTCAGCA	AATACTGCGC	GCTGACTCTT	AAGGACTAGT	TTCGCGCCCT
27121	TTCTCAAATT	TAAGCGCGAA	AACTACGTCA	TCTCCAGCGG	CCACACCCGG	CGCCAGCACC
27181	TGTCGTCAGC	GCCATTATGA	GCAAGGAAAT	TCCCACGCCC	TACATGTGGA	GTTACCAGCC
27241	ACAAATGGGA	CTTGCGGCTG	GAGCTGCCCA	AGACTACTCA	ACCCGAATAA	ACTACATGAG
27301	CGCGGGACCC	CACATGATAT	CCCGGGTCAA	CGGAATCCGC	GCCCACCGAA	ACCGAATTCT
27361	CCTCGAACAG	GCGGCTATTA	CCACCACACC	TCGTAATAAC	CTTAATCCCC	GTAGTTGGCC
27421	CGCTGCCCTG	GTGTACCAGG	AAAGTCCCGC	TCCCACCACT	GTGGTACTTC	CCAGAGACGC
27481	CCAGGCCGAA	GTTCAGATGA	CTAACTCAGG	GGCGCAGCTI	GCGGGCGGCT	TTCGTCACAG
27541	GGTGCGGTCG	CCCGGGCAGG	GTATAACTCA	CCTGAAAATC	AGAGGGCGAG	GTATTCAGCT
27601	CAACGACGAG	TCGGTGAGCT	CCTCTCTTGG	TCTCCGTCCG	GACGGGACAT	TTCAGATCGG
27661	CGGCGCTGGC	CGCTCTTCAT	TTACGCCCCG	TCAGGCGATC	CTAACTCTGC	AGACCTCGTC

FIG. 7K

					ATTGAGGAGT	
27781	GGTTTACTTC	AACCCCTTTT	CTGGACCTCC	CGGCCACTAC	CCGGACCAGT	TTATTCCCAA
					ATGACCAGTG	
					AAGTGCTTTG	
27961	CGGTGAGTTT	TGTTACTTTG	AATTGCCCGA	AGAGCATATC	GAGGGCCCGG	CGCACGGCGT
28021	CCGGCTCACC	ACCCAGGTAG	AGCTTACACG	TAGCCTGATT	CGGGAGTTTA	CCAAGCGCCC
28081	CCTGCTAGTG	GAGCGGGAGC	GGGGTCCCTG	TGTTCTGACC	GTGGTTTGCA	ACTGTCCTAA
28141	CCCTGGATTA	CATCAAGATC	TTTGTTGTCA	TCTCTGTGCT	GAGTATAATA	AATACAGAAA
28201	TTAGAATCTA	CTGGGGCTCC	TGTCGCCATC	CTGTGAACGC	CACCGTTTTT	ACCCACCCAA
28261	AGCAGACCAA	AGCAAACCTC	ACCTCCGGTT	TGCACAAGCG	GGCCAATAAG	TACCTTACCT
28321	GGTACTTTAA	CGGCTCTTCA	TTTGTAATTT	ACAACAGTTT	CCAGCGAGAC	GAAGTAAGTT
28381	TGCCACACAA	CCTTCTCGGC	TTCAACTACA	CCGTCAAGAA	AAACACCACC	ACCACCCTCC
28441	TCACCTGCCG	GGAACGTACG	AGTGCGTCAC	CGGTTGCTGC	GCCCACACCT	ACAGCCTGAG
28501	CGTAACCAGA	CATTACTCCC	ATTTTCCCAA	AACAGGAGGT	GAGCTCAACT	CCCGGAACTC
28561	AGGTCAAAAA	AGCATTTTGC	GGGGTGCTGG	GATTTTTTAA	TTAAGTATAT	GAGCAATTCA
28621	AGTAACTCTA	${\tt CAAGCTTGTC}$	TAATTTTTCT	${\tt GGAATTGGGG}$	TCGGGGTTAT	CCTTACTCTT
28681	GTAATTCTGT	${\tt TTATTCTTAT}$	ACTAGCACTT	CTGTGCCTTA	GGGTTGCCGC	CTGCTGCACG
28741	CACGTTTGTA	${\tt CCTATTGTCA}$	${\tt GCTTTTTAAA}$	CGCTGGGGGC	GACATCCAAG	ATGAGGTACA
28801	${\tt TGATTTTAGG}$	CTTGCTCGCC	${\tt CTTGCGGCAG}$	${\tt TCTGCAGCGC}$	TGCCAAAAAG	GTTGAGTTTA
28861	AGGAACCAGC	${\tt TTGCAATGTT}$	ACATTTAAAT	CAGAAGCTAA	TGAATGCACT	ACTCTTATAA
28921	AATGCACCAC	AGAACATGAA	AAGCTTATTA	TTCGCCACAA	AGACAAAATT	GGCAAGTATG
28981	CTGTATATGC	TATTTGGCAG	CCAGGTGACA	CTAACGACTA	TAATGTCACA	GTCTTCCAAG
29041	GTGAAAATCG	${\tt TAAAACTTTT}$	ATGTATAAAT	TTCCATTTTA	TGAAATGTGC	GATATTACCA
29101	TGTACATGAG	CAAACAGTAC	AAGTTGTGGC	CCCCACAAAA	GTGTTTAGAG	AACACTGGCA
29161	CCTTTTGTTC	CACCGCTCTG	CTTATTACAG	CGCTTGCTTT	GGTATGTACC	TTACTTTATC
29221	TCAAATACAA	AAGCAGACGC	AGTTTTATTG	ATGAAAAGAA	AATGCCTTGA	TTTTCCGCTT
29281	GCTTGTATTC	CCCTGGACAA	TTTACTCTAT	GTGGGATATG	CGCCAGGCGG	GAAAGATTAT
29341	ACCCACAACC	TTCAAATCAA	ACTTTCCTGG	ACGTTAGCGC	CTGACTTCTG	CCAGCGCCTG
29401	CACTGCAAAT	TTGATCAAAC	CCAGCTTCAG	CTTGCCTGCT	CCAGAGATGA	CCGGCTCAAC
29461	CATCGCGCCC	ACAACGGACT	ATCGCAACAC	CACTGCTACC	GGACTAAAAT	CTGCCCTAAA
29521	TTTACCCCAA	GTTCATGCCT	TTGTCAATGA	CTGGGCGAGC	TTGGGCATGT	GGTGGTTTTC
29581	CATAGCGCTT	ATGTTTGTTT	GCCTTATTAT	TATGTGGCTT	ATTTGTTGCC	TAAAGCGCAG
29641	ACGCGCCAGA	CCCCCATCT	ATAGGCCTAT	CATTGTGCTC	AACCCACACA	ATGAAAAAAT
29701	TCATAGATTG	GACGGTCTCA	AACCATGTTC	TCTTCTTTTA	CAGTATGATT	AAATGAGACA
29761	TGATTCCTCG	AGTCCTTATA	TTATTGACCC	TTGTTGCGCT	TTTCTGTGCG	TGCTCTACAT
29821	TGGCTGCGGT	CGCTCACATC	GAAGTAGATT	GCATCCCACC	TTTCACAGTT	TACCTGCTTT
29881	ACGGATTTGT	CACCCTTATC	CTCATCTGCA	GCCTCGTCAC	TGTAGTCATC	GCCTTCATTC
29941	AGTTCATTGA	CTGGATTTGT	GTGCGCATTG	CGTACCTTAG	GCACCATCCG	CAATACAGAG
30001	ACAGGACTAT	AGCTGATCTT	CTCAGAATTC	TTTAATTATG	AAACGGATTG	TCACTTTTGT
30061	TTTGCTGATT	TTCTGCGCCC	TACCTGTGCT	TTGCTCCCAA	ACCTCAGCGC	CTCCCAAAAG
30121	ACATATTTCC	TGCAGATTCA	CTCAAATATG	GAACATTCCC	AGCTGCTACA	ACAAACAGAG
30181	CGATTTGTCA	GAAGCCTGGT	TATACGCCAT	CATCTCTGTC	ATGGTTTTTT	GCAGTACCAT

FIG. 7L

30241	TTTTGCCCTA	GCCATATACC	CATACCTTGA	CATTGGTTGG	AATGCCATAG	ATGCCATGAA
30301	CCACCCTACT	TTCCCAGCGC	CCAATGTCAT	ACCACTGCAA	CAGGTTATTG	CCCCAATCAA
30361	TCAGCCTCGC	CCCCCTTCTC	CCACCCCCAC	TGAGATTAGC	TACTTTAATT	TGACAGGTGG
30421	AGATGACTGA	ATCTCTAGAT	CTAGAATTGG	ATGGAATTAA	CACCGAACAG	CGCCTACTAG
30481	AAAGGCGCAA	GGCGGCGTCC	GAGCGAGAAC	GCCTAAAACA	AGAAGTTGAA	GACATGGTTA
30541	ACCTGCACCA	GTGTAAAAGA	GGTATCTTTT	GTGTGGTCAA	GCAGGCCAAA	CTTACCTACG
30601	AAAAAACCAC	TACCGGCAAC	CGCCTTAGCT	ACAAGCTACC	CACCCAGCGC	CAAAAACTGG
30661	TGCTTATGGT	GGGAGAAAA	CCTATCACCG	TCACCCAGCA	CTCGGCAGAA	ACAGAAGGCT
30721	GCCTGCACTT	CCCCTATCAG	GGTCCAGAGG	ACCTCTGCAC	TCTTATTAAA	ACCATGTGTG
		TCTTATTCCA				
30841	ATCAGTCAGC	AAATCTTTGT	CCAGCTTATT	CAGCATCACC	TCCTTTCCCT	CCTCCCAACT
30901	CTGGTATTTC	AGCAGCCTTT	TAGCTGCGAA	CTTTCTCCAA	AGTCTAAATG	GGATGTCAAA
30961	TTCCTCATGT	TCTTGTCCCT	CCGCACCCAC	TATCTTCATA	TTGTTGCAGA	TGAAACGCGC
		GAAGACACCT				
31081	AACTGTGCCT	TTCCTTACCC	CTCCCTTTGT	GTCGCCAAAT	GGGTTCCAAG	AAAGTCCCCC
31141	CGGAGTGCTT	TCTTTGCGTC	TTTCAGAACC	TTTGGTTACC	TCACACGGCA	TGCTTGCGCT
31201	AAAAATGGGC	AGCGGCCTGT	CCCTGGATCA	GGCAGGCAAC	CTTACATCAA	ATACAATCAC
31261	TGTTTCTCAA	CCGCTAAAAA	AAACAAAGTC	CAATATAACT	TTGGAAACAT	CCGCGCCCCT
31321	TACAGTCAGC	TCAGGCGCCC	TAACCATGGC	CACAACTTCG	CCTTTGGTGG	TCTCTGACAA
31381	CACTCTTACC	ATGCAATCAC	AAGCACCGCT	AACCGTGCAA	GACTCAAAAC	TTAGCATTGC
31441	TACCAAAGAG	CCACTTACAG	TGTTAGATGG	AAAACTGGCC	CTGCAGACAT	CAGCCCCCT
		GATAACAACG				
31561	TGGTAGTCTG	GCTGTTACCA	TGGAAAACCC	ACTTTACAAC	AACAATGGAA	AACTTGGGCT
31621	CAAAATTGGC	GGTCCTTTGC	AAGTGGCCAC	CGACTCACAT	GCACTAACAC	TAGGTACTGG
31681	TCAGGGGGTT	GCAGTTCATA	ACAATTTGCT	ACATACAAAA	GTTACAGGCG	CAATAGGGTT
31741	TGATACATCT	GGCAACATGG	AACTTAAAAC	TGGAGATGGC	CTCTATGTGG	ATAGCGCCGG
31801	TCCTAACCAA	AAACTACATA	TTAATCTAAA	TACCACAAAA	GGCCTTGCTT	TTGACAACAC
31861	CGCAATAACA	ATTAACGCTG	GAAAAGGGTT	GGAATTTGAA	ACAGACTCCT	CAAACGGAAA
31921	TCCCATAAAA	ACAAAAATTG	GATCAGGCAT	ACAATATAAT	ACCAATGGAG	CTATGGTTGC
31981	AAAACTTGGA	ACAGGCCTCA	GTTTTGACAG	CTCCGGAGCC	ATAACAATGG	GCAGCATAAA
		CTTACTCTTT				
32101	AGATAAAGAC	TGCAAGCTAA	CTCTGGCGCT	AACAAAATGT	GGCAGTCAAA	TTTTGGGCAC
32161	TGTTTCAGCT	TTGGCAGTAT	CAGGTAATAT	GGCCTCCATC	AATGGAACTC	TAAGCAGTGT
32221	AAACTTGGTT	CTTAGATTTG	ATGACAACGG	AGTGCTTATG	TCAAATTCAT	CACTGGACAA
32281	ACAGTATTGG	AACTTTAGAA	ACGGGGACTC	CACTAACGGT	CAACCATACA	CTTATGCTGT
32341	TGGGTTTATG	CCAAACCTAA	AAGCTTACCC	AAAAACTCAA	AGTAAAACTG	CAAAAAGTAA
32401	TATTGTTAGC	CAGGTGTATC	TTAATGGTGA	CAAGTCTAAA	CCATTGCATT	TTACTATTAC
32461	GCTAAATGGA	ACAGATGAAA	CCAACCAAGT	AAGCAAATAC	TCAATATCAT	TCAGTTGGTC
32521	CTGGAACAGT	GGACAATACA	CTAATGACAA	ATTTGCCACC	AATTCCTATA	CCTTCTCCTA
32581	CATTGCCCAG	GAATAAAGAA	TCGTGAACCT	GTTGCATGTT	ATGTTTCAAC	GTGTTTATTT
32641	TTCAATTGCA	GAAAATTTCA	AGTCATTTT	CATTCAGTAG	TATAGCCCCA	CCACCACATA
32701	GCTTATACTA	ATCACCGTAC	CTTAATCAAA	CTCACAGAAC	CCTAGTATTC	AACCTGCCAC

FIG. 7M

			m> 0> 0> 0	mmmcmccccc	CCMCCCCMMX	አልሮአሮሮአሞሮአ
	CTCCCTCCCA					
	TATCATGGGT					
	AACGCTCATC					
	CCAGCTGCTG					
	AAGTCCACGC					
	GCAGCAGCGC					
	CAGTGGTCTC					
	CACAGCAGCG					
	TATTGTTTAA					
	AACCCACGTG					
33361	CGCTGGACAT	AAACATTACC	TCTTTTGGCA	TGTTGTAATT	CACCACCTCC	CGGTACCATA
	TAAACCTCTG					
33481	GCCCGCCGGC	TATGCACTGC	AGGGAACCGG	GACTGGAACA	ATGACAGTGG	AGAGCCCAGG
33541	ACTCGTAACC	ATGGATCATC	ATGCTCGTCA	TGATATCAAT	GTTGGCACAA	CACAGGCACA
33601	CGTGCATACA	CTTCCTCAGG	ATTACAAGCT	CCTCCCGCGT	CAGAACCATA	TCCCAGGGAA
33661	CAACCCATTC	CTGAATCAGC	GTAAATCCCA	CACTGCAGGG	AAGACCTCGC	ACGTAACTCA
33721	CGTTGTGCAT	${\tt TGTCAAAGTG}$	TTACATTCGG	GCAGCAGCGG	ATGATCCTCC	AGTATGGTAG
33781	CGCGTGTCTC	TGTCTCAAAA	GGAGGTAGGC	GATCCCTACT	GTACGGAGTG	CGCCGAGACA
33841	ACCGAGATCG	TGTTGGŤCGT	AGTGTCATGC	CAAATGGAAC	GCCGGACGTA	GTCATATTTC
33901	CTGAAGCAAA	ACCAGGTGCG	GGCGTGACAA	ACAGATCTGC	GTCTCCGGTC	TCGTCGCTTA
33961	GCTCGCTCTG	TGTAGTAGTT	GTAGTATATC	CACTCTCTCA	AAGCATCCAG	GCGCCCCTG
34021	GCTTCGGGTT	CTATGTAAAC	TCCTTCATGC	GCCGCTGCCC	TGATAACATC	CACCACCGCA
34081	GAATAAGCCA	CACCCAGCCA	ACCTACACAT	TCGTTCTGCG	AGTCACACAC	GGGAGGAGCG
34141	GGAAGAGCTG	GAAGAACCAT	GTTTTTTTT	TTTATTCCAA	AAGATTATCC	AAAACCTCAA
34201	AATGAAGATC	TATTAAGTGA	ACGCGCTCCC	CTCCGGTGGC	GTGGTCAAAC	TCTACAGCCA
34261	AAGAACAGAT	AATGGCATTT	GTAAGATGTT	GCACAATGGC	TTCCAAAAGG	CAAACTGCCC
34321	TCACGTCCAA	GTGGACGTAA	AGGCTAAACC	CTTCAGGGTG	AATCTCCTCT	ATAAACATTC
34381	CAGCACCTTC	AACCATGCCC	TTTTAATAAA	CATCTCGCCA	CCTTATCAAT	ATGTCTCTAA
34441	GCAAATCCCG	AATATTAAGT	CCGGCCATTG	TAAAAATCTG	CTCCAGAGCG	CCCTCCACCT
34501	TCAGCCTCAA	GCAGCGAATC	ATGATTGCAA	AAATTCAGGT	TCCTCACAGA	CCTGTATAAG
34561	ATTCAAAAGC	GGAACATTAA	CAAAAATACC	GCGATCCCGT	AGGTCCCTTC	GCAGGGCCAG
34621	CTGAACATAA	TCGTGCAGGT	CTGCACGGAC	CAGCGCGGCC	ACTTCCCCGC	CAGGAACCAT
34681	GACAAAAGAA	CCCACACTGA	TTATGACACG	CATACTCGGA	GCTATGCTAA	CCAGCGTAGC
34741	CCCGATGTAA	GCTTGTTGCA	TGGGCGGCGA	TATAAAATGC	AAGGTACTGC	TCAAAAAATC
34801	AGGCAAAGCC	TCGCGCAAAA	AAGCAAGCAC	ATCGTAGTCA	TGCTCATGCA	GATAAAGGCA
34861	GGTAAGTTCC	GGAACCACCA	CAGAAAAAGA	CACCATTTTT	CTCTCAAACA	TGTCTGCGGG
	TTCCTGCATA					
	TGTNTTACAA					
	TGACCGTAAA					
	TCCGGAGTCA					
	AAAAAGCGAC					
	GCCCCCATAG					

FIG. 7N

CCCTCCTGCC	TAGGCAAAAT	AGCACCCTCC	CGCTCCAGAA	CAACATACAG	CGCTTCCACA
GCGGCAGCCA	TAACAGTCAG	CCTTACCAGT	AAAAAAACCT	ATTAAAAAAC	ACCACTCGAC
GGACTAAAAA	ATGACGTAAC	GGTTAAAGTC	CACAAAAACC	ACCCAGAAAA	CCGCACGCGA
ACCTACGCCC	AGAAACGAAA	GCCAAAAAAC	CCACAACTTC	CTCAAATCTT	CACTTCCGTT
TTCCCACGAT	ACGTCACTTC	CCATTTTAAA	AAAAAACTAC	AATTCCCAAT	ACATGCAAGT
amacaaaa	TO A A A C C T A C	CTC A CCC GCC	CCGTTCCCAC	GCCCCGCGCC	ACGTCACAAA
CTCCACCCCC	TCATTATCAT	ATTGGCTTCA	ATCCAAAATA	AGGTATATTA	TTGATGATG
	GCGGCAGCCA ACGGCACCAG GGACTAAAAA ACCTACGCCC TTCCCACGAT TACTCCGCCC	GCGGCAGCCA TAACAGTCAG ACGGCACCAG CTCAATCAGT GGACTAAAAA ATGACGTAAC ACCTACGCCC AGAAACGAAA TTCCCACGAT ACGTCACTTC TACTCCGCCC TAAAACCTAC	GCGGCAGCCA TAACAGTCAG CCTTACCAGT ACGGCACCAG CTCAATCAGT CACAGTGTAA GGACTAAAAA ATGACGTAAC GGTTAAAGTC ACCTACGCCC AGAAACGAAA GCCAAAAAAC TTCCCACGAT ACGTCACTTC CCATTTTAAA TACTCCGCCC TAAAACCTAC GTCACCCGCC	GCGGCAGCCA TAACAGTCAG CCTTACCAGT AAAAAAACCT ACGGCACCAG CTCAATCAGT CACAGTGTAA AAAGGGCCAA GGACTAAAAA ATGACGTAAC GGTTAAAGTC CACAAAAACC ACCTACGCCC AGAAACGAAA GCCAAAAAAAC CCACAACTTC TTCCCACGAT ACGTCACTTC CCATTTTAAA AAAAAACTAC TACTCCGCCC TAAAACCTAC GTCACCGCC CCGTTCCCAC	CCCTCCTGCC TAGGCAAAAT AGCACCTCC CGCTCCAGAA CAACATACAG GCGGCAGCCA TAACAGTCAG CCTTACCAGT AAAAAAACCT ATTAAAAAAC ACGGCACCAG CTCAATCAGT CACAGTGTAA AAAGGGCCCAA GTACAGAGCG GGACTAAAAA ATGACGTAAC GGTTAAAGTC CACAAAAACC ACCCAGAAAA ACCTACGCCC AGAAACGAAA GCCAAAAAAAC CCACAACTTC CTCAAATCTT TTCCCACGAT ACGTCACTTC CCATTTTAAA AAAAAACTAC AATTCCCAAT TACTCCGCCC TAAAACCTAC GTCACCCGCC CCGTTCCCAC GCCCCGCGCC CTCCACCCCC TCATTATCAT ATTGGCTTCA ATCCAAAATA AGGTATATTA

1	CATCATCAAT	AATATACCTT	ATTTTGGATT	GAAGCCAATA	TGATAATGAG	GGGGTGGAGT
61	TTGTGACGTG	GCGCGGGGCG	TGGGAACGGG	GCGGGTGACG	TAGTAGTGTG	GCGGAAGTGT
121	GATGTTGCAA	GTGTGGCGGA	ACACATGTAA	GCGACGGATG	TGGCAAAAGT	GACGTTTTTG
				TTTTCGCGCG		
				CCATTTTCGC		
				TAGCGCGTAA		
				AGGTGTTTTT		
				TCAGCTGACG		
481				CAGCGAGTAG		
541				TATCTGCCAC		
				CGAAGAGGTA		
				CGAACTGTAT		
721	CGAAGATCCC	AACGAGGAGG	CGGTTTCGCA	GATTTTTCCC	GACTCTGTAA	TGTTGGCGGT
				GCCGGCGCCC		
841	CCTTTCCCGG	CAGCCCGAGC	AGCCGGAGCA	GAGAGCCTTG	GGTCCGGTTT	CTATGCCAAA
901	CCTTGTACCG	GAGGTGATCG	ATCTTACCTG	CCACGAGGCT	GGCTTTCCAC	CCAGTGACGA
961	CGAGGATGAA	GAGGGTGAGG	AGTTTGTGTT	AGATTATGTG	GAGCACCCCG	GGCACGGTTG
1021	CAGGTCTTGT	CATTATCACC	GGAGGAATAC	GGGGGACCCA	GATATTATGT	GTTCGCTTTG
				CAGTAAGTGA		
1141	TAGAGTGGTG	GGTTTGGTGT	GGTAATTTTT	TTTTTAATTT	TTACAGTTTT	GTGGTTTAAA
				CCTGTGTCTG		
1261	CCAGAACCGG	AGCCTGCAAG	ACCTACCCGC	CGTCCTAAAA	TGGCGCCTGC	TATCCTGAGA
1321	CGCCCGACAT	CACCTGTGTC	TAGAGAATGC	AATAGTAGTA	CGGATAGCTG	TGACTCCGGT
				GTGGTCCCGC		
				GTGGAATGTA		
1501	CCTGGGCAAC	CTTTGGACTT	GAGCTGTAAA	CGCCCCAGGC	CATAAGGTGT	AAACCTGTGA
1561	TTGCGTGTGT	GGTTAACGCC	TTTGTTTGCT	GAATGAGTTG	ATGTAAGTTT	AATAAAGGGT
				AATGGGGCGG		
1681	CGCCGTGGGC	TAATCTTGGT	TACATCTGAC	CTCATGGAGG	CTTGGGAGTG	TTTGGAAGAT
1741	TTTTCTGCTG	TGCGTAACTT	GCTGGAACAG	AGCTCTAACA	GTACCTCTTG	GTTTTGGAGG
1801	TTTCTGTGGG	GCTCATCCCA	GGCAAAGTTA	GTCTGCAGAA	TTAAGGAGGA	TTACAAGTGG
1861	GAATTTGAAG	AGCTTTTGAA	ATCCTGTGGT	GAGCTGTTTG	ATTCTTTGAA	TCTGGGTCAC
1921	CAGGCGCTTT	TCCAAGAGAA	GGTCATCAAG	ACTTTGGATT	TTTCCACACC	GGGGCGCGCT
1981	GCGGCTGCTG	TTGCTTTTTT	GAGTTTTATA	AAGGATAAAT	GGAGCGAAGA	AACCCATCTG
2041	AGCGGGGGGT	ACCTGCTGGA	TTTTCTGGCC	ATGCATCTGT	GGAGAGCGGT	TGTGAGACAC
2101	AAGAATCGCC	TGCTACTGTT	GTCTTCCGTC	CGCCCGGCGA	TAATACCGAC	GGAGGAGCAG
2161	CAGCAGCAGC	AGGAGGAAGC	CAGGCGGCGG	CGGCAGGAGC	AGAGCCCATG	GAACCCGAGA
				TACAGGTGGC		
2281	GACGCATTTT	GACAATTACA	GAGGATGGGC	AGGGGCTAAA	GGGGGTAAAG	AGGGAGCGGG
2341	GGGCTTGTGA	GGCTACAGAG	GAGGCTAGGA	ATCTAGCTTT	TAGCTTAATG	ACCAGACACC
2401	GTCCTGAGTG	TATTACTTTT	CAACAGATCA	AGGATAATTG	CGCTAATGAG	CTTGATCTGC
2461	TGGCGCAGAA	GTATTCCATA	GAGCAGCTGA	CCACTTACTG	GCTGCAGCCA	GGGGATGATT
				TGGCACTTAG		
2581	TCAGCAAACT	TGTAAATATC	AGGAATTGTT	GCTACATTTC	TGGGAACGGG	GCCGAGGTGG
2641	AGATAGATAC	GGAGGATAGG	GTGGCCTTTA	GATGTAGCAT	GATAAATATG	TGGCCGGGGG
2701	TGCTTGGCAT	GGACGGGGTG	GTTATTATGA	ATGTAAGGTT	TACTGGCCCC	AATTTTAGCG
2761	GTACGGTTTT	CCTGGCCAAT	ACCAACCTTA	TCCTACACGG	TGTAAGCTTC	TATGGGTTTA
2821	ACAATACCTG	TGTGGAAGCC	TGGACCGATG	TAAGGGTTCG	GGGCTGTGCC	TTTTACTGCT
2881	GCTGGAAGGG	GGTGGTGTGT	CGCCCCAAAA	GCAGGGCTTC	AATTAAGAAA	TGCCTCTTTG
2941	AAAGGTGTAC	CTTGGGTATC	CTGTCTGAGG	GTAACTCCAG	GGTGCGCCAC	AATGTGGCCT
3001	CCGACTGTGG	TTGCTTCATG	CTAGTGAAAA	GCGTGGCTGT	GATTAAGCAT	AACATGGTAT
3061	GTGGCAACTG	CGAGGACAGG	GCCTCTCAGA	TGCTGACCTG	CTCGGACGGC	AACTGTCACC
3121	TGCTGAAGAC	CATTCACGTA	GCCAGCCACT	CTCGCAAGGC	CTGGCCAGTG	TTTGAGCATA
3181	ACATACTGAC	CCGCTGTTCC	TTGCATTTGG	GTAACAGGAG	GGGGGTGTTC	CTACCTTACC
3241	AATGCAATTT	GAGTCACACT	AAGATATTGC	TTGAGCCCGA	GAGCATGTCC	AAGGTGAACC

2201	man naccocc	ርመምምር እር አጥር	Δ CCΔTGΔΔGΔ	TCTGGAAGGT	GCTGAGGTAC	GATGAGACCC
3301	TGAACGGGGI	CACACCCTCC	CACTGTGGCG	GTAAACATAT	TAGGAACCAG	CCTGTGATGC
3361	GCACCAGGIG	CAGACCAGCTG	AGGCCCGATC	ACTTGGTGCT	GGCCTGCACC	CGCGCTGAGT
3421	TGGATGTGAC	CCATCAACAT	ACACATTGAG	GTACTGAAAT	GTGTGGGCGT	GGCTTAAGGG
3481	TIGGCTCTAG	CGAIGAAGAI	CCCCCTCTTA	TGTAGTTTTG	TATCTGTTTT	GCAGCAGCCG
3541	TGGGAAAGAA	TATATAAGGI	TO COUNTY CATC	GAAGCATTGT	GAGCTCATAT	TTGACAACGC
3601	CCGCCGCCAT	MAGCACCAAC	COCCCOCACA	ATGTGATGGG	CTCCAGCATT	GATGGTCGCC
3661	GCATGCCCCC	ATGGGCCGGG	A CON COUNCY	CCTACGAGAC	CGTGTCTGGA	ACGCCGTTGG
3721	CCGTCCTGCC	CGCAAACTCT	ACTACCTIGA CCCCC	CTGCAGCCAC	CGCCCGCGG	ATTGTGACTG
3781	AGACTGCAGC	CTCCGCCGCC	GCTTCAGCCG	GTGCAGCTTC	CCCTTCATCC	GCCCGCGATG
3841	ACTTTGCTTT	CCTGAGCCCG	CTTGCAAGCA	ATTCTTTGAC	CCCCCAACTT	AATGTCGTTT
3901	ACAAGTTGAC	GGCTCTTTTG	GCACAATIGG	MUNICIPLICATE	CAACCCTTCC	TCCCCTCCCA
3961	CTCAGCAGCT	GTTGGATCTG	CGCCAGCAGG	TTTCTGCCCT	CATTTCCATC	AAGCAAGTGT
4021	ATGCGGTTTA	AAACATAAAT	AAAAAACCAG	ACTCTGTTTG	CCCCCACCAC	CGGTCTCGGT
4081	CTTGCTGTCT	TTATTTAGGG	GTTTTGCGCG	CGCGGTAGGC	CTCACTCTGG	ATGTTCAGAT
4141	CGTTGAGGGT	CCTGTGTATT	COCCOCCOCCA	CGTGGTAAAG	CTCCAGAGCT	TCATGCTGCG
4201	ACATGGGCAT	AAGCCCGTCT	CTGGGGTGGA	GGTAGCACCA	CCCCTCCTCC	CTABABATGT
4261	GGGTGGTGTT	GTAGATGATC	CAGTCGTAGC	AGGAGCGCTG	CULTUTUTE	ACAAAGCGGT
4321	CTTTCAGTAG	CAAGCTGATT	GCCAGGGGCA	GGCCCTTGGT	CUMCCACACA	ልጥጥጥጥ ልርርጥ
4381	TAAGCTGGGA	TGGGTGCATA	CGTGGGGATA	TGAGATGCAT	CTTGGACIGI	ACCACCACAG
4441	TGGCTATGTT	CCCAGCCATA	TCCCTCCGGG	GATTCATGTT	ACCANAGECC	TOCANGACA
4501	TGTATCCGGT	GCACTTGGGA	AATTTGTCAT	GTAGCTTAGA	AGGAAATGCG	AUCCCAAUCC
4561	TGGAGACGCC	CTTGTGACCT	CCAAGATTTT	CCATGCATTC	GICCATAAIG	MACALLAGO MACALLAGO
4621	GCCCACGGGC	GGCGGCCTGG	GCGAAGATAT	TTCTGGGATC	ACTAACGICA	CCACACTCCC
4681	CCAGGATGAG	ATCGTCATAG	GCCATTTTTA	CAAAGCGCGG	GCGGAGGGTG	AMENCECACE
4741	GTATAATGGT	TCCATCCGGC	CCAGGGGCGT	AGTTACCCTC	ACAGATTIGC	ATTICCCACG
4801	CTTTGAGTTC	AGATGGGGGG	ATCATGTCTA	CCTGCGGGGC	GATGAAGAAA	MCGGIIICCG
4861	GGGTAGGGGA	GATCAGCTGG	GAAGAAAGCA	GGTTCCTGAG	CAGCTGCGAC	TTACCGCAGC
4921	CGGTGGGCCC	GTAAATCACA	CCTATTACCG	GGTGCAACTG	GTAGTTAAGA	GAGCTGCAGC
4001	ጥር ር ር ር ጥር ልጥር	CCTGAGCAGG	GGGGCCACTT	CGTTAAGCAT	GTCCCTGACT	CGCATGTTT
E0/11	CCCTCACCAA	ATCCCCCAGA	AGGCGCTCGC	CGCCCAGCGA	TAGCAGTTCT	TGCAAGGAAG
E101	ር ሃ ሃ ሃርብላቡብላብ	CAACGGTTTG	AGACCGTCCG	CCGTAGGCAT	GCTTTTGAGC	GTTTGACCAA
5161	CCACTTCCAG	CCCCTCCCAC	AGCTCGGTCA	CCTGCTCTAC	GGCATCTCGA	TCCAGCATAT
E221	CTCCTCCTTT	CCCCCCTTCC	GGCGGCTTTC	GCTGTACGGC	AGTAGTCGGT	GCTCGTCCAG
E201	ACCCCCCACC	GTCATGTCTT	TCCACGGGCG	CAGGGTCCTC	GTCAGCGTAG	TCTGGGTCAC
E2/1	CCTCAACCCC	TECECTCEGG	GCTGCGCGCT	GGCCAGGGTG	CGCTTGAGGC	TGGTCCTGCT
5401	CCTCCTCAAC	CGCTGCCGGT	CTTCGCCCTG	CGCGTCGGCC	AGGTAGCATT	TGACCATGGI
5161	ርጥሮ እጥ እርጥሮር	AGCCCCTCCG	CGGCGTGGCC	CTTGGCGCGC	AGCTTGCCCT	TGGAGGAGGC
5521	CCCCCACGAG	GGGCAGTGCA	GACTTTTGAG	GGCGTAGAGC	TTGGGCGCGA	GAAATACCGA
5591	TTCCCCCCAC	TAGGCATCCG	CGCCGCAGGC	CCCGCAGACG	GTCTCGCATT	CCACGAGCCA
E C 11	CCTCACCTCT	CCCCCTTCGG	GGTCAAAAAC	CAGGTTTCCC	CCATGCTTT	TGATGCGTTT
5701	CMMPCCMCTC	GTTTCCATGA	GCCGGTGTCC	: ACGCTCGGTG	ACGAAAAGGC	TGTCCGTGTC
5761	CCCCTATACA	GACTTGAGAG	GCCTGTCCTC	GAGCGGTGTT	CCGCGGTCCT	CCTCGTATAG
E031	አ አ አ ርጥር ርር ልር	САСТСТСАСА	CAAAGGCTCG	CGTCCAGGCC	AGCACGAAGG	AGGCTAAGTG
5001	CCACCCCTAC	CGGTCGTTGT	CCACTAGGGG	; GTCCACTCGC	TCCAGGGTGT	GAAGACACAT
50/1	CTCCCCCTCT	TCGGCATCAA	GGAAGGTGAT	TGGTTTGTAG	GTGTAGGCCA	CGTGACCGGG
6001	ጥርጥጥርርጥር እ እ	GGGGGGCTAT	AAAAGGGGGT	GGGGGCGCGT	TCGTCCTCAC	TUTUTTUCGU
6001	3 TO 1 TO C	CCCAGGGCCA	CCTGTTGGGG	TGAGTACTCC	CTCTGAAAAG	CGGGCATGAC
6131	TTCTCCCCTT	ACATTGTCAG	TTTCCAAAAA	CGAGGAGGAT	TTGATATTCA	CCTGGCCCGC
0121	COMCAMOCOCIA	TACACCCTCC	CCCCATCCAT	CTGGTCAGAA	AAGACAATCT	TTTTGTTGTC
C241	አአሮርመውርርጥር	CCAAACGACC	CGTAGAGGGC	: GTTGGACAGC	AACTTGGCGA	TGGAGCGCAG
6241	COMMOCOMM	, GCZZZCGZCC	CGGCGCGCTC	CTTGGCCGCG	ATGTTTAGCT	GCACGTATTC
6301	. GG111GG171	LACCCCAT	CGGGAAAGAG	GGTGGTGCGC	TCGTCGGGCA	CCAGGTGCAC
0301	COCCAACO	, CUCCACCUTT	CCCTCACAA	GTCAACGCTG	GTGGCTACCT	CTCCGCGTAG
6421	. GUGUCAACCG	COCTICIONS	GGGIGYCYYG	. CTTGCGC16	CAGAATGGCG	GTAGGGGGTC
6481	GUGUTUGTTO	, GICCAGCAGA	, CGMCWCCCGC(CACCCTABAC	ACCCCGGGCA	GCAGGCGCGC
6541	TAGCTGCGTC	TUGTUUGGG	, 661616616	, caccoinanc		

6601	GTCGAAGTAG	TCTATCTTGC	ATCCTTGCAA	GTCTAGCGCC	TGCTGCCATG	CGCGGGCGGC
6661	AAGCGCGCGC	TCGTATGGGT	TGAGTGGGGG	ACCCCATGGC	ATGGGGTGGG	TGAGCGCGGA
6721	GGCGTACATG	CCGCAAATGT	CGTAAACGTA	GAGGGGCTCT	CTGAGTATTC	CAAGATATGT
6781	AGGGTAGCAT	CTTCCACCGC	GGATGCTGGC	GCGCACGTAA	TCGTATAGTT	CGTGCGAGGG
6841	AGCGAGGAGG	TCGGGACCGA	GGTTGCTACG	GGCGGGCTGC	TCTGCTCGGA	AGACTATCTG
6901	CCTGAAGATG	GCATGTGAGT	TGGATGATAT	GGTTGGACGC	TGGAAGACGT	TGAAGCTGGC
6961	GTCTGTGAGA	CCTACCGCGT	CACGCACGAA	GGAGGCGTAG	GAGTCGCGCA	GCTTGTTGAC
7021	CAGCTCGGCG	GTGACCTGCA	CGTCTAGGGC	GCAGTAGTCC	AGGGTTTCCT	TGATGATGTC
7081	ATACTTATCC	TGTCCCTTTT	TTTTCCACAG	CTCGCGGTTG	AGGACAAACT	CTTCGCGGTC
7141	TTTCCAGTAC	TCTTGGATCG	GAAACCCGTC	GGCCTCCGAA	CGGTAAGAGC	CTAGCATGTA
7201	GAACTGGTTG	ACGGCCTGGT	AGGCGCAGCA	TCCCTTTTCT	ACGGGTAGCG	CGTATGCCTG
7261	CGCGGCCTTC	CGGAGCGAGG	TGTGGGTGAG	CGCAAAGGTG	TCCCTGACCA	TGACTTTGAG
7321	GTACTGGTAT	TTGAAGTCAG	TGTCGTCGCA	TCCGCCCTGC	TCCCAGAGCA	AAAAGTCCGT
7381	GCGCTTTTTG	GAACGCGGAT	TTGGCAGGGC	GAAGGTGACA	TCGTTGAAGA	GTATCTTTCC
7441	CGCGCGAGGC	ATAAAGTTGC	GTGTGATGCG	GAAGGGTCCC	GGCACCTCGG	AACGGTTGTT
7501	AATTACCTGG	GCGGCGAGCA	CGATCTCGTC	AAAGCCGTTG	ATGTTGTGGC	CCACAATGTA
7561	AAGTTCCAAG	AAGCGCGGGA	TGCCCTTGAT	GGAAGGCAAT	TTTTTAAGTT	CCTCGTAGGT
	GAGCTCTTCA					
7681	GGAAGCGACG	AATGAGCTCC	ACAGGTCACG	GGCCATTAGC	ATTTGCAGGT	GGTCGCGAAA
7741	GGTCCTAAAC	TGGCGACCTA	TGGCCATTTT	TTCTGGGGTG	ATGCAGTAGA	AGGTAAGCGG
7801	GTCTTGTTCC	CAGCGGTCCC	ATCCAAGGTT	CGCGGCTAGG	TCTCGCGCGG	CAGTCACTAG
7861	AGGCTCATCT	CCGCCGAACT	TCATGACCAG	CATGAAGGGC	ACGAGCTGCT	TCCCAAAGGC
7921	CCCCATCCAA	GTATAGGTCT	CTACATCGTA	GGTGACAAAG	AGACGCTCGG	TGCGAGGATG
7981	CGAGCCGATC	GGGAAGAACT	GGATCTCCCG	CCACCAATTG	GAGGAGTGGC	TATTGATGTG
	GTGAAAGTAG					
	GCAGTACTGG					
	CACAAGGAAG					
	TACTTCGGCT					
8281	CACCACGCCG	CGCGAGCCCA	AAGTCCAGAT	GTCCGCGCGC	GGCGGTCGGA	GCTTGATGAC
	AACATCGCGC					
	GAGCTCCTGC					
	CCTAATTTCC					
8521	CGGCGCGACT	ACGGTACCGC	GCGGCGGCG	GTGGGCCGCG	GGGGTGTCCT	TGGATGATGC
8581	ATCTAAAAGC	GGTGACGCGG	GCGAGCCCCC	GGAGGTAGGG	GGGGCTCCGG	ACCCGCCGGG
8641	AGAGGGGGCA	GGGGCACGTC	GCCCCCCCC	GCGGGCAGGA	GCTGGTGCTG	CGCGCGTAGG
	TTGCTGGCGA					
	ACGACGGGCC					
	TTGACGGCGG					
	TCGGCCATGA					
	GTGGCGGCGA					
	TCGTTCCAGA					
9061	TGCGCGAGAT	TGAGCTCCAC	GTGCCGGGCG	AAGACGGCGT	AGTTTCGCAG	GCGCTGAAAG
9121	AGGTAGTTGA	GGGTGGTGGC	GGTGTGTTCT	GCCACGAAGA	AGTACATAAC	CCAGCGTCGC
9181	AACGTGGATT	CGTTGATATC	CCCCAAGGCC	TCAAGGCGCT	CCATGGCCTC	GTAGAAGTCC
	ACGGCGAAGT					
9301	CGGATGAGCT	CGGCGACAGT	GTCGCGCACC	TCGCGCTCAA	AGGCTACAGG	GGCCTCTTCT
9361	TCTTCTTCAA	TCTCCTCTTC	CATAAGGGCC	TCCCCTTCTT	CTTCTTCTGG	CGGCGGTGGG
9421	GGAGGGGGGA	CACGGCGGCG	ACGACGGCGC	ACCGGGAGGC	GGTCGACAAA	GCGCTCGATC
	ATCTCCCCGC					
	AGTTGGAAGA					
9601	AGGGATACGG	CGCTAACGAT	GCATCTCAAC	AATTGTTGTG	TAGGTACTCC	GCCGCCGAGG
9661	GACCTGAGCG	AGTCCGCATC	GACCGGATCG	GAAAACCTCT	CGAGAAAGGC	GTCTAACCAG
9721	TCACAGTCGC	AAGGTAGGCT	GAGCACCGTG	GCGGGCGCA	GCGGGCGCG	GTCGGGGTTG
9781	TTTCTGGCGG	AGGTGCTGCT	GATGATGTAA	TTAAAGTAGG	CGGTCTTGAG	ACGGCGGATG
9841	GTCGACAGAA	GCACCATGTC	CTTGGGTCCG	GCCTGCTGAA	TGCGCAGGCG	GTCGGCCATG

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		CGTTTTGACA	TCCCCCCACC	ጥርጥጥርጥልርጥ	AGTCTTGCAT	GAGCCTTTCT
9901	CCCCAGGCTT	CTTCTTCTCC	TCGGCGCAGG	CCTCCATCTC	TTGCATCTAT	CGCTGCGGCG
9961	ACCGGCACTT	TTGGCCGTAG	CECCCCCCC	CTTCCTCCCA	TCCGTGTGAC	CCCGAAGCCC
10021	GCGGCGGAGT	GAAGCAGGGC	ma composes	ACA ACCCCCCT	CCCCTAATAT	GGCCTGCTGC
10081	CTCATCGGCT	GGGTAGACTG	CARCOCATCC	ACAACGCGC1	AGCGGTGGTA	TGCGCCCGTG
10141	ACCTGCGTGA	AAGTGCAGTT	GAAGTCATCC	CACCACACAA	CCCTCTCCTC	ACCCGGCTGC
10201	TTGATGGTGT	AAGTGCAGTT	ACCCCATAACG	CCCCTCCACT	CANATACGTA	GTCGTTGCAA
10261	GAGAGCTCGG	TGTACCTGAG	ACGCGAGTAA	P V CACC L C CAG 1	CAAATACGTA	GTAGAGGGGC
10321	GTCCGCACCA	GGTACTGGTA	TCCCACCAAA	AAGIGCGGCG	ACATA ACCCG	ATGATATCCG
10381	CAGCGTAGGG	TGGCCGGGGC TGGACATCCA	CCTCATCACCCC	CCCCCCCCCCC	TGGAGGCGCG	CGGAAAGTCG
10441	TAGATGTACC	TCCAGATGTT	CCCCACCCCC	A A A A A CTCCT	CCATGGTCGG	GACGCTCTGG
10501	CGGACGCGGT	GCGCGCAATC	CERCAGUGGU	MARAAGIGCI MACACCCTCC	AAAAGGAGAG	CCTGTAAGCG
10561	CCGGTCAGGC	GCGCGCAATC	CTTGACGCTC	CCCAACCGIGC	TCATGGCGGA	CGACCGGGGT
10621	GGCACTCTTC	CGTGGTCTGG	TGGATAAATT	TGCAAGGG1A	TACCGCCCCC	GTGTCGAACC
10681	TCGAGCCCCG	TATCCGGCCG	TCCGCCGTGA	CONCOMPTE	CCTTCCTTCC	AGGCGCGGCG
10741	CAGGTGTGCG	ACGTCAGACA	ACGGGGGAGT	CCCCCCCACC	CTAACCCCTT	AGGCTGGAAA
10801	GCTGCTGCGC	TAGCTTTTTT	GGCCACTGGC	CGCGCGCAGC	WATER COURT	CCCTTGAGTC
10861	GCGAAAGCAT	TAAGTGGCTC	GCTCCCTGTA	GCCGGAGGGI	CCCAACGGGG	CTTTCCCTCC
10921	GCGGGACCCC	CGGTTCGAGT	CTCGGACCGG	CCGGACTGCG	CCCACCACCC	CCTTTTTTCC
10981	CCGTCATGCA	AGACCCCGCT	TGCAAATTCC	TCCGGAAACA	CECCECACCA	CCCCCAAGAG
11041	TTTTCCCAGA	TGCATCCGGT	GCTGCGGCAG	MACCOMMONTO	CTCCTCAGCA	AGGAGGGGGG
11101	CAAGAGCAGC	GGCAGACATG	CAGGGCACCC	CARTACCARC	CIACCGCGIC	CCGCCCCCG
11161	ACATCCGCGG	TTGACGCGGC	AGCAGATGGT	GATTACGAAC	maccaccecc	СТСТСТСАС
11221	CACTACCTGG	ACTTGGAGGA	GGGCGAGGGC	TGGCGCGCGC	CCMACCMCCC	CCCCCACAAC
11281	CGGTACCCAA	GGGTGCAGCT	GAAGCGTGAT	ACGCGTGAGG	CCCATCGAAA	GTTCCACGCA
11341	CTGTTTCGCG	ACCGCGAGGG	AGAGGAGCCC	GAGGAGATGC	mcccccaaaa	CCACTTTCAC
11401	GGGCGCGAGC	TGCGGCATGG	CCTGAATCGC	GAGCGGTTGC	TGCGCGAGGA	CCACCTCCTA
11461	CCCGACGCGC	GAACCGGGAT	TAGTCCCGCG	CGCGCACACG	166C6GCCGC	TANCARCCAC
11521	ACCGCATACG	AGCAGACGGT	GAACCAGGAG	ATTAACTTTC	MAAAAAGC11	CTCCCACTTT
11581	GTGCGTACGC	TTGTGGCGCG	CGAGGAGGTG	GCTATAGGAC	TGATGCATCT	COUCCULIT
11641	GTAAGCGCGC	TGGAGCAAAA	CCCAAATAGC	AAGCCGCTCA	TGGCGCAGCI	ACTACACCCC
11701	GTGCAGCACA	GCAGGGACAA	CGAGGCATTC	AGGGATGCGC	TGCTAAACAI	CCACCACCCC
11761	GAGGGCCGCT	GGCTGCTCGA	TTTGATAAAC	ATCCTGCAGA	GCATAGTGGI	CCTCCCCAAC
11821	AGCTTGAGCC	TGGCTGACAA	GGTGGCCGCC	ATCAACTATT	CCATGCTTAG	CCTGGGCAAG
11881	TTTTACGCCC	GCAAGATATA	CCATACCCCT	TACGTTCCCA	TAGACAAGGA	CCTCCCCCTT
11941	GAGGGGTTCT	ACATGCGCAT	GGCGCTGAAG	GTGCTTACCT	TGAGCGACGA	CCTGGGGGTT
12001	TATCGCAACG	AGCGCATCCA	CAAGGCCGTG	AGCGTGAGCC	GGCGGCGCGA	CCATACACAC
12061	CGCGAGCTGA	TGCACAGCCT	GCAAAGGGCC	CTGGCTGGCA	CGGGCAGCGG	ACCCCCCCCCC
12121	GCCGAGTCCT	ACTTTGACGC	GGGCGCTGAC	CTGCGCTGGG	CCCCAAGCCG	ACGCGCCCIG
12181	GAGGCAGCTG	GGGCCGGACC	TGGGCTGGCG	GTGGCACCCG	CGCGCGCTGG	CAACGICGGC
12241	GGCGTGGAGG	AATATGACGA	GGACGATGAG	TACGAGCCAG	AGGACGGCGA	GTACTAAGCG
12301	GTGATGTTTC	TGATCAGATG	ATGCAAGACG	CAACGGACCC	GGCGGTGCGG	GCGGCGCTGC
12361	AGAGCCAGCC	GTCCGGCCTT	AACTCCACGG	ACGACTGGCG	CCAGGTCATG	GACCGCATCA
12421	TGTCGCTGAC	TGCGCGCAAT	CCTGACGCGT	TCCGGCAGCA	GCCGCAGGCC	AACCGGCTCT
12/21	CCCC A A ጥጥርጥ	GGAAGCGGTG	GTCCCGGCGC	GCGCAAACCC	CACGCACGAG	AAGGTGCTGG
125/1	CCMTCCTMAAA	_	GAAAACAGGG	CCATCCGGCC	CGACGAGGCC	GGCCIGGICI
12601	ACGACGCGCT	CCTTCAGCGC	GTGGCTCGTT	ACAACAGCGG	CAACGTGCAG	ACCAACCIGG
12661	ACCECCTECT	CCCCCATCTC	CGCGAGGCCG	TGGCGCAGCG	TGAGCGCGCG	CAGCAGCAGG
12721	CC A ACCTGGG	ርጥርር ልጥርርጥጥ	GCACTAAACG	CCTTCCTGAG	TACACAGCCC	GCCAACGIGC
12781	CCCCCCCACA	GGAGGACTAC	ACCAACTTTG	TGAGCGCACT	GCGGCTAATG	GIGACIGAGA
12041	CACCCCAAAC	TCACCTCTAC	CAGTCTGGGC	CAGACTATTT	TTTCCAGACC	AGTAGACAAG
12001	CCCTCCAGAC	CCTAAACCTG	AGCCAGGCTT	' TCAAAAACTT	GCAGGGGCTG	1666666166
12061	CCCCMCCCAC	NOCCCACCCC	-CCGACCGTGT	' CTAGCTTGCT	' GACGCCCAAC	TUGUGUUTGI
12021	T	' እልጥልርርርርርር	TTCACGGACA	GTGGCAGCGT	GTCCCGGGAC	ACATACCIAG
12001	COCACOOCCO	• ሮ እሮ <mark>እርጥርጥ</mark> እር	- CGCGAGGCCA	TAGGTCAGGC	GCATGTGGAC	GAGCATACTT
13141	TCCAGGAGAT	TACAAGTGTC	AGCCGCGCGC	TGGGGCAGGA	GGACACGGGC	AGCCTGGAGG

12201	011000m111	CM & CCMCCMC	3CC33CCCCC	CCCACAACAT	CCCCTCGTTG	СУСУСТТАУ
					CGTGAGCCTT	
					GCGCAACATG	
					CTACTTGCAT	
					GCACTGGCTA	
					TGGATTCCTC	
					AGAGTTGCAA	
					AAGCAGCTTG	
					AAGCTTGATA	
					GGAGTACCTA	
					TCCCAACAAC	
					GGAGCACAGG	
					GCGGGGTCTG	
13981	ACGATGACTC	GGCAGACGAC	AGCAGCGTCC	TGGATTTGGG	AGGGAGTGGC	AACCCGTTTG
14041	CGCACCTTCG	CCCCAGGCTG	GGGAGAATGT	TTTAAAAAAA	AAAAAGCATG	ATGCAAAATA
					TGTATTCCCC	
					GAGAGTGTGG	
					CTGGACCCGC	
					CGTTACTCTG	
					TCAACGGATG	
					ATTCAAAACA	
					CGGTCGCACT	
					GAGTTCATGT	
					GACAATCAGG	
					TCCGAGACCA	
					GGCAGACAGA	
14701	CCTTATGAAC	AMCGCGGGTCG	A COMMOCACIA	CLIGAMAGIG	AGACTGGGGT	TOTAL CONTROL OF THE STATE OF T
14/01	GGAAAGCGAC	ATCGGGGTAA	CCCMAMAMAC	AAACCAACTIC	TTCCATCCAG	A C A T C A T T T T T
					AGCAACTTGT	
					GATGATCTGG	
14941	CAAGCGGCAA	CCCTTCCAGG	MCC3 CCCCM3	CCACCCCACC	TTGAAAGATG	ACACCCAACA
					GGCGCGGAAG	
					GATCATGCCA	
					GAAGCAGCGG	
					AAACCGGTGA	
					AATGACAGCA	
15361	GTACCGCAGC	TGGTACCTTG	CATACAACTA	CGGCGACCCT	CAGACCGGAA	TCCGCTCATG
15421	GACCCTGCTT	TGCACTCCTG	ACGTAACCTG	CGGCTCGGAG	CAGGTCTACT	GGTCGTTGCC
					CAGATCAGCA	
					TACAACGACC	
					TTCAATCGCT	
					GTCAGTGAAA	
					GGAGGAGTCC	
					AAGGCCCTGG	
15841	GCCGCGCGTC	CTATCGAGCC	GCACTTTTTG	AGCAAGCATG	TCCATCCTTA	TATCGCCCAG
15901	CAATAACACA	GGCTGGGGCC	TGCGCTTCCC	AAGCAAGATG	TTTGGCGGGG	CCAAGAAGCG
15961	CTCCGACCAA	CACCCAGTGC	GCGTGCGCGG	GCACTACCGC	GCGCCCTGGG	GCGCGCACAA
					GACGCGGTGG	
					GACGCGGCCA	
					CGGAGGCGCG	
16201	CCACCGCCGC	CGACCCGGCA	CTGCCGCCCA	ACGCGCGGCG	GCGGCCCTGC	TTAACCGCGC
16261	ACGTCGCACC	GGCCGACGGG	CGGCCATGCG	GGCCGCTCGA	AGGCTGGCCG	CGGGTATTGT
16321	CACTGTGCCC	CCCAGGTCCA	GGCGACGAGC	GGCCGCCGCA	GCAGCCGCGG	CCATTAGTGC
					GACTCGGTTA	
16441	CCTCCCCTC	CGCACCCGCC	CCCCGCGCAA	CTAGATTGCA	AGAAAAAACT	ACTTAGACTC
10441	-31000010	COUNCECCOCC				

16501	СФАСФСФФСФ	ATGTATCCAG	CGGCGGCGGC	GCGCAACGAA	GCTATGTCCA	AGCGCAAAAT
10001	CANACAACAC	ATCCTCCACC	TCATCGCGCC	GGAGATCTAT	GGCCCCCCGA	AGAAGGAAGA
16621	CCACCATTAC	AAGCCCCGAA	AGCTAAAGCG	GGTCAAAAAG	AAAAAGAAAG	ATGATGATGA
1 (6 0 1	ጥር እ እርጥጥር እር	CACCAGGTGG	AACTGCTGCA	CGCTACCGCG	CCCAGGCGAC	GGGTACAGTG
1 (7 / 1	CARACCTCCA	CCCCTAAAAC	GTGTTTTGCG	ACCCGGCACC	ACCGTAGTCT	TTACGCCCGG
1 (0 0 1	THE REPORT OF THE PROPERTY OF	ACCCCCACCT	ACAAGCGCGT	GTATGATGAG	GTGTACGGCG	ACGAGGACCI
10001	CCMTCACCAC	GCCAACGAGC	GCCTCGGGGA	GTTTGCCTAC	GGAAAGCGGC	ATAAGGACAT
10001	CCTCCCCTTC	CCGCTGGACG	AGGGCAACCC	AACACCTAGC	CTAAAGCCCG	TAACACTGCA
16921	GCTGGCGTTG	CCCGCGCTTG	CACCGTCCGA	AGAAAAGCGC	GGCCTAAAGC	GCGAGTCTGG
10001	GCAGGIGCIG	CCCACCGTGC	ACCTGATGGT	ACCCAAGCGC	CAGCGACTGG	AAGATGTCTT
17041	TGACTIGGCA	ACCGTGGAAC	CTGGGCTGGA	GCCCGAGGTC	CGCGTGCGGC	CAATCAAGCA
1/101	GGAAAAAATG	GGACTGGGCG	TCCAGACCGT	GGACGTTCAG	ATACCCACTA	CCAGTAGCAC
1/101	GGTGGCGCCG	ACCGCCACAG	ACCCCATCGA	GACACAAACG	TCCCCGGTTG	CCTCAGCGGT
17221	CAGTATTGCC	GCGGTGCAGG	CCCTCCCTCC	GGCCGCGTCC	AAGACCTCTA	CGGAGGTGCA
1/281	GGCGGATGCC	TGGATGTTTC	CCCTTTCACC	CCCCGGCGC	CCGCGCGGTT	CGAGGAAGTA
17341	AACGGACCCG	AGCGCGCTAC	TCCCCGATA	TGCCCTACAT	CCTTCCATTG	CGCCTACCCC
17401	CGGCGCCGCC	GGCTACACCT	ACCCCCCCAG	AAGACGAGCA	ACTACCCGAC	GCCGAACCAC
17461	CGGCTATCGT	CGCCGCCGCC	CTCCCCCTCC	CCAGCCCGTG	CTGGCCCCGA	TTTCCGTGCG
17521	CACTGGAACC	CGCCGAAGGAG	CCACCACCCT	GGTGCTGCCA	ACAGCGCGCT	ACCACCCCAG
17581	CAGGGTGGCT	AAGCCGGTCT	TTCTCCTTCT	TGCAGATATG	GCCCTCACCT	GCCGCCTCCG
17641	CATCGTTTAA	CCGGGATTCC	CACCAACAAT	GCACCGTAGG	AGGGGCATGG	CCGGCCACGG
17701	TTTCCCCGGTG	GGCATGCGTC	CTCCCCACCA	CCGCCGCGG	CGCGCGTCGC	ACCGTCGCAT
17761	CCTGACGGGC	ATCCTGCCCC	TCCTTATTCC	ACTGATCGCC	GCGGCGATTG	GCGCCGTGCC
17821	GCGCGGCGGT	TCCGTGGCCT	TCCTTATTCC	GAGACACTGA	тталаласаа	GTTGCATGTG
17881	CGGAATTGCA	AATAAAAAGT	CTCGACTCTC	ACCCTCCCTT	GGTCCTGTAA	CTATTTTGTA
17941	GAAAAATCAA	CATCAACTTT	CCCTCTCTCC	CCCCGCGACA	CGGCTCGCGC	CCGTTCATGG
18001	GAATGGAAGA	AGATATCGGC	ACCACCA ATA	TCACCCCTCC	CGCCTTCAGC	TGGGGCTCGC
18061	GAAACTGGCA	CATTAAAAAT	MUCHGCAAIA	CCCTTAACAA	CTATGGCAGC	AAGGCCTGGA
18121	TGTGGAGCGG	AGGCCAGATG	CUCACCONTA	ACTTCA AACA	GCAAAATTTC	CAACAAAAGG
18181	ACAGCAGCAC	CCTGGCCTCT	CIGAGGGAIA	CCCTCCTCCA	CCTGGCCAAC	CAGGCAGTGC
18241	TGGTAGATGG	TAACAGTAAG	CUTCATCCC	CCCCTCCCGT	AGAGGAGCCT	CCACCGGCCG
18301	AAAATAAGAT	GTCTCCAGAG	CTTGATCCCC	AAAGCGTCC	GCGCCCCGAC	AGGGAAGAAA
18361	TGGAGACAGT	GCAAATAGAC	CACCCTCCCT	CCTACCACCA	GGCACTAAAG	CAAGGCCTGC
18421	CTCTGGTGAC	TCCCATCGCG	CCCATCCCT	CCCCACTCCT	GGGCCAGCAC	ACACCCGTAA
18481	CCACCACCCG	GCCTCCCCCC	CCCAIGGCIA	ACCACAAACC	TETECTECCA	GGCCCGACCG
18541	CGCTGGACCT	AACCCGTCCT	ACCCCCCCCCC	CCCTCCCCC	CCCCCCAGC	GGTCCGCGAT
18601	CCGTTGTTGT	CGTAGCCAGT	AGCCGCGCG1	AAACCACACT	CAACAGCATC	GTGGGTCTGG
18661	CGTTGCGGCC	CGTAGCCAGT	CCACCAMCTGGC	TOTO ATACC	TAACGTGTCG	TATGTGTGTC
18721	GGGTGCAATC	CCTGAAGCGC	CGACGAIGCI	CTCCTCACC	CCCCCCCCCC	CGCTTTCCAA
18781	ATGTATGCGT	CCATGTCGCC	MCCCCCACAC	CTGCTGAGCC	CACATCTCGG	GCCAGGACGC
18841	GATGGCTACC	CTGAGCCCCG	CCCTCCTCCA	GTTTTGCCCGC	GCCACCGAGA	CGTACTTCAG
18901	CTCGGAGTAC	AAGTTTAGAA	ACCCCACGCT	CCCCCCTACG	CACGACGTGA	CCACAGACCG
18961	CCTGAATAAC	TTGACGCTGC	CCETCATCC	TCTCCACCGT	GAGGATACTG	CGTACTCGTA
19021	GTCCCAGCG1	TIGACGCIGC	CHCHCCCC	TO ACCOTOTO	CTGGACATGG	CTTCCACGTA
19081	CAAGGCGCGG	TICACCCTAG	TO T	CCCTACTTT	AAGCCCTACT	CTGGCACTGC
19141	CTTTGACATC	CGCGGCGTGC	A CCCTTCCCCC	NAME	CAATGGGATG	AAGCTGCTAC
19201	CTACAACGCC	CIGGCICCCA	AGGGIGCCCC	CCATCACAA	CAACACGAAG	TAGACGAGCA
19261	TGCTCTTGAA	ATAAACCTAG	AAGAAGAGGGA	CCACCCCCCT	TATTCTGGTA	TAAATATTAC
19321	AGCTGAGCAG	CAAAAAACTC	WCGIWIIIGG	יייייייייייייייייייייייייייייייייייייי	AAATATGCCG	ATAAAACATT
19381	AAAGGAGGGT	ATTCAAATAG	GIGICGWAGG	CACCAPCE	ACTGAAATTA	ATCATGCAGC
19441	TCAACCTGAA	CCTCAAATAG	CHACACATOTOR	CANACCAM	TACGGTTCAT	ATGCAAAACC
19501	TGGGAGAGTC	CTTAAAAAGA	NACCCAMPO	1 DIAJORANDO 1 POR A A CONTROL	CANANTOCAN	AGCTAGAAAG
19561	. CACAAATGAA	AATGGAGGGC	AAGGCATICI	TGIAAAGCAC	CCAGCCAATG	GTGATAACTT
19621	. TCAAGTGGAA	A ATGCAATTTT	TCTCAACTAC	, <u>1</u> GAGGCGACC	CAAACCCCAG	ACACTCATAT
19681	. GACTCCTAA	A GIGGIATIGI	ACAGIGAAGA	CACTURE TO THE	CTAATGGGCC	AACAATCTAT
19741	TTCTTACATO	S CCCACTATTA	, AGGAAGGIAA	CICICGAGA		

						TGTATTACAA
						CTGTTGTAGA
			AGCTTTCATA			
			GGAATCAGGC			
20041	TATTGAAAAT	CATGGAACTG	AAGATGAACT	TCCAAATTAC	TGCTTTCCAC	TGGGAGGTGT
20101	GATTAATACA	GAGACTCTTA	CCAAGGTAAA	ACCTAAAACA	GGTCAGGAAA	ATGGATGGGA
20161	AAAAGATGCT	ACAGAATTTT	CAGATAAAAA	TGAAATAAGA	GTTGGAAATA	ATTTTGCCAT
20221	GGAAATCAAT	CTAAATGCCA	ACCTGTGGAG	AAATTTCCTG	TACTCCAACA	TAGCGCTGTA
			ACAGTCCTTC			
20341	CTACGACTAC	ATGAACAAGC	GAGTGGTGGC	TCCCGGGTTA	GTGGACTGCT	ACATTAACCT
20401	TGGAGCACGC	TGGTCCCTTG	ACTATATGGA	CAACGTCAAC	CCATTTAACC	ACCACCGCAA
			CAATGTTGCT			
20521	CCAGGTGCCT	CAGAAGTTCT	TTGCCATTAA	AAACCTCCTT	CTCCTGCCGG	GCTCATACAC
			AGGATGTTAA			
			GCATTAAGTT			
			CCTCCACGCT			
			TCTCCGCCGC			
			TCCCCTCCCG			
			AAACCCCATC			
			ACCTAGATGG			
			CTTCTGTCAG			
			GCTCAGTTGA			
			TGGTACAAAT			
			ACAAGGACCG			
			ATGATACTAA			
			GATTTGTTGG			
			CCTATCCGCT			
			ATCGCACCCT			
			ACCTGGGCCA			
			TGGATCCCAT			
			GTGTGCACCG			
			CCGGCAACGC			
			CAGTGAGCAG			
			CACCTATGAC			
21841	AAGCTCGCCT	GCGCCATAGT	CAATACGGCC	GGTCGCGAGA	CTGGGGGCGT	ACACTGGATG
21901	GCCTTTGCCT	GGAACCCGCA	CTCAAAAACA	TGCTACCTCT	TTGAGCCCTT	TGGCTTTTCT
21961	GACCAGCGAC	TCAAGCAGGT	TTACCAGTTT	GAGTACGAGT	CACTCCTGCG	CCGTAGCGCC
22021	ATTGCTTCTT	CCCCGACCG	CTGTATAACG	CTGGAAAAGT	CCACCCAAAG	CGTACAGGGG
22081	CCCAACTCGG	CCGCCTGTGG	ACTATTCTGC	TGCATGTTTC	TCCACGCCTT	TGCCAACTGG
22141	CCCCAAACTC	CCATGGATCA	CAACCCCACC	ATGAACCTTA	TTACCGGGGT	ACCCAACTCC
22201	ATGCTCAACA	GTCCCCAGGT	ACAGCCCACC	CTGCGTCGCA	ACCAGGAACA	GCTCTACAGC
22261	TTCCTGGAGC	GCCACTCGCC	CTACTTCCGC	AGCCACAGTG	CGCAGATTAG	GAGCGCCACT
			CATGTAAAAA			
			TCTCGGGTGA			
			CTGCCGCGCA			
			CCACTTAAAC			
			GCGCACCATC		-	
			GCCTCCGCCC			
			CGCCGGGTGG			
			CTCCGCGTTG			
			GTGCCCAGGC			
			CTGGGCGTTA			
			CTTTGCGCCT			
			GCCGCGTCG			
22701	AMONOCACCA	CAMMMCCCCC	CCACCGGTTC	100000000C	WCCLIGCOIC	PCPCACCACC
23U4T	MICIGCACCA	CHITICGGCC	CCMCCGG11C	I TCACGAICT	100CC11GCT	VOVC I OC I CC

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23101	TTCAGCGCGC	GCTGCCCGTT	TTCGCTCGTC	ACATCCATTT	TOTAL COCCA	CCCCTGCAGC
23161	ATCATAATGC	TTCCGTGTAG	ACACTTAAGC	TCGCCTTCGA	TCTCAGCGCA	CCACACCACC
03001	CACAACCCCC	A CCCCCCCCCCCCC	ርጥርርጥር አጥርር	TTGTAGGTCA	CCTCTGCAAA	CGACIGCAGG
23281	TACGCCTGCA	GGAATCGCCC	CATCATCGTC	ACAAAGGTCT	TGTTGCTGGT	A CCMMCCA CM
22241	MCC N N CCCCCC	CCTCCTCCTC	GTTCAGCCAG	GTCTTGCATA	CGGCCGCCAG	MGCIICCHCI
22401	ጥርርጥር እርርር እ	CTACTTTGAA	GTTCGCCTTT	AGATCGTTAT	CCACGTGGTA	CITGICCAIC
22461	ACCCCCCCCC	CACCCTCCAT	GCCCTTCTCC	CACGCAGACA	CGATCGGCAC	ACTUAGUGG
22521	ምምር እጥር እርርር	ጥል ልጥጥጥር ልርጥ	TTCCGCTTCG	CTGGGCTCTT	CCTCTTCCTC	110001000
22501	AMACCACCCC	CCACTGGGTC	GTCTTCATTC	AGCCGCCGCA	CTGTGCGCTT	ACCICCITIG
22641	CCATCCTTCA	TTACCACCGC	TEGETTECTE	AAACCCACCA	TTTGTAGCGC	CACATCITCI
22701	CUMMTCUMTCOM	CCCTCTCCCAC	GATTACCTCT	GGTGATGGCG	GGCGCTCGGG	CIIGGGAGAA
22761	CCCCCCTTCT	<u> ተመተመተመተመተ</u>	GGGCGCAATG	GCCAAATCCG	CCGCCGAGGT	CGATGGCCGC
22021	CCCCTCCCTC	TGCGCGGCAC	CAGCGCGTCT	TGTGATGAGT	CTTCCTCGTC	CICGGACICG
22001	A TO A C C C C C C C C	ጥር ልጥር ርርር ርጥጥ	TTTTGGGGGC	GCCCGGGGAG	GCGGCGGCGA	CGGGGACGGG
22041	CACCACACCT	CCTCCATGGT	TGGGGGACGT	CGCGCCGCAC	CGCGTCCGCG	CICGGGGGIG
24001	CHARACCCCCC	ርርጥርርጥርጥጥር	CCGACTGGCC	ATTTCCTTCT	CCTATAGGCA	GAAAAAGAIC
24061	አመድሮ አርመሮ አር	ጥሮርልርልልርልል	GGACAGCCTA	ACCGCCCCT	CTGAGTTCGC	CACCACCGCC
24121	THE THE PROPERTY OF THE PROPER	CCCCCAACGC	GCCTACCACC	TTCCCCGTCG	AGGCACCCC	GCIIGAGGAG
24101	CACCAACTCA	TTATCGAGCA	GGACCCAGGT	TTTGTAAGCG	AAGACGACGA	GGACCGCICA
24241	CHACCAACAC	ACCATAAAA	GCAAGACCAG	GACAACGCAG	AGGCAAACGA	GGAACAAGIC
24201	CCCCCCCCCCCC	ACGAAAGGCA	TGGCGACTAC	CTAGATGTGG	GAGACGACGT	GCTGTTGAAG
24261	CARCTCCACC	CCCACTCCCC	CATTATCTGC	GACGCGTTGC	AAGAGCGCAG	CGATGIGCCC
24421	CTCCCCATAG	CCCATCTCAG	CCTTGCCTAC	GAACGCCACC	TATTCTCACC	GCGCGTACCC
24401	CCCNNNCCCC	AACAAAACGG	CACATGCGAG	CCCAACCCGC	GCCTCAACTT	CTACCCCGIA
24541	MMMCCCCCCCC	CACACCTCCT	TCCCACCTAT	CACATCTTTT	TCCAAAACTG	CAAGATACCC
24601	CTATCCTCCC	GTGCCAACCG	CAGCCGAGCG	GACAAGCAGC	TGGCCTTGCG	GCAGGGGGC1
24561	ርጥር እጥ እርርጥር	ልጥልጥሮርሮሮጥሮ	GCTCAACGAA	GTGCCAAAAA	TCTTTGAGGG	TCTTGGACGC
24721	CACCACAACC	CCCCCCCAAA	CGCTCTGCAA	CAGGAAAACA	GCGAAAATGA	AAGICACICI
24701	CCACTCTTCC	TCCAACTCGA	GGGTGACAAC	GCGCGCCTAG	CCGTACTAAA	ACGCAGCAIC
24041	CACCTCACCC	ልሮጥጥጥርሮሮጥ ል	CCCGGCACTT	AACCTACCCC	CCAAGGTCAT	GAGCACAGIC
24001	አጥር እርጥር እርር	TCATCGTGCG	CCGTGCGCAG	CCCCTGGAGA	GGGATGCAAA	1-1-1-GCAAGAA
24061	CANACAGAGG	ACCCCCTACC	CGCAGTTGGC	GACGAGCAGC	TAGCGCGCTG	GCTTCAAACG
25021	CCCCACCCTC	CCCACTTGGA	GGAGCGACGC	AAACTAATGA	TGGCCGCAGT	GCTCGTTACC
25001	CTCCACCTTC	ACTCCATCCA	GCGGTTCTTT	GCTGACCCGG	AGATGCAGCG	CAAGCTAGAG
25141	ር እ እ እር እጥጥርር	ልሮ ሞልሮልሮሮሞሞ	TCGACAGGGC	TACGTACGCC	AGGCCTGCAA	GATCICCAAC
25201	CTCCACCTCT	CCAACCTGGT	CTCCTACCTT	GGAATTTTGC	ACGAAAACCG	CCTTGGGCAA
25261	እ አ ርርጥርር ርጥጥር	ልጥጥርሮልሮርሮ	CAAGGGCGAG	GCGCGCCGCG	ACTACGTCCG	CGACTGCGTT
25221	ጥ አ ር ጥጥ አ ጥጥጥር	ጥልጥርርጥልሮልሮ	CTGGCAGACG	GCCATGGGCG	TTTGGCAGCA	GTGCTTGGAG
25201	ር አርጥርር አ አርር	TO A ACCACOT	GCAGAAACTG	CTAAAGCAAA	ACTTGAAGGA	CCTATGGACG
25443	CCCMMCAACC	ልሮሮርርጥሮርርጥ	GGCCGCGCAC	CTGGCGGACA	TCATTTCCC	CGAACGCCIG
25501	ርጥጥአ አ አ አርርር	TCCDDCAGGG	TCTGCCAGAC	TTCACCAGTC	AAAGCATGTT	GCAGAACIII
25561	አ ርር አ አ ርጥጥጥ እ	TCCTACACCC	CTCAGGAATC	TTGCCCGCCA	CCTGCTGTGC	ACTICCIAGC
25621	CACOOOCCOC	·	CCCCGAATGC	CCTCCGCCGC	TTTGGGGCCA	CIGCIACCII
25601	CTCCACCTAC	CCAACTACCT	TGCCTACCAC	TCTGACATAA	TGGAAGAUGT	GAGCGGIGAC
25741	CCTCTACTCC	አርጥርጥሮልርጥ ር	TCGCTGCAAC	CTATGCACCC	CGCACCGCTC	CCIGGIIIGC
25001	እ አጥጥር <u>ርር</u> አርር	· ጥርርጥጥልልሮGA	AAGTCAAATT	ATCGGTACCT	' TTGAGCTGCA	GGGTCCCTCG
25001	CCTCACCAG	ACTOCIONO	TCCGGGGTTG	AAACTCACTC	CGGGGCTGTG	GACGTCGGCT
72801	TARBURAL OF THE	AGICCGCGCC Agrammaaa	TGAGGACTAC	CACGCCCACG	AGATTAGGTT	CTACGAAGAC
25001	CAATCCCCCC	CCCCAAATGC	GGAGCTTACC	GCCTGCGTCA	TTACCCAGGG	CCACATTCTT
72227	CCCCANDOCC	, CGCCAAAIGC Bacccaacaa	CAAAGCCCCGC	CAAGAGTTTC	TGCTACGAAA	GGGACGGGG
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20101	TATCAGCAGC	, AGCCGCGGGC	L ACCACCAATA	CTGGGACAGT	CAGGCAGAGG	AGGTTTTGGA
20221	10000000000000000000000000000000000000	T CACCACAGACA	TCCAACACTC	GGAGAGCCTA	GACGAGGAAG	CTTCCGAGGT
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26341	CGAAGAGGTC	, ICAGACGAAP	CACCGICACC			

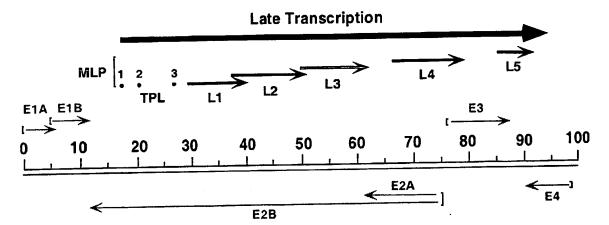
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CATCTACCAC AAATACACCC L ACCGGACTTA CATCTACCAC AAATACACCC L ACCGGACTTA CATCTACCAC AAATACACCC L ACCGGACTTA CATCTACCAC AAATACACCC L ACCTTGGGCAC TGTTGTGGTGTT CTCCATAGGGA	L GTCCCGCTCC CACCACTGTG GTACTTCCCA GAGACGCCCA L ACTCAGGGGC GCAGCTTGCG GGCGGCTTTC TAACTCACCT GACAATCAGA GGGCGAGGTA TTCAGCTCAA CGCTTGGTCT CCGTCCGGAC GGGACATTTC AGATCGGCGG CGCCTCGTCA GGCAATCCTA ACTCTGCAGA CCTCGTCCTC TTGGAACTCT GCAATCATA TGAGGAGTTTG TGCCATCGGT TTGGAACTCT GCAATCATA TGAGGAGTTTG TGCCATCGGT CGGACGGCTA CGACTGAATG TTAAGTGGAG AGGCAGAGCA TCCACTGTCG CCGCCACAAG TGCTTTGCCC GCGACTCCGG TGCCCGAGGA TCATATCGAG GGCCCGGCGC ACGGCGTCCG ACCCCGTAG CCTGATTCGG GAGTTTACCC AGCGCCCCCT TGCCCGAGGA TCATATCGAG GGCCCGGCGC ACGGCGTCCG GACCCTGTGT TCTCACTGTG ATTTGCAACT GTCCTAACCT TACACGCATC TCTGCTGAG TATAATAAAT ACAGAAATTA CGGCATCCTG TAACACCCAC CGTCTTCACC CGCCCAAGCA CGCCATCTT TACACCCAC CGTCTTCACC CGCCCAAGCA TGCCGGGAC GTACGAGTGC GTCACCGCC GCTGCACCAC CCAGACTTTT TCCGGACAGA CCTCAATAAC TCTGTTTACC AAAACCCTTA GGGTATTCTA ATTCAGGTT TCCATCAGAA CCACAGAGA CCTCAATAAC TCTGTTTACC AAAACCCTTA GGGTATTCTA ATTCAGGTT TCCATCAGAA CAACTCTACG GGCTATTCTA ATTCAGGTT TCCATCAGAA CAACTCTACG GGCTATTCTA ATTCAGGTT TCTTAGAATC CTATGGAATA TCCTTAGGTT ACTCACCCT TCTTAGAATC CTATGTAAAAT TCCTAGGTTT ACTCACCCT TAACACGTT AAGGTACATAA TCCTAGGTTT ACTCACCCT TAACACCTT AAGGTACATAA TCCTAGGTTT ACTCACCCTT TAAACGCTGG CTTATAAAAT GCCACACAA ACATGAAAAG CTGCTTATTC AAGTATGCTG TTTATCCTAACCCCT TAATGTTACA AAGTATGCTG TTTATGCTAT TTGGCAGCCA GGTGACACTA AAGTATGCTG TTTATGCTAT TTGGCAGCCA GTGTACACTA AAGTATGCTA AAAGTCATAA ACATTAAAA TTGCCAGCTA TTTTTCCAGGGTA AAAGTCATAA ACATTAAAA TTGCCAGCCC TTATTTTC ACTCGAGCTT TCTGCTGCAC TGCTATTACT TTTTTTCAGGCCC CTCTATATTA AAAGCCACAAA ACAGTATAAG TTGCCAGCTC CTCTATATTA AAAGCCACAAA ACAGTATAAG TTGTGGCCCC CTCTATATTA AAAGCTAATA TTGGCAGCCA TTTTTTGAGG ACTAGGACTT TCTGCTGCAC TGCTATTACT TTTTTTTCAGGCACCTA ATTACACTTT TTTTTTCAGAGTACACAAAAAG CAGCACACAA ACAGTATAAG TTTTACAGTGC CTCTATATTA AAAGCTAAAAAAAAAAAAAAAAAAAAAAA	CCACACCTCG TAATAACCTT AATCCCGTA GTTGGCCCGC TGCCCTGGTG GTCCCGCTCC CACCACTGTG GTACTTCCCA GAGACGCCA GGCCGAAGTT ACTCAGGGGC GACACTCGG GGCGCTTTC TAACTCACCT GACAATCAGA GGGCGAGGTA TTCACCTCAA CGACCAGTCG CGCTTGGTCT CCGTCCGGAC GGCACTTCC AGATCGCGC GCCCGCCGT CGCCTGGTCT CCGTCCGGAC GGCACTTCC AGATCGCGC CGCCCGTCTCT TGAGCCGGC CGCCTGGTCA GGCAATCCTA ACTCTGCAGA CCTCGTCCT TGAGCCGGC CCACTATCCG GACAATTATT GAGGAGTTTG TGCCATCGGT CTACTTTAAC GACCTCCCGG CCACTATCCG GATCAATTTA TTCCTAACTT TGACCGGGTA CGGCGGCCACAAG TCCTTTGCCC GCGACCACCG TGAGTTTTGC CGCCGAGGA TCATATCGA GGCCCGGCG ACGACTCCG TGAGTTTGC TCCACTGTC CCGCCACAAG TGCTTTGCCC GCGACCCCCC GCTACCTGGT TTCACCGTAG CCTGATTCGG GAGTTACCC AGCGCCCCCCT GCTACTTGGC GACCCTGTCT TCTCACTGTG ATTTGCAACT GTCCTAACCT TGGATTACAT GTTGCCCGTAG CCTGATTCGG GAGTTACCC AGCGCCCCCCT GCTAGTTGGC GGCCATCCTG TAAACGCCAC CGTCTACCC GCCCAAGCA AACCAACGCG CGCATCCTG TAAACGCCAC CGTCTCACC CGCCCAAGCA AACCAACGAG CGTACTCTTA ACATCTCCC CTCTGTGATT TACAACAGTT TCAACCAGA CCTACGAGAAAACCCTTA ACATCTCCC CTCTAGCTACCT TACACCAGA AAAACCCTTA ACATCTCCC CTCTAGCTAC TCCATCAGAA AAACACACCAC CACACCTTTT TCCGGACAGA CCTCAATAAC TCCATCAGAA AAACACCAC CAACCCTTA GGGTATTAGG CCAAAGGCGC GCTCACCAC ACCTACCGCC CCAGACTTTT TCCGGACAGA CCTCAATAAC TCCTTTTACC CACCTCTCG GGCTATTCTA ATTCAGGTTT TCCTAGAATC GGGTTATGAA AAAACCCTTA GGGTATTCTA ATTCAGGTTT TCTTAGAACAGCGC CACACCACAA ACCCTTATACC TCTTTATTC TTATACTAAC GCTCACCTGC GGGTTATGAA CAACCCTTA TGCATTTATT GTCAACCTTTTACC GGGTTAGCAC CAACCCTTAAAACCCTTA ACCCAAGAACACCTT TTATACTATCA TCCTAGACACACACACACACACACACACACACACACACAC

		ACAATGATGG	**************************************	TTCCACCCAC	тсааасасат	GTTCTTTTCT
29701	CTACACCCAA	GATTAAATGA	CACATCATAGA	CTCCACTTTT	TATATTACTG	ACCCTTGTTG
29761	CTTACAGTAT	TGCGTGCTCC	ACATGATIC	CCCTTTCTC	CATCGAAGTA	GACTGCATTC
29821	CGCTTTTTTG	AGTCTATTTG	ACATIGGCIG	CGGIIICICA CGGIIICICA	CACCCTCATC	TGCAGCCTCA
29881	CAGCCTTCAC	AGTCTATTTG	A TOTAL COURT	TIGICACCCI TOTCACCCCT	CTCTCTCTCCCC	TTTGCATATC
29941	TCACTGTGGT	CATCGCCTTT TCCCCAGTAC	ATCCAGTGCA	CONTROCTOR	CCTTCTTAGA	ATTCTTTAAT
30001	TCAGACACCA	TCCCCAGTAC	AGGGACAGGA	TATAGCIGA TATAGCIGA	CTATCTCCCT	TTTGTTCCCC
30061	TATGAAATTT	ACTGTGACTT	TTCTGCTGAT	CACATTCACT	CCTATCIGCGI	ATATTCCAAG
30121	GACCTCCAAG	CCTCAAAGAC GAAAAAAGCG	ATATATCATG	ACCCTCCTTA	TATCCAATCA	TCTCTGTTAT
30181	TTGCTACAAT	AGTACCATCT	MACCOCCON CC	WACCIGGIIV	TACCOTTCACA	TTGGCTGGAA
30241	GGTGTTCTGC	GCCATGAACC	ACCCAACCCT	CCCCCCCCCCCC	CCTATCCTTC	CACTGCAACA
30301	ACGAATAGAT	GCCATGAACC	ACCCAACTTT	TCACCCTCGC	CCCACTTCTC	CCACCCCCAC
30361	AGTTGTTGCC	TACTTTAATC	TUCCAGUCAA	ACAMCACMCA	CACCCTAGAT	CTAGAAATGG
30421	TGAAATCAGC	TACTTTAATC	TAACAGGAGG	ANACACCCAC	CACCACTRICATE	GAGCAACAGC
30481	ACGGAATTAT	TACAGAGCAG AGAGCTCCAA	CGCCTGCTAG	AAAGACGCAG	CTCCAAAAGG	CCTATCTTTT
30541	GCATGAATCA	AGAGCTCCAA	GACATGGTTA	ACTIGCACCA ACACTA ATAC	CACCGGACAC	CGCCTTAGCT
30601	GTCTGGTAAA	GCAGGCCAAA	GICACCTACG	MCAGIAAIAC	CCCACAAAAC	CCCATTACCA
30661	ACAAGTTGCC	AACCAAGCGT	CAGAAATTGG	CCARTCACTC	ACCUTCTCAA	GGACCTGAGG
30721	TAACTCAGCA	CTCGGTAGAA	ACCGAAGGCT	COCOCO	TOTTO TOTO	ΤΤΤΑΑСΤΑΑΤ
30781	ATCTCTGCAC	CCTTATTAAG	ACCCTGTGCG	A A THE A CHITTA	CCAAATTTCT	CTCCAGTTTA
30841	AAAAAAAAT	AATAAAGCAT	CACTTACTTA	COCOCCOATA	CCACCTTCCT	CCTGGCTGCA
30901	TTCAGCAGCA	CCTCCTTGCC	TOCAL A TOTAL	CICIGGIAII	CTTCCTCTCC	ATCCGCACCC
30961	AACTTTCTCC	ACAATCTAAA	TGGAATGTCA	GITICCICCI	CTCAACATAC	CTTCAACCCC
31021	ACTATCTTCA	TGTTGTTGCA	GATGAAGCGC	GCAAGACCGI	CIGAAGAIAC	ጥርርጥርርርጥጥጥ
31081	GTGTATCCAT	ATGACACGGA	AACCGGTCCT	CCAACTGIGC	TOTO TOTO TO	CCTATCCGAA
31141	GTATCCCCCA	ATGGGTTTCA	AGAGAGTCCC	CCTGGGGTAC	CCAACCCCCT	CTCTCTGGAC
31201	CCTCTAGTTA	CCTCCAATGG	CATGCTTGCG	CTCAAAATGG	CCCCACCTCT	CAAAAAAACC
31261	GAGGCCGGCA	ACCTTACCTC	CCAAAATGTA	ACCACTGTGA	TTACCTCACA	ACCCCTAACT
31321	AAGTCAAACA	TAAACCTGGA	AATATCTGCA	CCCCTCACAG	TIACCICAGA	AGCCCIAACI
31381	GTGGCTGCCG	CCGCACCTCT	AATGGTCGCG	GGCAACACAC	A A C C A C C C C C T	CACACTCTCA
31441	CCGCTAACCG	TGCACGACTC	CAAACTTAGC	ATTIGCCACCC	CCACCCATAC	CACAGIGICA
31501	GAAGGAAAGC	TAGCCCTGCA	AACATCAGGC	CCCCTCACCA	CCACCGATAG	TCACTTCAAA
31561	ACTATCACTG	CCTCACCCCC	TCTAACTACT	GCCACTGGTA	ACCCCCCTCC	TORCITORIA
31621	GAGCCCATTT	ATACACAAAA	TGGAAAACTA	GGACTAAAGT	CTCCCC CTCC	TITGCATGIA
31681	ACAGACGACC	TAAACACTTT	GACCGTAGCA	ACTGGTCCAG	GIGIGACIAI	TAMIANIACI TATIONALACI
31741	TCCTTGCAAA	CTAAAGTTAC	TGGAGCCTTG	GGTTTTGATT	CACAAGGCAA	ተርያብርርኒኒርር
31801	AATGTAGCAG	GAGGACTAAG	GATTGATTCT	CAAAACAGAC	GCCTTATACT	ጥርጥጥጥጥስጥል ፕሬአተርያ፣ ነላሪ፣
31861	TATCCGTTTG	ATGCTCAAAA	CCAACTAAAT	CTAAGACTAG	THE TANK OF THE TA	ጥልሮልርርጥጥሮል
31921	AACTCAGCCC	ACAACTTGGA	TATTAACTAC	AACAAAGGCC	ACCCCCCCCC	CUTTCACCTCA
31981	AACAATTCCA	AAAAGCTTGA	GGTTAACCTA	AGCACTGCCA	AGGGGIIGAI	TGCACCAAAC
32041	ACAGCCATAG	CCATTAATGC	AGGAGATGGG	CTTGAATTTG	GITCACCIAA	CARCCAMC
32101	ACAAATCCCC	TCAAAACAAA	AATTGGCCAT	GGCCTAGAAT	CTCCCATTAC	ACTACCAAAC
32161	GTTCCTAAAC	TAGGAACTGG	CCTTAGTTTT	GACAGCACAG	CAMCMCCMAA	CTCTACACTA
32221	DTAATAAAA	ATAAGCTAAC	TTTGTGGACC	ACACCAGCTC	ANDCOCCOAC	CTGTAGACTA
32281	AATGCAGAGA	AAGATGCTAA	ACTCACTTTG	GTCTTAACAA	CARDIGGCAG	TCAAATACTT
32341	GCTACAGTTT	CAGTTTTGGC	TGTTAAAGGC	AGTTTGGCTC	CAMIMICIOG	AACAGTTCAA
32401	AGTGCTCATC	TTATTATAAG	ATTTGACGAA	AATGGAGTGC	TACTAAACAA	TTCCTTCCTG
32461	GACCCAGAAT	ATTGGAACTT	TAGAAATGGA	GATCTTACTG	AAGGCACAGC	CTATACAAAC
32521	GCTGTTGGAT	TTATGCCTAA	CCTATCAGCT	TATCCAAAAT	CTCACGGTAA	AACTGCCAAA
32581	AGTAACATTG	TCAGTCAAGT	TTACTTAAAC	GGAGACAAAA	CTAAACCTGI	AACACTAACC
22641	አ ጥጥ አ ር አ ር ጥ አ አ	ACCCTACACA	GGAAACAGGA	GACACAACTO	CAAGTGCATA	CICIAIGICA
32701	TTTTCATGGG	ACTGGTCTGG	CCACAACTAC	ATTAATGAAA	TATTTGCCAC	ATCCTCTTAC
22761	እርጥጥጥጥ ተር እባ	' እር <u>እጥፕ</u> ርርርር	. AGAATAAAGA	ATCGTTTGTG	TTATGTTTCA	ACGIGITIAL
22021	መመመመር እ አመጥር	י ראכאאאדיי	CAAGTCATTI	' TTCATTCAGI	' AGTATAGCCC	CALLACTACA
22001	መስ ርርመመስ መስር	* ACATCACCGT	· ACCTTAATCA	. AACTCACAGA	ACCCTAGTAL	TUAACCIGCC
32941	ACCTCCCTCC	CAACACACAC	AGTACACAGI	CCTTTCTCCC	. CGGCTGGCCT	TAAAAAGCAT

33001	CATATCATGG	GTAACAGACA	TATTCTTAGG	TGTTATATTC	CACACGGTTT	CCTGTCGAGC
33061	CAAACGCTCA	TCAGTGATAT	TAATAAACTC	CCCGGGCAGC	TCACTTAAGT	TCATGTCGCT
33121	GTCCAGCTGC	TGAGCCACAG	GCTGCTGTCC	AACTTGCGGT	TGCTTAACGG	GCGGCGAAGG
33181	AGAAGTCCAC	GCCTACATGG	GGGTAGAGTC	ATAATCGTGC	ATCAGGATAG	GGCGGTGGTG
33241	CTGCAGCAGC	GCGCGAATAA	ACTGCTGCCG	CCGCCGCTCC	GTCCTGCAGG	AATACAACAT
33301	GGCAGTGGTC	TCCTCAGCGA	TGATTCGCAC	CGCCCGCAGC	ATAAGGCGCC	TTGTCCTCCG
33361	GGCACAGCAG	CGCACCCTGA	TCTCACTTAA	ATCAGCACAG	TAACTGCAGC	ACAGCACCAC
33421	AATATTGTTC	AAAATCCCAC	AGTGCAAGGC	GCTGTATCCA	AAGCTCATGG	CGGGGACCAC
33481	AGAACCCACG	TGGCCATCAT	ACCACAAGCG	CAGGTAGATT	AAGTGGCGAC	CCCTCATAAA
33541	CACGCTGGAC	ATAAACATTA	CCTCTTTTGG	CATGTTGTAA	TTCACCACCT	CCCGGTACCA
33601	TATAAACCTC	TGATTAAACA	TGGCGCCATC	CACCACCATC	CTAAACCAGC	TGGCCAAAAC
33661	CTGCCCGCCG	GCTATACACT	GCAGGGAACC	GGGACTGGAA	CAATGACAGT	GGAGAGCCCA
33721	GGACTCGTAA	CCATGGATCA	TCATGCTCGT	CATGATATCA	ATGTTGGCAC	AACACAGGCA
33781	CACGTGCATA	CACTTCCTCA	GGATTACAAG	CTCCTCCCGC	GTTAGAACCA	TATCCCAGGG
33841	AACAACCCAT	TCCTGAATCA	GCGTAAATCC	CACACTGCAG	GGAAGACCTC	GCACGTAACT
	CACGTTGTGC					
33961	AGCGCGGGTT	TCTGTCTCAA	AAGGAGGTAG	ACGATCCCTA	CTGTACGGAG	TGCGCCGAGA
	CAACCGAGAT					
	TCCTGAAGCA					
	TAGATCGCTC					
34201	TGGCTTCGGG	TTCTATGTAA	ACTCCTTCAT	GCGCCGCTGC	CCTGATAACA	TCCACCACCG
34261	CAGAATAAGC	CACACCCAGC	CAACCTACAC	ATTCGTTCTG	CGAGTCACAC	ACGGGAGGAG
34321	CGGGAAGAGC	TGGAAGAACC	ATGTTTTTTT	TTTTATTCCA	AAAGATTATC	CAAAACCTCA
34381	AAATGAAGAT	CTATTAAGTG	AACGCGCTCC	CCTCCGGTGG	CGTGGTCAAA	CTCTACAGCC
34441	AAAGAACAGA	TAATGGCATT	TGTAAGATGT	TGCACAATGG	CTTCCAAAAG	GCAAACGGCC
34501	CTCACGTCCA	AGTGGACGTA	AAGGCTAAAC	CCTTCAGGGT	GAATCTCCTC	TATAAACATT
34561	CCAGCACCTT	CAACCATGCC	CAAATAATTC	TCATCTCGCC	ACCTTCTCAA	TATATCTCTA
	AGCAAATCCC					
	TTCAGCCTCA					
	GATTCAAAAG					
34801	GCTGAACATA	ATCGTGCAGG	TCTGCACGGA	CCAGCGCGGC	CACTTCCCCG	CCAGGAACCT
	TGACAAAAGA					
34921	CCCCGATGTA	AGCTTTGTTG	CATGGGCGGC	GATATAAAAT	GCAAGGTGCT	GCTCAAAAAA
34981	TCAGGCAAAG	CCTCGCGCAA	AAAAGAAAGC	ACATCGTAGT	CATGCTCATG	CAGATAAAGG
35041	CAGGTAAGCT	CCGGAACCAC	CACAGAAAAA	GACACCATTT	TTCTCTCAAA	CATGTCTGCG
35101	GGTTTCTGCA	TAAACACAAA	ATAAAATAAC	AAAAAAACAT	TTAAACATTA	GAAGCCTGTC
35161	TTACAACAGG	AAAAACAACC	CTTATAAGCA	TAAGACGGAC	TACGGCCATG	CCGGCGTGAC
35221	CGTAAAAAAA	CTGGTCACCG	TGATTAAAAA	GCACCACCGA	CAGCTCCTCG	GTCATGTCCG
	GAGTCATAAT					
	CGACCGAAAT					
	ATAGGAGGTA					
	TGCCTAGGCA					
35521	CCTAACAGTC	AGCCTTACCA	GTAAAAAAGA	AAACCTATTA	AAAAAACACC	ACTCGACACG
35581	GCACCAGCTC	AATCAGTCAC	AGTGTAAAAA	AGGGCCAAGT	GCAGAGCGAG	TATATATAGG
35641	ACTAAAAAAT	GACGTAACGG	TTAAAGTCCA	CAAAAAACAC	CCAGAAAACC	GCACGCGAAC
	CTACGCCCAG					
35761	CCCACGTTAC	GTAACTTCCC	ATTTTAAGAA	AACTACAATT	CCCAACACAT	ACAAGTTACT
35821	CCGCCCTAAA	ACCTACGTCA	CCCGCCCCGT	TCCCACGCCC	CGCGCCACGT	CACAAACTCC
35881	ACCCCCTCAT	TATCATATTG	GCTTCAATCC	AAAATAAGGT	ATATTATTGA	TGATG

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Structure of the Ad6 Genome



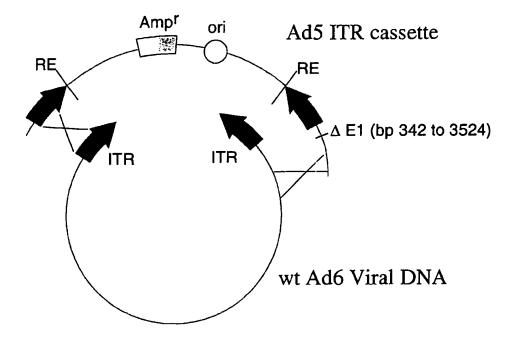
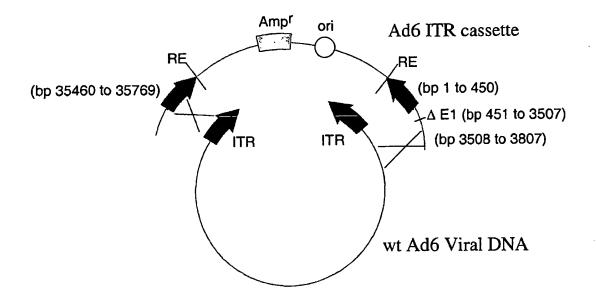
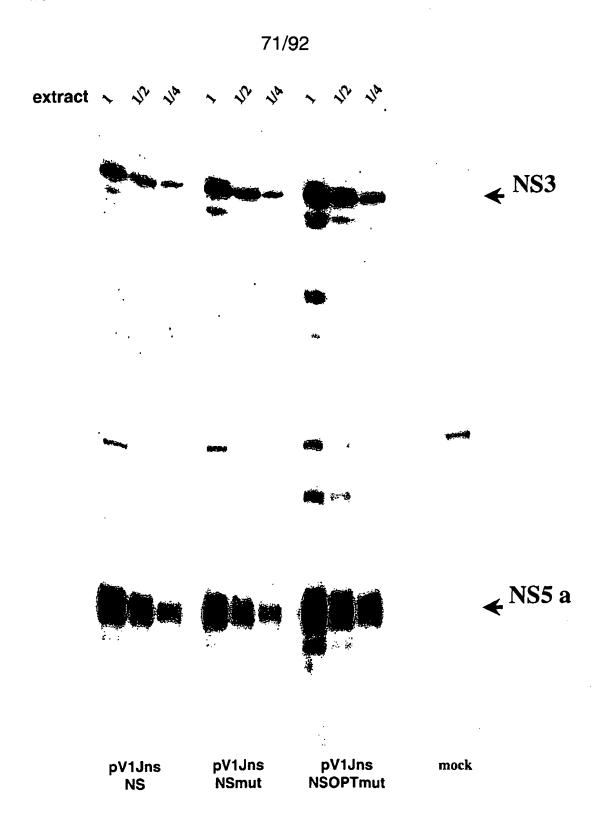


FIG. 10





Western blot on whole-cell extracts from 293 cells transfected with plasmid DNA expressing the different HCV NS cassettes. Mature NS3 and NS5A products were detected with specific antibodies.

FIG. 12

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					Pep pool				
	mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L(NS35b)	M(NS5b)	1480(CD8 ep)	DMSC
	#31	41	135	19	44	25	17	137	8
V44 NG	#32	121	783	77	144	13	22	604	4
	#33	8	32	3	11	6	6	43	3
	#34	16	139	13	47	31	25	151	2
	#35	21	101	40	32	21	20	75	1
pV1jns-NS	#36	18	26	24	25	5	7	29	6
	#37	19	73	15	39	8	20	49	2
	#38	133	575	74	345	75	63	515	5
	#39	40	183	10	85	14	9	148	2
	#40	66	465	29	111	15	16	189	0
	Geomean	33	146	21	57	15	16	123	na

					Pep pool				
	mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L(NS35b)	M(NS5b)	1480(CD8 ep)	DMSC
	#41	39	293	58	187	5	4	248	1
	#42	21	220	46	107	26	10	189	4
	#43	76	134	12	78	8	6	144	2
	#44	30	45	20	52	4	8	40	4
pV1jns-NSmut	#45	36	100	17	56	4	6	116	3
•	#46	67	172	16	138	8	9	145	3
	#47	34	131	28	38	9	5	118	1
•	#48	55	316	43	107	9	7	277	5
	#49	6	131	5	25	4	1	91	0
	#49 #50	13	93	11	11	5	1	76	1
	Geomean	30	142	20	61	7	5	126	na

					Pep pool				
	mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L(NS35b)	M(NS5b)	1480(CD8 ep)	DMSC
	#51	53	409	34	84	11	25	271	4
	#52	140	660	65	276	23	36	377	2
	#53	58	553	48	105	23	18	564	1
	#54	50	105	35	134	10	16	80	2
V1jns-NSOPTmut	#55	14	80	11	35	4	7	91	6
•	#56	14	342	30	101	23	14	207	1
	#57	63	325	66	239	17	24	123	1
	#58	75	542	66	168	127	93	191	0
	#59	65	468	40	124	18	23	344	4
	#60	27	142	48	16	7	8	77	0
	Geomean	45	295	40	99	16	20	188	na

IFNY ELIspot on splenocytes from C57black6 mice immunized with two injections of 25µg DNA/dose with GET of plasmid vectors expressing the different HCV NS cassettes. Data are expressed as SFC/106 PBMC.

FIG. 13A

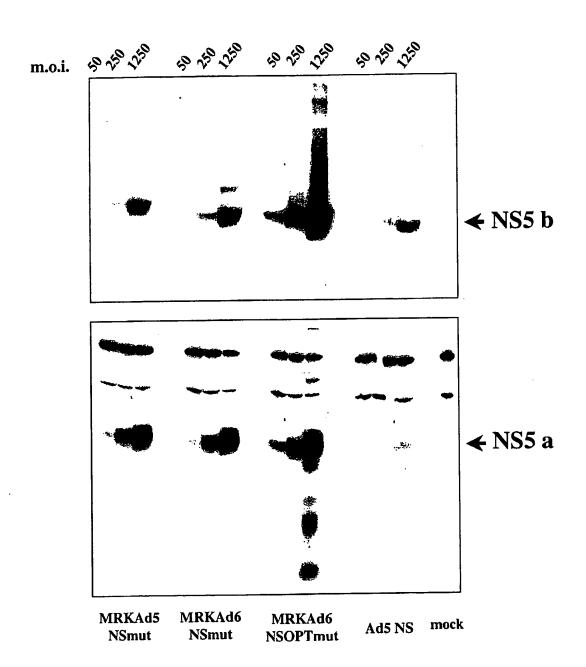
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		Pep pool						
	mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L(NS35b)	M(NS5b)	DMSO
	#51	219	699	634	486	487	264	34
	#52	67	302	347	167	111	87	9
	#53	59	460	400	246	244	136	26
	#54	139	817	685	236	547	223	24
	#55	96	904	542	277	256	337	17
pV1jns-NS	#56	225	603	686	156	350	240	56
	#57	44	288	211	148	100	141	4
	#58	37	262	221	53	58	62	3
	#59	131	975	928	159	305	284	14
	#60	93	475	464	77	206	113	12
	geo mean	111	579	512	201	266	189	20
		Pep pool						
	mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L(NS35b)	M(NS5b)	DMSO
	#61	72	840	515	219	278	249	19
	#62	294	1881	1266	365	434	411	63
	#63	73	415	422	103	141	99	41
	#64	66	824	486	175	162	144	18
pV1jns-NSmut	#66	24	313	168	53	47	42	5
	#67	15	230	253	94	25	39	2
	#68	53	354	252	89	101	86	15
	#69	271	895	909	518	322	285	74
	#70	417	1303	1186	468	557	267	34
	geo mean	143	784	606	232	230	180	30
				Pe	p pool			
	mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L(NS35b)	M(NS5b)	DMSO
	#71	206	944	890	342	207	397	47
	#72	393	1655	1151	575	626	401	72
	#73	123	522	515	319	223	198	21
NOODE .	#74	500	1414	1419	878	1035	1122	137
V1jns-NSOPTmut	#75	286	812	873	382	543	267	31
	#76	224	1143	942	218	420	281	22
	#77	95	643	630	169	385	218	15
	#78	401	1302	1068	538	608	623	12
	#79	108	1190	914	199	265	215	4
	#80	122	511	546	189	286	190	13
	geo mean	209	941	854	331	406	329	24

IFNy ELIspot on splenocytes from BalbC mice immunized with two injections of 50µg DNA/dose with GET of plasmid vectors expressing the different HCV NS cassettes. Data are expressed as SFC/10⁶ PBMC.

FIG. 13B

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Western blot on whole-cell extracts from HeLa cells infected at different multiplicity of infection (m.o.i.; indicated at the top) with Adenovectors expressing the different HCV NS cassettes. Mature NS5B and NS5A products were detected with specific antibodies.

FIG. 14

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				Pep pool			
mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L+M(NS35b)	1480(CD8	ep)DMSO
#1	14	492	9	27	10	554	7
#2	8	440	2	26	5	438	0
#3	12	92	5	12	7	73	4
#4	16	388	6	40	6	228	2
#6	8	210	4	31	3	238	3
#7	7	133	13	16	0	128	9
#8	11	342	25	55	22	267	12
#9	5	345	0	45	5	285	3
#10	22	888	3	65	25	799	1
Geomean	10	305	na	31	na	269	na

MRKAd5-NSmut

Ad5-NS

	•			Pep poor			
mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L+M(NS35b)	1480(CD8	ep)DMSO
#11	14	1009	13	75	7	751	6
#12	15	695	3	39	9	552	1
#13	12	389	4	20	7	352	3
#14	7	459	6	50	1	274	1
#15	5	549	3	22	6	485	0
#16	10	631	1	6	4	600	3
#17	5	257	3	9	1	245	3
#18	13	659	6	43	7	555	1
#19	12	758	1	37	5	669	0
#20	22	1380	5	163	8	1003	4
Geomean	10	615	3	31	4	504	na

				Pep pool			
mouse	F(NS3p)	G(NS3h)	H(NS4)	I(NS5a)	L+M(NS35b)	1480(CD8	ep)DMSO
#21	6	584	5	27	4	491	2
#22	6	231	3	12	3	235	0
#23	8	482	1	18	1	511	0
#24	14	1120	6	38	10	1004	5
#25	1	311	3	9	0	382	1
#26	29	903	3	60	5	751	5
#27	35	1573	4	40	4	1277	4
#28	7	406	5	15	1	443	3
#29	4	461	3	12	3	515	_ 3
Beomean	8	567	3	21	па	554	na

MRKAd6-NSmut

IFNy ELISPOT on splenocytes from C57black6 mice immunized with two injections of 10^9 vp/dose of Adenovectors expressing the different HCV NS cassettes. Data are expressed as SFC/ 10^6 PBMC.

FIG. 15

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	Ads	5-NS 10 ¹⁰ vp/d	ose
Pep pools	96074	134T	063Q
F (NS3p)	374	11	74
G (NS3h)	359	1070	1455
H (NS4)	376	30	64
I (NS5a)	240	40	63
L (NS5b)	226	29	121
M (NS5b)	511	23	35
DMSO	128	3	31

	MRK Ad	MRK Ad6-NSmut 10 ¹⁰ vp/dose					
Pep pools	S207	035Q	057 Q				
F (NS3p)	363	382	150				
G (NS3h)	180	316	119				
H (NS4)	126	113	62				
1 (NS5a)	1780	688	114				
L (NS5b)	447	111	81				
M (NS5b)	153	38	16				
DMSO	9	6	9				

IFNY ELISPOT on PBMC from Rhesus monkeys immunized with one injection of 10^{10} vp/dose of Adenovectors expressing the different HCV NS cassettes. Data are expressed as SFC/ 10^6 PBMC.

FIG. 16A

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	MRK Ad5-NSmut 10 ¹⁰ vp/dose					
Pep pools	S201	075Q	137Q			
F (NS3p)	928	69	254			
G (NS3h)	317	436	98			
H (NS4)	56	101	45			
I (NS5a)	1530	1100	413			
L (NS5b)	149	23	92			
M (NS5b)	398	32	80			
DMSO	29	6	29			

	MRK Ad6-I	NSOPTmut 1	10 ¹⁰	vp/dose
Pep pools	98D209	106Q		113Q
F (NS3p)	3110	263		404
G (NS3h)	2115	642		1008
H (NS4)	373	72		19
I (NS5a)	103	37		347
L (NS5b)	149	22		10
M (NSSb)	314	428	-	19
DMSO	0	1		3

IFNy ELISPOT on PBMC from Rhesus monkeys immunized with one injection of 10^{10} vp/dose of Adenovectors expressing the different HCV NS cassettes. Data are expressed as SFC/ 10^6 PBMC.

FIG. 16B

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	Ad5-NS 10 ¹¹ vp/dose						
Pep pools	99C008	97N104	97X008	99C026			
F (NS3p)	28	1026	579	889			
G (NS3h)	1279	188	103	2453			
H (NS4)	18	39	138	109			
I (NS5a)	131	1068	172	141			
L (NS5b)	78	144	103	32			
M (NS5b)	24	68	47	84			
DMSO	3	16	1	19			

	MR	KAd6-NSmu	it 10 ¹¹ vp/	dose
Pep pools	98C047	97C055	93G	97X014
F (NS3p)	477	25	93	1022
G (NS3h)	959	398	81	1513
H (NS4)	36	14	99	53
I (NS5a)	171	45	1237	98
L (NS5b)	18	32	23	51
M (NS5b)	88	4	13	40
DMSO	8	3	1	5

IFN γ ELISPOT on PBMC from Rhesus monkeys immunized with two injections of 10^{11} vp/dose of Adenovectors expressing the different HCV NS cassettes. Data are expressed as SFC/10⁶ PBMC.

FIG. 16C

79/92

	MRI	KAd5-NSm	ut 10 ¹¹ vp/	dose
Pep pools	99C059	<i>99C060</i>	97X009	96069
F (NS3p)	28	81	1308	1618
G (NS3h)	2600	161	1008	123
H (NS4)	31	74	101	40
1 (NS5a)	181	99	69	96
L (NS5b)	24	31	40	20
M (NS5b)	11	58	38	164
DMSO	6	15	1	16
	<u></u>	L		l

IFNy ELISPOT on PBMC from Rhesus monkeys immunized with two injections of 10^{11} vp/dose of Adenovectors expressing the different HCV NS cassettes. Data are expressed as SFC/ 10^6 PBMC.

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	MRK Ad	5-NSmut 10	10 vp/dose
Pep pools	S201	075 <u>Q</u>	137 <u>Q</u>
pool F (NS3p)	881	1755	73
pool G (NS3h)	573		•
pool H (NS4)		3541	
pool I (NS5a)	2094		39
pool L (NS5b)			
pool M (NS5b)	756		
DMSO	319	117	44

	MRK Ad6-N	ISOPTmut 10	10 vp/dose
Pep pools	98D209	106Q	113 <u>Q</u>
pool F (NS3p)	5073	84	952
pool G (NS3h)	2376	160	3325
pool H (NS4)	700		
pool I (NS5a)			1106
pool L (NS5b)			
pool M (NS5b)	530	706	
DMSO	43	47	28

	MRK Ad	6-NSmut 10	¹⁰ vp/dose
Pep pools	S207	035Q	057Q
pool F (NS3p)	118	480	
pool G (NS3h)		196	
pool H (NS4)			
pool I (NS5a)	3340	933	
pool L (NS5b)	118		
pool M (NS5b)			
DMSO .	145	34	

IFN γ ICS on PBMC from Rhesus monkeys immunized with two injections at four weeks interval with 10^{10} vp/dose of Adenovectors expressing the different HCV NS cassettes. Data are expressed as number of positive IFN γ /CD3/CD8 per 10^6 lymphocytes.

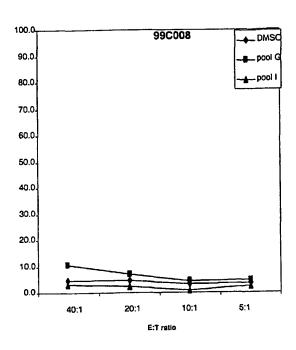
81/92

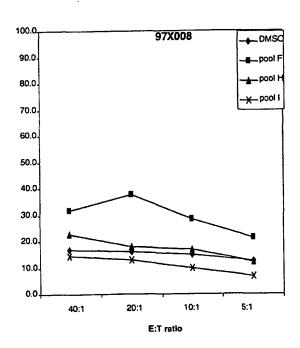
	^	d5-NS 10	11 vp/do	se
Pep pools	99C008	97N104	97X008	99C026
F (NS3p)		1703	1136	615
G (NS3h)	3153			2787
H (NS4)				
I (NS5a)		2233		
L (NS5b)				
M (NS5b)				
DMSO	125	98	130	0
	MRKA	\d6-NSm	ut 10 ¹¹ v	p/dose
Pep pools	98C047	97C055	93G	97X014
F (NS3p)	1024			948
G (NS3h)	3246	353		1074
H (NS4)			316	
I (NS5a)			6224	
L (NS5b)				
M (NS5b)				
DMSO	49	23	37	93
	MRKA	ıd5-NSmı	ut 10 ¹¹ vj	o/dose
Pep pools	99C059	99C060	97X009	96069
F (NS3p)	-		2266	5053
G (NS3h)	2434	316	1018	
H (NS4)	•			· ·
I (NS5a)				
L (NS5b)				
M (NS5b)				205
DMSO	13	110	119	15

IFNY ICS on PBMC from Rhesus monkeys immunized with two injections at four weeks interval with 10¹¹ vp/dose of Adenovectors expressing the different HCV NS cassettes. Data are expressed as number of positive IFNy/CD3/CD8 per 10⁶ lymphocytes.

FIG. 17B



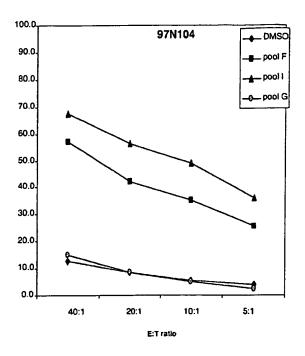


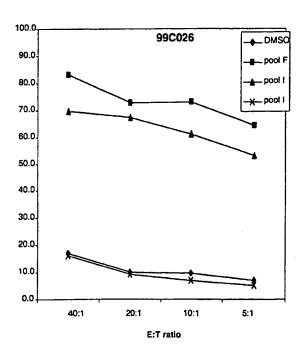


Bulk CTL assays on PBMC from Rhesus monkeys immunized with two injections of 10¹¹vp/dose of Ad5-NS.

FIG. 18A

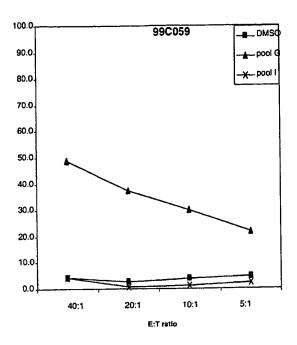


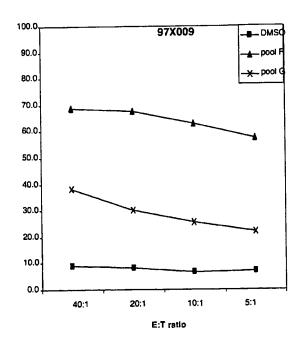




Bulk CTL assays on PBMC from Rhesus monkeys immunized with two injections of 10¹¹vp/dose of Ad5-NS.

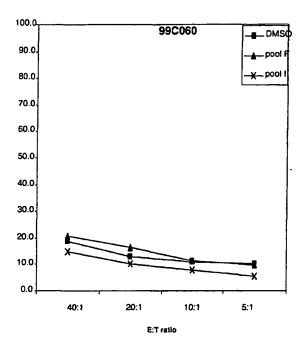
FIG. 18B

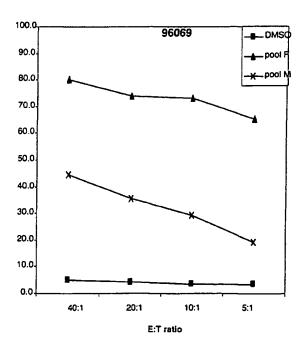




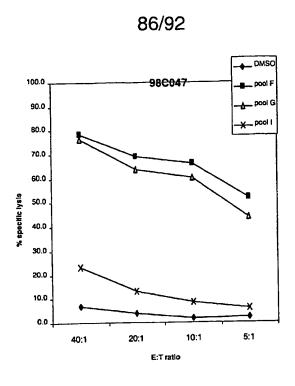
Bulk CTL assays on PBMC from Rhesus monkeys immunized with two injections of 1011vp/dose of MRKAd5-NSmut.

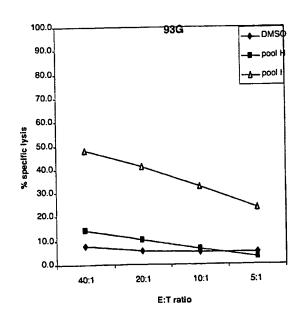






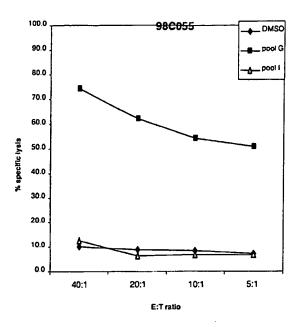
Bulk CTL assays on PBMC from Rhesus monkeys immunized with two injections of 1011vp/dose of MRKAd5-NSmut

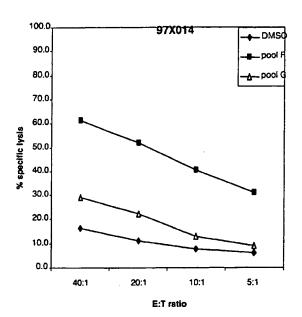




Bulk CTL assays on PBMC from Rhesus monkeys immunized with two injections of 1011vp/dose of MRKAd6-NSmut.







Bulk CTL assays on PBMC from Rhesus monkeys immunized with two injections of 10¹¹vp/dose of MRKAd6-NSmut.

FIG. 18F

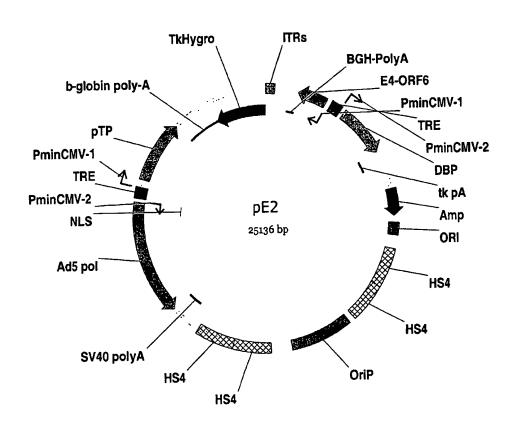


FIG. 19

1	GCCACCATGG	CCCCCATCAC	CGCCTACAGC	CAGCAGACCA	GGGGCCTGCT
51	GGGCTGCATC	ATCACCAGCC	TGACCGGACG	CGACAAGAAC	CAGGTGGAGG
101	GAGAGGTGCA	GGTGGTGAGC	ACCGCTACCC	AGAGCTTCCT	GGCCACCTGC
151	GTGAACGGCG	TGTGCTGGAC	CGTGTACCAC	GGAGCCGGAA	GCAAGACCCT
201	GGCCGGACCC	AAGGCCCTA	TCACCCAGAT	GTACACCAAT	GTGGATCAGG
251	ATCTGGTGGG	CTGGCAGGCC	CCTCCCGGAG	CCAGGAGCCT	GACACCCTGT
301	ACCTGTGGAA	GCAGCGACCT	GTACCTGGTG	ACACGCCACG	CCGATGTGAT
351	CCCCGTGAGG	CGCAGGGGCG	ATTCTCGCGG	AAGCCTGCTG	AGCCCTAGGC
401	CCGTGAGCTA	CCTGAAGGGC	AGCAGCGGAG	GACCCCTGCT	GTGTCCTTCT
451	GGCCATGCCG	TGGGCATTTT	TCGCGCTGCC	GTGTGTACCA	GGGGCGTGGC
501	CAAAGCCGTG	GATTTTGTGC	CCGTGGAAAG	CATGGAGACC	ACCATGCGCA
551	GCCCTGTGTT	CACCGACAAC	AGCTCTCCCC	CTGCCGTGCC	CCAATCATTC
601	CAGGTGGCTC	ACCTGCACGC	CCCTACCGGA	TCTGGCAAGA	GCACCAAGGT
651	GCCCGCTGCC	TACGCCGCTC	AGGGCTACAA	GGTGCTGGTG	CTGAACCCCA
701	GCGTGGCCGC	TACCCTGGGC	TTCGGCGCTT	ACATGAGCAA	GGCCCATGGC
751	ATCGACCCCA	ACATCCGCAC	AGGCGTGCGC	ACCATCACCA	CCGGAGCTCC
801	CGTGACCTAC	AGCACCTACG	GCAAGTTCCT	GGCCGATGGA	GGCTGCAGCG
851	GAGGAGCCTA	CGACATCATC	ATCTGCGACG	AGTGCCACAG	CACCGACAGC
901	ACCACCATCC	TGGGCATTGG	CACCGTGCTG	GATCAGGCCG	AAACAGCTGG
951	AGCCAGGCTG	GTGGTGCTGG	CCACAGCTAC	CCCTCCTGGC	AGCGTGACCG
1001	TGCCCCATCC	CAATATCGAG	GAGGTGGCCC	TGAGCAACAC	AGGCGAGATC
1051	CCCTTCTACG	GCAAGGCCAT	CCCCATCGAG	GCCATCCGCG	GAGGCAGGCA
1101	CCTGATCTTC	TGCCACAGCA	AGAAGAAGTG	CGACGAGCTG	GCTGCCAAGC
1151	TGAGCGGACT	GGGCATCAAC	GCCGTGGCCT	ACTACAGGGG	CCTGGACGTG
1201	TCAGTGATCC	CCACCATCGG	CGATGTGGTG	GTGGTGGCCA	CCGACGCCCT
1251	GATGACAGGC	TACACCGGAG	ACTTCGACAG	CGTGATCGAC	TGCAACACCT
1301	GCGTGACCCA	GACCGTGGAC	TTCAGCCTGG	ACCCCACCTT	CACCATCGAA
1351	ACCACCACCG	TGCCTCAGGA	TGCTGTGAGC	AGGAGCCAGA	GGCGCGGACG
1401	CACCGGAAGG	GGCAGGCGCG	GAATTTATCG	CTTTGTGACC	CCTGGCGAAA
1451	GGCCCTCTGG	CATGTTCGAC	AGCAGCGTGC	TGTGCGAGTG	CTACGACGCT
1501	GGCTGCGCTT	GGTACGAGCT	GACACCCGCT	GAAACCAGCG	TGCGCCTGCG
1551	CGCTTATCTG	AATACCCCTG	GCCTGCCCGT	GTGTCAGGAC	CACCTGGAGT

FIG. 20A

1601		CGTGTTCACA			
1651		AGCAGGCTGG			
1701		TGTGCTAGGG			
1751	TGTGGAAGTG	CCTGATCCGC	CTGAAGCCCA	CCCTGCACGG	CCCTACCCCT
1801	CTGCTGTACC	GCCTGGGAGC	CGTGCAGAAC	GAGGTGACCC	TGACCCACCC
1851	CATCACCAAG	TACATCATGG	CCTGCATGAG	CGCTGATCTG	GAAGTGGTGA
1901	CCAGCACCTG	GGTGCTGGTG	GGAGGCGTGC	TGGCCGCTCT	GGCTGCCTAC
1951	TGCCTGACCA	CCGGAAGCGT	GGTGATCGTG	GGACGCATCA	TCCTGAGCGG
2001	AAGGCCCGCT	ATCGTGCCCG	ATCGCGAGTT	CCTGTACCAG	GAGTTCGACG
2051	AGATGGAGGA	GTGTGCCAGC	CACCTGCCCT	ACATCGAGCA	GGGCATGCAG
2101	CTGGCCGAAC	AGTTCAAGCA	GAAGGCCCTG	GGCCTGCTGC	AGACAGCCAC
2151	CAAACAGGCC	GAAGCTGCCG	CTCCCGTGGT	GGAAAGCAAG	TGGAGGGCCC
2201	TGGAGACCTT	CTGGGCTAAG	CACATGTGGA	ACTTCATCTC	TGGCATCCAG
2251	TACCTGGCCG	GACTGAGCAC	CCTGCCTGGC	AACCCCGCTA	TCGCCAGCCT
2301	GATGGCCTTC	ACCGCTAGCA	TCACCTCTCC	CCTGACCACC	CAGAGCACCC
2351	TGCTGTTCAA	CATTCTGGGC	GGATGGGTGG	CCGCTCAGCT	GGCCCCTCCT
2401	TCAGCTGCTT	CTGCCTTTGT	GGGCGCTGGC	ATTGCCGGAG	CCGCTGTGGG
2451	CAGCATTGGC	CTGGGCAAAG	TGCTGGTGGA	TATTCTGGCT	GGCTATGGCG
2501	CTGGCGTGGC	CGGAGCCCTG	GTGGCCTTCA	AGGTGATGAG	CGGAGAGATG
2551	CCCAGCACCG	AGGACCTGGT	GAACCTGCTG	CCTGCCATTC	TGAGCCCTGG
2601	AGCCCTGGTG	GTGGGCGTGG	TGTGTGCTGC	CATTCTGAGG	CGCCATGTGG
2651	GACCCGGAGA	GGGCGCTGTG	CAGTGGATGA	ACCGCCTGAT	CGCCTTCGCC
2701	TCTCGCGGAA	ACCACGTGAG	CCCTACCCAC	TACGTGCCTG	AGAGCGACGC
2751	CGCTGCCAGG	GTGACCCAGA	TCCTGAGCAG	CCTGACCATC	ACCCAGCTGC
2801	TGAAGCGCCT	GCACCAGTGG	ATCAACGAGG	ACTGCAGCAC	ACCCTGCAGC
2851	GGAAGCTGGC	TGAGGGACGT	GTGGGACTGG	ATCTGCACCO	TGCTGACCGA
2901	CTTCAAGACO	TGGCTGCAGA	GCAAGCTGCT	GCCCCAACTC	CCTGGCGTGC
2951	CCTTCTTCTC	ATGCCAGCGC	GGATACAAGG	GCGTGTGGAG	GGGCGATGGC
3001	ATCATGCAGA	A CCACCTGTCC	: CTGCGGAGCC	CAGATCACAC	GCCACGTGAA
3051	GAACGGCAG	ATGCGCATCG	TGGGCCCTA	A GACCTGCAGO	AACACCTGGC
3101	ACGGCACCT	r CCCCATCAAC	GCCTACACCA	A CCGGACCCT	G CACACCCAGC
3151	CCTGCTCCC	A ACTACAGCAG	GGCCCTGTGC	G AGGGTGGCT	G CCGAGGAGTA

3201	CGTGGAGGTG	ACCAGGGTGG	GAGACTTCCA	CTACGTGACC	GGAATGACCA
3251	CCGACAACGT	GAAGTGTCCC	TGTCAGGTGC	CCGCTCCCGA	ATTTTTTACC
3301	GAAGTGGATG	GCGTGCGCCT	GCATCGCTAT	GCCCCTGCCT	GTAGGCCCCT
3351	GCTGCGCGAA	GAAGTGACCT	TCCAGGTGGG	CCTGAACCAG	TACCTGGTGG
3401	GCAGCCAGCT	GCCCTGCGAG	CCTGAGCCCG	ATGTGGCCGT	GCTGACCAGC
3451	ATGCTGACCG	ACCCCAGCCA	CATCACAGCC	GAAACCGCTA	AAAGGCGCCT
3501	GGCCAGGGGC	TCTCCTCCAA	GCCTGGCCTC	AAGCAGCGCT	AGCCAGCTGT
3551	CTGCTCCCAG	CCTGAAGGCC	ACCTGCACCA	CCCACCACGT	GAGCCCCGAC
3601	GCCGACCTGA	TCGAGGCCAA	CCTGCTGTGG	CGCCAGGAGA	TGGGCGGCAA
3651	CATCACCCGC	GTGGAGAGCG	AGAACAAGGT	GGTGGTGCTG	GACAGCTTCG
3701	ACCCCCTGCG	CGCCGAGGAG	GACGAGCGCG	AGGTGAGCGT	GCCCGCCGAG
3751	ATCCTGCGCA	AGAGCAAGAA	GTTCCCCGCT	GCCATGCCCA	TCTGGGCTAG
3801	ACCTGATTAC	AACCCTCCCC	TGCTGGAGAG	CTGGAAGGAC	CCTGATTACG
3851	TGCCTCCAGT	GGTGCATGGC	TGTCCTCTGC	CTCCCATTAA	AGCCCCTCCT
3901	ATTCCACCTC	CTAGGCGCAA	AAGGACCGTG	GTGCTGACAG	AAAGCAGCGT
3951	GAGCTCTGCT	CTGGCCGAAC	TGGCCACCAA	GACCTTTGGC	AGCAGCGAGA
4001	GCTCTGCCGT	GGACAGCGGA	ACAGCCACCG	CTCTGCCTGA	CCAGGCCAGC
4051	GACGACGGCG	ATAAGGGCAG	CGATGTGGAG	AGCTATAGCA	GCATGCCTCC
4101	CCTGGAAGGC	GAACCTGGCG	ATCCCGATCT	GAGCGATGGC	AGCTGGAGCA
4151	CCGTGAGCGA	AGAGGCCAGC	GAGGACGTGG	TGTGTTGCAG	CATGAGCTAC
4201	ACCTGGACAG	GCGCTCTGAT	CACACCCTGC	GCTGCCGAGG	AGAGCAAGCT
4251	GCCCATCAAC	GCCCTGAGCA	ACAGCCTGCT	GAGGCACCAC	AACATGGTGT
4301	ACGCCACCAC	CAGCAGGTCT	GCCGGACTGA	GGCAGAAGAA	GGTGACCTTC
4351	GACCGCCTGC	AGGTGCTGGA	CGACCACTAC	CGCGATGTGC	TGAAGGAGAT
4401	GAAGGCCAAG	GCCAGCACCG	TGAAGGCCAA	GCTGCTGAGC	GTGGAGGAGG
4451	CCTGCAAGCT	GACCCCCCC	CACAGCGCCA	AGAGCAAGTT	CGGCTACGGC
4501	GCCAAGGACG	TGCGCAACCT	GAGCAGCAAG	GCCGTGAACC	ACATCCACAG
4551	CGTGTGGAAG	GACCTGCTGG	AGGACACCGT	GACCCCCATC	GACACCACCA
4601	TCATGGCCAA	GAACGAGGTG	TTCTGCGTGC	AGCCCGAGAA	GGGCGGCCGC
4651	AAGCCCGCTC	GCCTGATCGT	GTTCCCCGAT	CTGGGCGTGC	GCGTGTGCGA
4701	GAAGATGGCC	CTGTACGACG	TGGTGAGCAC	CCTGCCTCAG	GTGGTGATGG
4751	GCTCAAGCTA	CGGCTTCCAG	TACAGCCCTG	GCCAGCGCGT	GGAGTTCCTG

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4801	GTGAACACCT	GGAAGAGCAA	GAAGAACCCC	ATGGGCTTCA	GCTACGACAC
4851	ACGCTGCTTC	GACAGCACCG	TGACCGAGAA	CGACATCCGC	GTGGAGGAGA
4901	GCATCTACCA	GTGCTGCGAC	CTGGCCCCTG	AGGCCAGGCA	GGCCATCAAG
4951	AGCCTGACCG	AGCGCCTGTA	CATCGGAGGC	CCTCTGACCA	ACAGCAAGGG
5001	ACAGAACTGC	GGATACAGGC	GCTGTAGGGC	CTCTGGCGTG	CTGACCACCA
5051	GCTGTGGCAA	CACCCTGACC	TGCTACCTGA	AGGCCAGCGC	TGCCTGTCGC
5101	GCTGCCAAGC	TGCAGGACTG	CACCATGCTG	GTGAACGCCG	CTGGCCTGGT
5151	GGTGATTTGT	GAAAGCGCTG	GCACCCAGGA	AGATGCTGCC	AGCCTGCGCG
5201	TGTTCACCGA	GGCCATGACC	AGGTACTCTG	CCCCTCCCGG	AGACCCCCCT
5251	CAGCCCGAAT	ACGACCTGGA	GCTGATCACC	AGCTGCTCAA	GCAACGTGAG
5301	CGTGGCTCAC	GACGCCAGCG	GAAAGCGCGT	GTACTACCTG	ACACGCGATC
5351	CCACCACCCC	TCTGGCTCGC	GCTGCCTGGG	AAACCGCTCG	CCATACACCC
5401	GTGAACAGCT	GGCTGGGCAA	CATCATCATG	TACGCCCCTA	CCCTGTGGGC
5451	TCGCATGATC	CTGATGACCC	ACTTCTTCAG	CATCCTGCTG	GCTCAGGAGC
5501	AGCTGGAGAA	GGCCCTGGAC	TGCCAGATTT	ACGGCGCTTG	CTACAGCATC
5551	GAGCCCCTGG	ACCTGCCCCA	AATCATCGAG	CGCCTGCACG	GCCTGTCTGC
5601	CTTCAGCCTG	CACAGCTACA	GCCCTGGCGA	AATTAATCGC	GTGGCCAGCT
5651	GTCTGCGCAA	ACTGGGCGTG	CCTCCTCTGC	GCGTGTGGAG	GCATAGGGCT
5701	AGGAGCGTGA	GGGCTAGGCT	GCTGAGCCAG	GGAGGCAGGG	CCGCTACCTG
5751	TGGAAAGTAC	CTGTTCAACT	GGGCCGTGAA	GACCAAGCTG	AAGCTGACCC
5801	CTATCCCTGC	CGCTAGCCAG	CTGGACCTGA	GCGGATGGTT	CGTGGCTGGC
5851	TACAGCGGAG	GCGACATCTA	CCACAGCCTG	TCTCGCGCTC	GCCCTCGCTG
5901	GTTCATGCTG	TGCCTGCTGC	TGCTGAGCGT	GGGCGTGGGC	ATCTACCTGC
5951	TGCCCAACCG	CTAAA			

FIG. 20D

IN THE PCT RECEIVING OFFICE OF THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s):

Merck & Co., Inc

PCT Serial No.:

To Be Assigned

Case No.: PCT ITR0015Y

US/RO

Filing date:

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Authorized Officer:

For:

HEPATITIS C VIRUS VACCINE

To Be Assigned

Assistant Commissioner of Patents

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NUCLEOTIDE AND/OR AMINO ACID SEQUENCE DISCLOSURE, PCT RULE 5.2

Sir:

As required under PCT Rule 5.2, Applicant respectfully encloses a paper (64 pages) and a computer readable form of the Sequence Listing for the above-identified PCT International Application, filed on even date herewith.

I hereby state that the content of the paper and computer readable forms of the Sequence Listing, submitted in accordance with WIPO and Standard ST.23 and under PCT Rule 13ter.1, respectively, are the same.

Respectfully submitted,

Bv

Sheldon O. Heber Reg. No. 38,179

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SEQUENCE LISTING

<110> Merck & Co. Inc., and Istituto Di Ricerche Di Biologia Molecolare P. Angeletti S.P.A. <120> HEPATITIS C VIRUS VACCINE <130> ITR0015Y <150> 60/363,774 <151> 2002-03-13 <150> 60/328,655 <151> 2001-10-11 <160> 17 <170> FastSEQ for Windows Version 4.0 <210> 1 <211> 1985 <212> PRT <213> Artificial Sequence <220> <223> Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide Met Ala Pro Ile Thr Ala Tyr Ser Gln Gln Thr Arg Gly Leu Leu Gly 10 Cys Ile Ile Thr Ser Leu Thr Gly Arg Asp Lys Asn Gln Val Glu Gly 25 30 20 Glu Val Gln Val Val Ser Thr Ala Thr Gln Ser Phe Leu Ala Thr Cys 40 Val Asn Gly Val Cys Trp Thr Val Tyr His Gly Ala Gly Ser Lys Thr 60 55 Leu Ala Gly Pro Lys Gly Pro Ile Thr Gln Met Tyr Thr Asn Val Asp 75 70 Gln Asp Leu Val Gly Trp Gln Ala Pro Pro Gly Ala Arg Ser Leu Thr 90 85 Pro Cys Thr Cys Gly Ser Ser Asp Leu Tyr Leu Val Thr Arg His Ala 110 105 100 Asp Val Ile Pro Val Arg Arg Gly Asp Ser Arg Gly Ser Leu Leu 120 125 115 Ser Pro Arg Pro Val Ser Tyr Leu Lys Gly Ser Ser Gly Gly Pro Leu 140 135 130 Leu Cys Pro Ser Gly His Ala Val Gly Ile Phe Arg Ala Ala Val Cys 155 150 Thr Arg Gly Val Ala Lys Ala Val Asp Phe Val Pro Val Glu Ser Met 175 170 Glu Thr Thr Met Arg Ser Pro Val Phe Thr Asp Asn Ser Ser Pro Pro 185 190 Ala Val Pro Gln Ser Phe Gln Val Ala His Leu His Ala Pro Thr Gly

200

195

205

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Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala Thr Leu Gly Phe Gly
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Ala Tyr Met Ser Lys Ala His Gly Ile Asp Pro Asn Ile Arg Thr Gly
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Val Arg Thr Ile Thr Thr Gly Ala Pro Val Thr Tyr Ser Thr Tyr Gly
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        260
Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly Ala Tyr Asp Ile Ile
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Ile Cys Asp Glu Cys His Ser Thr Asp Ser Thr Thr Ile Leu Gly Ile
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Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly Ala Arg Leu Val Val
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Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr Val Pro His Pro Asn
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Ile Glu Glu Val Ala Leu Ser Asn Thr Gly Glu Ile Pro Phe Tyr Gly
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Lys Ala Ile Pro Ile Glu Ala Ile Arg Gly Gly Arg His Leu Ile Phe
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Cys His Ser Lys Lys Cys Asp Glu Leu Ala Ala Lys Leu Ser Gly
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Ile Pro Thr Ile Gly Asp Val Val Val Val Ala Thr Asp Ala Leu Met
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Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile Asp Cys Asn Thr Cys
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			Arg 660					665					6/0		
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	690	Pro	Tyr			695					700				
705	Lys		Leu		710					715					120
Ala			Val	725	Glu				730					/35	
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			Gly 820					825					830		
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865			Val		870					875					880
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	930		Leu			935					940				
945			Ser		950					955					960
			Thr	965					970					9/5	
			Gly 980	Val	Pro			985		•			990		
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Thr	Thr			104	5				105	0				102	
Leu	Trp	Arg	Val 106		Ala	Glu	Glu	Tyr 106		. Glu	\Val	Thr	Arg 107	val 0	Gly

rap	Phe	Hic	Ther	Val	ጥከዮ	Glv	Met	Thr	ጥኮኮ	Δen	Acn	Val	T.VS	Cvs	Pro
		107	5				108	0				108	5		
Cys	Gln 109		Pro	Ala	Pro	Glu 109		Phe	Thr	Glu	Val 110		Gly	Val	Arg
Leu	His	Arg	Tyr	Ala	Pro	Ala	Cys	Arg	Pro	Leu	Leu	Arg	Glu	Glu	Val
110					1110					111					1120
Thr	Phe	Gln	Val	Gly 112		Asn	Gln	Tyr	Leu 1130		Gly	Ser	Gln	Leu 1139	
Cys	Glu	Pro	Glu 1140		Asp	Val	Ala	Val 1145		Thr	Ser	Met	Leu 1150		Asp
Pro	Ser		Ile		Ala	Glu		Ala		Arg	Arg		Ala		Gly
_		115		.		C	1160		31 -	C	~ 1	1169		21.	D=0
	Pro 1170)				1175	5				1180)			
	Leu	Lys	Ala	Thr			Thr	His	His			Pro	Asp	Ala	
118					1190		_	_		1199				_	1200
	Ile			1209	5				1210	כ				1215	5
Thr	Arg	Val	Glu 1220		Glu	Asn	Lys	Val 1225		Val	Leu	Asp	Ser 1230		Asp
Pro	Leu	Arg	Ala	Glu	Glu	Asp	Glu	Arg	Glu	Val	Ser	Val	Pro	Ala	Glu
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Ile	Leu 1250		Lys	Ser	Lys	Lys 1255		Pro	Ala	Ala	Met 1260		Ile	Trp	Ala
Δτα	Pro		ጥኒታ	λen	Pro			T.en	Glu	Ser			Asp	Pro	Asp
126		rap	1 y L	A311	1270		Dou	204	014	1275		_,,			1280
	Val	Pro	Pro	Val			Glv	Cvs	Pro			Pro	Tle	Lvs	
131	Vai	110	110	1289			017	-	1290					1295	
Pro	Pro	Ile		Pro		Arg	Arg		Arg		Val	Val		Thr	
			1300					1305			m\	T	1310		01
Ser	Ser	Val	Ser	Ser	Ala	Leu	Ala	GIU	ьeu	Ala	THE	гĀ2	Thr	Pne	GIA
		1315	5				1320)				1325	i		
	Ser Ser	1315	5				1320)				1325	i		
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Ser	Ser	1315 Glu)	Ser	Ser	Ala Asp	Val 1335 Gly	1320 Asp) Ser	Gly	Thr Ser	Ala 1340 Asp	1325 Thr	Ala	Leu	Pro Tyr
Ser Asp	Ser 1330 Gln	1315 Glu) Ala	Ser Ser	Ser Asp	Ala Asp 1350	Val 1335 Gly	132(Asp Asp	Ser Lys	Gly Gly	Thr Ser 1355	Ala 1340 Asp	1325 Thr Val	Ala Glu	Leu Ser	Pro Tyr 1360
Ser Asp	Ser 1330 Gln	1315 Glu) Ala	Ser Ser	Ser Asp Pro	Ala Asp 1350 Leu	Val 1335 Gly	132(Asp Asp	Ser Lys	Gly Gly Pro	Thr Ser 1355 Gly	Ala 1340 Asp	1325 Thr Val	Ala Glu	Leu Ser Leu	Pro Tyr 1360 Ser
Ser Asp 134 Ser	Ser 1330 Gln Ser	1315 Glu) Ala Met	Ser Ser Pro	Ser Asp Pro	Ala Asp 1350 Leu	Val 1335 Gly Olu	1320 Asp Asp Gly	Ser Lys Glu	Gly Gly Pro 1370	Thr Ser 1355 Gly	Ala 1340 Asp Asp	1325 Thr Val	Ala Glu Asp	Leu Ser Leu 1375	Pro Tyr 1360 Ser
Ser Asp 134 Ser	Ser 1330 Gln	1315 Glu) Ala Met	Ser Ser Pro	Ser Asp Pro 1365 Ser	Ala Asp 1350 Leu	Val 1335 Gly Olu	1320 Asp Asp Gly	Ser Lys Glu	Gly Gly Pro 1370 Glu	Thr Ser 1355 Gly	Ala 1340 Asp Asp	1325 Thr Val	Ala Glu Asp Asp	Leu Ser Leu 1375 Val	Pro Tyr 1360 Ser
Ser Asp 134: Ser Asp	Ser 1330 Gln Ser Gly	1315 Glu Ala Met	Ser Ser Pro Trp 1380	Ser Asp Pro 1365 Ser	Ala Asp 1350 Leu Thr	Val 1335 Gly Glu Val	1320 Asp Asp Gly Ser	Ser Lys Glu Glu 1385	Gly Gly Pro 1370 Glu	Thr Ser 1355 Gly) Ala	Ala 1340 Asp Asp	1325 Thr Val Pro	Ala Glu Asp Asp 1390	Leu Ser Leu 1375 Val	Pro Tyr 1360 Ser Val
Ser Asp 134: Ser Asp	Ser 1330 Gln Ser	1315 Glu Ala Met Ser	Ser Ser Pro Trp 1380	Ser Asp Pro 1365 Ser	Ala Asp 1350 Leu Thr	Val 1335 Gly Glu Val	1320 Asp Asp Gly Ser	Ser Lys Glu Glu 1385 Thr	Gly Gly Pro 1370 Glu	Thr Ser 1355 Gly) Ala	Ala 1340 Asp Asp	1325 Thr Val Pro Glu	Ala Glu Asp Asp 1390 Thr	Leu Ser Leu 1375 Val	Pro Tyr 1360 Ser Val
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Leu Ile Val Phe Pro Asp Leu Gly Val Arg Val Cys Glu Lys Met Ala	
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## (19) World Intellectual Property Organization International Bureau



## - 1 SOOTA BIRLOTO I BIRLOTO BELLE BIRLOTO BELLE BIRLOTO BELLE BIRLOTO BIRLOTO BIRLOTO BIRLOTO BIRLOTO BIRLOTO

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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.
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#### Published:

- with international search report
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments
- (88) Date of publication of the international search report:
  30 October 2003

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

31588

(54) Title: HEPATITIS C VIRUS VACCINE

(57) Abstract: The present invention features Ad6 vectors and a nucleic acid encoding a Met-NS3-NS4A-NS4B-NS5A-NS5B polypeptide containing an inactive NS5B RNA-dependent RNA polymerase region. The nucleic acid is particularly useful as a component of an adenovector or DNA plasmid vaccine providing a broad range of antigens for generating an HCV specific cell mediated immune (CMI) response against HCV.

## INTERNATIONAL SEARCH REPORT

International application No.
PCT/US02/32512

A. CLAS	SSIFICATION OF SUBJECT MATTER		[*			
IPC(7)	: C12N 15/40, 15/51, 15/85, 15/86, 15/861; A6	1K 48/00				
US CL	US CL : 514/44; 424/93.2; 435/320.1, 455, 456					
	According to International Patent Classification (IPC) or to both national classification and IPC					
	DS SEARCHED					
Minimum do	cumentation searched (classification system followed	by classification symbols)				
U.S. : 5	14/44; 424/93.2; 435/320.1, 455, 456		}			
Documentati	Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched					
2000000						
	ta base consulted during the international search (nar	ne of data base and, where practicable, s	earch terms useu)			
Please See C	ontinuation Sheet					
C. DOC	UMENTS CONSIDERED TO BE RELEVANT					
Category *	Citation of document, with indication, where ap	propriate, of the relevant passages	Relevant to claim No.			
X	US 6,127,116 A (RICE et al.) 03 October 2000 (03	.10.2000), column 45, lines 18-57.	1, 2			
A	WO 01/30812 A2 (CHIRON CORPORATION) 03	May 2001 (03.05.2001).	1-54			
			1.54			
A	WO 97/47358 A1 (MERCK & CO., INC.) 18 Dece	mber 1997 (18.12.1997).	1-54			
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Further	r documents are listed in the continuation of Box C.	See patent family annex.				
* 5	pecial categories of cited documents:	"T" later document published after the inte date and not in conflict with the applie	emational filing date or priority			
"A" documen	t defining the general state of the art which is not considered to be	principle or theory underlying the inve	ention			
of partice	ilar relevance	"X" document of particular relevance; the	claimed invention cannot be			
"E" carlier a	pplication or patent published on or after the international filing date	considered novel or cannot be conside	red to involve an inventive step			
	t which may throw doubts on priority claim(s) or which is cited to	when the document is taken alone				
"L" document establish	the publication date of another citation or other special reason (as	"Y" document of particular relevance; the				
specified		considered to involve an inventive ste combined with one or more other such	documents, such combination			
"O" documen	t referring to an oral disclosure, use, exhibition or other means	being obvious to a person skilled in th	o azt			
	t published prior to the international filing date but later than the	"&" document member of the same patent	family			
priority	inte claimed					
Date of the	actual completion of the international search	Date of mailing of the international sea	rch report			
Date of the	sound somptoness of the management of the	0.2 SEP 2003				
	3 (09.07.2003)		<del></del>			
	nailing address of the ISA/US	Think & O.S.	to for			
	il Stop PCT, Attn: ISA/US mmissioner for Patents	Scott D. Priebe				
P.0	D. Box 1450	Telephone No. (703) 308-0196	•			
	exandrin, Virginia 22313-1450	1010photho 110. (100) 500 0250				
Hacsimile N	n. <i>(</i> 703)305-3230	i				

Facsimile No. (703)305-3230
Form PCT/ISA/210 (second sheet) (July 1998)

### INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/32512

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)		
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:		
Claim Nos.: because they relate to subject matter not required to be searched by this Authority, namely:		
Claim Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:		
Claim Nos.:  because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).		
Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)		
This International Searching Authority found multiple inventions in this international application, as follows: Please See Continuation Sheet		
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.		
<ol> <li>As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.</li> </ol>		
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:		
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report		
is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-54		
Remark on Protest  The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.		

Form PCT/ISA/210 (continuation of first sheet(1)) (July 1998)

	PCT/US02/32512	
INTERNATIONAL SEARCH REPORT		
INTERNATIONAL SEARCH REPORT		
BOX II. OBSERVATIONS WHERE UNITY OF INVENTION IS LA This application contains the following inventions or groups of inventions which inventive concept under PCT Rule 13.1. In order for all inventions to be search paid.	are not so linked as to form a single general	
-		
Group I, claim(s) 1-54, drawn to a nucleic acid encoding a HCV polyprotein.		
1 -	1 . 10 . 1	
Group II, claim(s) 55-59, drawn to a chimeric adenovirus vector comprising sequence derived from human adenovirus serotypes 5 and		
6.		
T 177 to establish to a ringle general inventi-	ve concept under PCT Rule 13.1 because, under PCT	
The inventions listed as Groups I and II do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons:		
The technical footing of invention Lie a miciele Scid encoding a DOIVD	folem derived from an ric v poryprotein, whereas are	
It is a chimeric adenoviral vector comprising a ne	terologous seguence. These two leatures are not	
related. Invention I does not require vector of invention II, nor does is the vector	or of invention II required to contain the	
polymicleotides of invention I.		
<b>F</b> ,		
Continuation of B. FIELDS SEARCHED Item 3:		
A COLUMN THE CARLES CARLES RIOSIS SCISEARCH, USPT. PGPB. DEF	RWENT, GENBANK, GENESEQ	
search terms: HCV, hepatitis C virus, vaccine, NS5B, NS5B near inactiv? or n	on-functional, SEQ ID NO: 1, SEQ ID NO: 2	
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